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(54) Title: RNA INTERFERENCE MEDIATED INHIBITION OF SEVERE ACUTE RESPIRATORY SYNDROME (SARS) VIRUS GENE EXPRESSION USING SHORT INTERFERING NUCLEIC ACID (siNA)

(57) Abstract: The present invention comprises compounds, compositions, and methods useful for modulating the expression of genes associated with respiratory and pulmonary disease, such as severe acute respiratory syndrome (SARS) virus genes, using short interfering nucleic acid (siNA) molecules. This invention also comprises compounds, compositions, and methods useful for modulating the expression and activity of SARS virus genes, or other genes involved in pathways of SARS virus gene expression and/or activity by RNA interference (RNAi) using small nucleic acid molecules. In particular, the instant invention features small nucleic acid molecules, such as short interfering nucleic acid (siNA), short interfering RNA (siRNA), double-stranded RNA (dsRNA), micro-RNA (miRNA), and short hairpin RNA (shRNA) molecules and methods used to modulate the expression of SARS virus RNA.



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RNA INTERFERENCE MEDIATED INHIBITION OF SEVERE ACUTE RESPIRATORY SYNDROME (SARS) VIRUS GENE EXPRESSION USING SHORT INTERFERING NUCLEIC ACID (siNA)

This application claims the benefit of U.S. Provisional Application No. 60/462,874, filed April 15, 2003, and is a continuation-in-part of U.S. Patent Application No. 10/757,803, filed January 14, 2004, which is a continuation-in-part of U.S. Patent Application No. 10/720,448, filed November 24, 2003, which is a continuation-in-part of U.S. Patent Application No. 10/693,059, filed October 23, 2003, which is a continuation-in-part of U.S. Patent Application No. 10/444,853, filed May 23, 2003. This application is also a continuation-in-part of US Patent Application No. 10/427,160, filed April 30, 2003.

Reference is made to International Patent Application No. PCT/US03/05346, filed February 20, 2003, and International Patent Application No. PCT/US03/05028, filed February 20, 2003, both of which claim the benefit of U.S. Provisional Application No. 60/358,580 filed February 20, 2002, U.S. Provisional Application No. 60/363,124 filed March 11, 2002, U.S. Provisional Application No. 60/386,782 filed June 6, 2002, U.S. Provisional Application No. 60/406,784 filed August 29, 2002, U.S. Provisional Application No. 60/408,378 filed September 5, 2002, U.S. Provisional Application No. 60/409,293 filed September 9, 2002, and U.S. Provisional Application No. 60/440,129 filed January 15, 2003. Reference is also made to International Patent Application No. PCT/US02/15876 filed May 17, 2002.

All the listed applications are hereby incorporated by reference herein in their entireties, including the drawings.

Field Of The Invention

The present invention concerns compounds, compositions, and methods for the study, diagnosis, and treatment of diseases and conditions that respond to the modulation of severe acute respiratory syndrome (SARS) associated cornavirus (SARS virus) gene expression and/or activity. The present invention also concerns compounds, compositions, and methods relating to conditions and diseases that respond to the modulation of expression and/or activity of genes involved in SARS virus pathways of

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gene expression, including cellular genes that are involved in SARS virus infection. Specifically, the invention comprises small nucleic acid molecules, such as short interfering nucleic acid (siNA), short interfering RNA (siRNA), double-stranded RNA (dsRNA), micro-RNA (miRNA), and short hairpin RNA (shRNA) molecules capable of mediating RNA interference (RNAi) against severe acute respiratory syndrome (SARS) associated cornavirus gene expression.

Background Of The Invention

The following is a discussion of relevant art pertaining to RNAi. The discussion is provided only for understanding of the invention that follows. The summary is not an admission that any of the work described below is prior art to the claimed invention.

RNA interference refers to the process of sequence-specific post-transcriptional gene silencing in animals mediated by short interfering RNAs (siRNAs) (Zamore et al., 2000, Cell, 101, 25-33; Fire et al., 1998, Nature, 391, 806; Hamilton et al., 1999, Science, 286, 950-951; Lin et al., 1999, Nature, 402, 128-129; Sharp, 1999, Genes & Dev., 13:139-141; and Strauss, 1999, Science, 286, 886). The corresponding process in plants (Heifetz et al., International PCT Publication No. WO 99/61631) is commonly referred to as post-transcriptional gene silencing or RNA silencing and is also referred to as quelling in fungi. The process of post-transcriptional gene silencing is thought to be an evolutionarily-conserved cellular defense mechanism used to prevent the expression of foreign genes and is commonly shared by diverse flora and phyla (Fire et al., 1999, Trends Genet., 15, 358). Such protection from foreign gene expression may have evolved in response to the production of double-stranded RNAs (dsRNAs) derived from viral infection or from the random integration of transposon elements into a host genome via a cellular response that specifically destroys homologous single-stranded RNA or viral genomic RNA. The presence of dsRNA in cells triggers the RNAi response through a mechanism that has yet to be fully characterized. This mechanism appears to be different from other known mechanisms involving double stranded RNA-specific ribonucleases, such as the interferon response that results from dsRNA-mediated activation of protein kinase PKR and 2',5'-oligoadenylate synthetase resulting in nonspecific cleavage of mRNA by ribonuclease L (see for example US Patent Nos.

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6,107,094; 5,898,031; Clemens et al., 1997, J. Interferon & Cytokine Res., 17, 503-524; Adah et al., 2001, Curr. Med. Chem., 8, 1189).

The presence of long dsRNAs in cells stimulates the activity of a ribonuclease III enzyme referred to as dicer (Bass, 2000, Cell, 101, 235; Zamore et al., 2000, Cell, 101, 25-33; Hammond et al., 2000, Nature, 404, 293). Dicer is involved in the processing of the dsRNA into short pieces of dsRNA known as short interfering RNAs (siRNAs) (Zamore et al., 2000, Cell, 101, 25-33; Bass, 2000, Cell, 101, 235; Berstein et al., 2001, Nature, 409, 363). Short interfering RNAs derived from dicer activity are typically about 21 to about 23 nucleotides in length and comprise about 19 base pair duplexes (Zamore et al., 2000, Cell, 101, 25-33; Elbashir et al., 2001, Genes Dev., 15, 188). Dicer has also been implicated in the excision of 21 and 22-nucleotide small temporal RNAs (stRNAs) from precursor RNA of conserved structure that are implicated in translational control (Hutvagner et al., 2001, Science, 293, 834). The RNAi response also features an endonuclease complex, commonly referred to as an RNA-induced silencing complex (RISC), which mediates cleavage of single-stranded RNA having sequence complementary to the antisense strand of the siRNA duplex. Cleavage of the target RNA takes place in the middle of the region complementary to the antisense strand of the siRNA duplex (Elbashir et al., 2001, Genes Dev., 15, 188).

RNAi has been studied in a variety of systems. Fire et al., 1998, Nature, 391, 806, 20 were the first to observe RNAi in C. elegans. Bahramian and Zarbl, 1999, Molecular and Cellular Biology, 19, 274-283 and Wianny and Goetz, 1999, Nature Cell Biol., 2, 70, describe RNAi mediated by dsRNA in mammalian systems. Hammond et al., 2000, Nature, 404, 293, describe RNAi in Drosophila cells transfected with dsRNA. Elbashir et al., 2001, Nature, 411, 494 and Tuschl et al., International PCT Publication No. WO 25 01/75164, describe RNAi induced by introduction of duplexes of synthetic 21-nucleotide RNAs in cultured mammalian cells including human embryonic kidney and HeLa cells. Recent work in Drosophila embryonic lysates (Elbashir et al., 2001, EMBO J., 20, 6877 and Tuschl et al., International PCT Publication No. WO 01/75164) has revealed certain requirements for siRNA length, structure, chemical composition, and sequence that are 30 essential to mediate efficient RNAi activity. These studies have shown that 21nucleotide siRNA duplexes are most active when containing 3'-terminal dinucleotide

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overhangs. Furthermore, complete substitution of one or both siRNA strands with 2'-deoxy (2'-H) or 2'-O-methyl nucleotides abolishes RNAi activity, whereas substitution of the 3'-terminal siRNA overhang nucleotides with 2'-deoxy nucleotides (2'-H) was shown to be tolerated. Single mismatch sequences in the center of the siRNA duplex were also shown to abolish RNAi activity. In addition, these studies also indicate that the position of the cleavage site in the target RNA is defined by the 5'-end of the siRNA guide sequence rather than the 3'-end of the guide sequence (Elbashir *et al.*, 2001, *EMBO J.*, 20, 6877). Other studies have indicated that a 5'-phosphate on the target-complementary strand of a siRNA duplex is required for siRNA activity and that ATP is utilized to maintain the 5'-phosphate moiety on the siRNA (Nykanen *et al.*, 2001, *Cell*, 107, 309).

Studies have shown that replacing the 3'-terminal nucleotide overhanging segments of 21-mer siRNA duplex having two-nucleotide 3'-overhangs deoxyribonucleotides does not have an adverse effect on RNAi activity. Replacing up to four nucleotides on each end of the siRNA with deoxyribonucleotides has been reported to be well tolerated, whereas complete substitution with deoxyribonucleotides results in no RNAi activity (Elbashir et al., 2001, EMBO J., 20, 6877 and Tuschl et al., International PCT Publication No. WO 01/75164). In addition, Elbashir et al., supra, also report that substitution of siRNA with 2'-O-methyl nucleotides completely abolishes RNAi activity. Li et al., International PCT Publication No. WO 00/44914, and Beach et al., International PCT Publication No. WO 01/68836 preliminarily suggest that siRNA may include modifications to either the phosphate-sugar backbone or the nucleoside to include at least one of a nitrogen or sulfur heteroatom, however, neither application postulates to what extent such modifications would be tolerated in siRNA molecules, nor provides any further guidance or examples of such modified siRNA. Kreutzer et al., Canadian Patent Application No. 2,359,180, also describe certain chemical modifications for use in dsRNA constructs in order to counteract activation of double-stranded RNAdependent protein kinase PKR, specifically 2'-amino or 2'-O-methyl nucleotides, and nucleotides containing a 2'-O or 4'-C methylene bridge. However, Kreutzer et al. similarly fails to provide examples or guidance as to what extent these modifications would be tolerated in dsRNA molecules.

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Parrish et al., 2000, Molecular Cell, 6, 1077-1087, tested certain chemical modifications targeting the unc-22 gene in C. elegans using long (>25 nt) siRNA transcripts. The authors describe the introduction of thiophosphate residues into these siRNA transcripts by incorporating thiophosphate nucleotide analogs with T7 and T3 RNA polymerase and observed that RNAs with two phosphorothioate modified bases also had substantial decreases in effectiveness as RNAi. Further, Parrish et al. reported that phosphorothicate modification of more than two residues greatly destabilized the RNAs in vitro such that interference activities could not be assayed. Id. at 1081. The authors also tested certain modifications at the 2'-position of the nucleotide sugar in the long siRNA transcripts and found that substituting deoxynucleotides for ribonucleotides produced a substantial decrease in interference activity, especially in the case of Uridine to Thymidine and/or Cytidine to deoxy-Cytidine substitutions. Id. In addition, the authors tested certain base modifications, including substituting, in sense and antisense strands of the siRNA, 4-thiouracil, 5-bromouracil, 5-iodouracil, and 3-(aminoallyl)uracil for uracil, and inosine for guanosine. Whereas 4-thiouracil and 5-bromouracil substitution appeared to be tolerated, Parrish reported that inosine produced a substantial decrease in interference activity when incorporated in either strand. Parrish also reported that incorporation of 5-iodouracil and 3-(aminoallyl)uracil in the antisense strand resulted in a substantial decrease in RNAi activity as well.

The use of longer dsRNA has been described. For example, Beach *et al.*, International PCT Publication No. WO 01/68836, describes specific methods for attenuating gene expression using endogenously-derived dsRNA. Tuschl *et al.*, International PCT Publication No. WO 01/75164, describe a *Drosophila in vitro* RNAi system and the use of specific siRNA molecules for certain functional genomic and certain therapeutic applications; although Tuschl, 2001, *Chem. Biochem.*, 2, 239-245, doubts that RNAi can be used to cure genetic diseases or viral infection due to the danger of activating interferon response. Li *et al.*, International PCT Publication No. WO 00/44914, describe the use of specific long (141 bp-488 bp) enzymatically synthesized or vector expressed dsRNAs for attenuating the expression of certain target genes. Zernicka-Goetz *et al.*, International PCT Publication No. WO 01/36646, describe certain methods for inhibiting the expression of particular genes in mammalian cells using certain long (550 bp-714 bp), enzymatically synthesized or vector expressed dsRNA

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molecules. Fire et al., International PCT Publication No. WO 99/32619, describe particular methods for introducing certain long dsRNA molecules into cells for use in inhibiting gene expression in nematodes. Plaetinck et al., International PCT Publication No. WO 00/01846, describe certain methods for identifying specific genes responsible for conferring a particular phenotype in a cell using specific long dsRNA molecules. Mello et al., International PCT Publication No. WO 01/29058, describe the identification of specific genes involved in dsRNA-mediated RNAi. Pachuck et al., International PCT Publication No. WO 00/63364, describe certain long (at least 200 nucleotide) dsRNA constructs. Deschamps Depaillette et al., International PCT Publication No. WO 99/07409, describe specific compositions consisting of particular dsRNA molecules combined with certain anti-viral agents. Waterhouse et al., International PCT Publication No. 99/53050 and 1998, PNAS, 95, 13959-13964, describe certain methods for decreasing the phenotypic expression of a nucleic acid in plant cells using certain dsRNAs. Driscoll et al., International PCT Publication No. WO 01/49844, describe specific DNA expression constructs for use in facilitating gene silencing in targeted organisms.

Others have reported on various RNAi and gene-silencing systems. For example, Parrish et al., 2000, Molecular Cell, 6, 1077-1087, describe specific chemically-modified dsRNA constructs targeting the unc-22 gene of C. elegans. Grossniklaus, International PCT Publication No. WO 01/38551, describes certain methods for regulating polycomb gene expression in plants using certain dsRNAs. Churikov et al., International PCT Publication No. WO 01/42443, describe certain methods for modifying genetic characteristics of an organism using certain dsRNAs. Cogoni et al., International PCT Publication No. WO 01/53475, describe certain methods for isolating a Neurospora silencing gene and uses thereof. Reed et al., International PCT Publication No. WO 01/68836, describe certain methods for gene silencing in plants. Honer et al., International PCT Publication No. WO 01/70944, describe certain methods of drug screening using transgenic nematodes as Parkinson's Disease models using certain dsRNAs. Deak et al., International PCT Publication No. WO 01/72774, describe certain Drosophila-derived gene products that may be related to RNAi in Drosophila. Arndt et al., International PCT Publication No. WO 01/92513 describe certain methods for mediating gene suppression by using factors that enhance RNAi. Tuschl et al.,

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International PCT Publication No. WO 02/44321, describe certain synthetic siRNA Pachuk et al., International PCT Publication No. WO 00/63364, and Satishchandran et al., International PCT Publication No. WO 01/04313, describe certain methods and compositions for inhibiting the function of certain polynucleotide sequences using certain long (over 250 bp), vector expressed dsRNAs. Echeverri et al., International PCT Publication No. WO 02/38805, describe certain C. elegans genes identified via RNAi. Kreutzer et al. International PCT Publications Nos. WO 02/055692, WO 02/055693, and EP 1144623 B1 describes certain methods for inhibiting gene expression using dsRNA. Graham et al., International PCT Publications Nos. WO 99/49029 and WO 01/70949, and AU 4037501 describe certain vector expressed siRNA molecules. Fire et al., US 6,506,559, describe certain methods for inhibiting gene expression in vitro using certain long dsRNA (299 bp-1033 bp) constructs that mediate RNAi. Martinez et al., 2002, Cell, 110, 563-574, describe certain single stranded siRNA constructs, including certain 5'-phosphorylated single stranded siRNAs that mediate RNA interference in Hela cells. Harborth et al., 2003, Antisense & Nucleic Acid Drug Development, 13, 83-105, describe certain chemically and structurally modified siRNA molecules. Chiu and Rana, 2003, RNA, 9, 1034-1048, describe certain chemically and structurally modified siRNA molecules.

McCaffrey et al., 2002, Nature, 418, 38-39, describes the use of certain siRNA constructs targeting a chimeric SARS NS5B protein/luciferase transcript in mice.

Randall *et al.*, 2003, *PNAS USA*, 100, 235-240, describe certain siRNA constructs targeting SARS RNA in Huh7 hepatoma cell lines.

SUMMARY OF THE INVENTION

This invention comprises compounds, compositions, and methods useful for modulating the expression of genes associated with the development or maintenance of SARS virus infection, acute respiratory failure, viral pneumonia, and/or other disease states associated with SARS virus infection,, using short interfering nucleic acid (siNA) molecules. This invention also comprises compounds, compositions, and methods useful

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for modulating the expression and activity of severe acute respiratory syndrome (SARS) associated cornavirus or genes involved in severe acute respiratory syndrome (SARS) associated cornavirus gene expression and/or activity by RNA interference (RNAi) using small nucleic acid molecules. In particular, the instant invention features small nucleic acid molecules, such as short interfering nucleic acid (siNA), short interfering RNA (siRNA), double-stranded RNA (dsRNA), micro-RNA (miRNA), and short hairpin RNA (shRNA) molecules and methods used to modulate the expression of severe acute respiratory syndrome (SARS) associated cornavirus. For convenience, all forms of the small nucleic acid molecules of the invention (e.g., siRNA, dsRNA, micro-RNA, etc.) are referred to herein as "siNA," unless expressly stated otherwise.

A siNA of the invention can be unmodified or chemically-modified. A siNA of the instant invention can be chemically synthesized, expressed from a vector or enzymatically synthesized. The instant invention also features various chemically-modified synthetic short interfering nucleic acid (siNA) molecules capable of modulating repeat expansion gene expression or activity in cells by RNA interference (RNAi). The use of chemically-modified siNA improves various properties of native siNA molecules through increased resistance to nuclease degradation in vivo and/or through improved cellular uptake. Further, contrary to earlier published studies, siNA having multiple chemical modifications retains its RNAi activity. The siNA molecules of the instant invention are useful reagents and are useful in methods for a variety of therapeutic, diagnostic, target validation, genomic discovery, genetic engineering, and pharmacogenomic applications.

In one embodiment, the invention comprises one or more siNA molecules (and methods of using them) that independently or in combination modulate the expression of gene(s) encoding SARS virus. Specifically, the present invention comprises siNA molecules that modulate the expression of SARS proteins, for example, proteins encoded by SARS virus genome, such as Genbank Accession Nos. in Table I.

In one embodiment, the invention comprises one or more siNA molecules (and methods of using them) that independently or in combination modulate the expression of genes representing cellular targets for SARS virus infection, such as cellular receptors,

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cell surface molecules, cellular enzymes, cellular transcription factors, and/or cytokines, second messengers, and cellular accessory molecules.

Due to the high sequence variability of the SARS genome, selection of siNA molecules for broad therapeutic applications preferably involve the conserved regions of the SARS genome. In one embodiment, the present invention comprises siNA molecules that target the conserved regions of the SARS genome, such as the polymerase encoding region of the SARS virus genomic RNA. Therefore, siNA molecules of the invention are designed to target all the different isolates of SARS. siNA molecules designed to target conserved regions of various SARS isolates enable efficient inhibition of SARS replication in diverse patient populations and ensure the effectiveness of the siNA molecules against SARS quasi species that evolve due to mutations in the non-conserved regions of the SARS genome. Therefore, a single siNA molecule can be targeted against all isolates of SARS by designing the siNA molecule to interact with conserved nucleotide sequences of SARS (such conserved sequences are expected to be present in the RNA of all SARS isolates).

In one embodiment, a siNA molecule is designed to target the 3'-untranslated region and/or the shared leader sequence of genomic SARS RNA transcripts. Because SARS cornavirus mRNAs are nested with the genomic RNA and share common 3' region and polyA region, a single siNA targeting the 3'-end can target all transcripts plus the genomic RNA.

In one embodiment, a siNA molecule of the invention targets both the plus (genomic) strand RNA and minus strand RNA of the SARS virus. Because the SARS virus generates a minus strand RNA from plus strand genomic RNA, a double stranded siNA molecule targeting the plus strand will also target the minus strand, thus allowing a single double-stranded siNA to target both the plus (genomic) and the minus strand of the SARS virus. For example, a double stranded siNA molecule targeting the 3'-end of the SARS virus genomic strand will also target the 3'-end of the the minus strand, thus allowing a single double-stranded siNA to target both the plus and the minus strand of the SARS virus.

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In one embodiment, the invention comprises one or more siNA molecules (and methods of using them) that independently or in combination modulate the expression of gene(s) encoding SARS virus and/or cellular proteins associated with the maintenance or development of SARS virus infection and/or acute respiratory failure, viral pneumonia, such as genes encoding sequences comprising those sequences referred to by GenBank Accession Nos. shown in Table I, referred to herein generally as SARS. The description below of the various aspects and embodiments of the invention is provided with reference to exemplary severe acute respiratory syndrome (SARS) associated cornavirus genes, generally referred to herein as SARS. However, such reference is meant to be exemplary only and the various aspects and embodiments of the invention are also directed to other genes that express alternate SARS genes, such as mutant SARS genes, splice variants of SARS genes, and genes encoding different strains of SARS, as well as as cellular targets for SARS, such as those described herein. The various aspects and embodiments are also directed to other genes involved in SARS pathways, including genes that encode cellular proteins involved in the maintenance and/or development of SARS virus infection and/or acute respiratory failure or other genes that express other proteins associated with SARS virus infection, such as cellular proteins that are utilized in the SARS life-cycle. Such additional genes can be analyzed for target sites using the methods described herein for SARS. Thus, the inhibition and the effects of such inhibition of the other genes can be performed as described herein. In other words, the term "SARS" as it is defined herein below and recited in the described embodiments, is meant to encompass genes associated with the development or maintenance of SARS virus infection, such as genes which encode SARS polypeptides, including polypeptides of different strains of SARS, mutant SARS genes, and splice variants of SARS genes, as well as cellular genes involved in SARS pathways of gene expression, replication, and/or SARS activity. Also, the term "SARS" as it is defined herein and recited in the described embodiments, is meant to encompass SARS viral gene products and cellular gene products involved in SARS virus infection, such as those described herein. Thus, each of the embodiments described herein with reference to the term "SARS" are applicable to all of the virus, cellular and viral protein, peptide, polypeptide, and/or polynucleotide molecules covered by the term "SARS" as that term is defined herein.

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In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a severe acute respiratory syndrome virus (e.g., SARS) gene, wherein said siNA molecule comprises about 19 to about 23 base pairs. Preferably the number of based pairs in the siNA molecule is 18, 19, 20, 21, 22, 23, or 24.

In one embodiment, the invention features a siNA molecule that down-regulates expression of a SARS gene, for example, wherein the SARS gene comprises SARS encoding sequence. In one embodiment, the invention features a siNA molecule that down-regulates expression of a SARS gene, for example, wherein the SARS gene comprises SARS non-coding sequence or regulatory elements involved in SARS gene expression.

In one embodiment, the invention features a siNA molecule having RNAi activity against SARS RNA, wherein the siNA molecule comprises a sequence complementary to any RNA having SARS encoding sequence, such as those sequences having GenBank Accession Nos. shown in Table I. In another embodiment, the invention features a siNA molecule having RNAi activity against SARS RNA, wherein the siNA molecule comprises a sequence complementary to an RNA having other SARS encoding sequence, for example other mutant SARS genes not shown in Table I but known in the art to be associated with respiratory and/or pulmonary disease, SARS virus infection and/or acute respiratory failure, viral pneumonia, impeded respiration, respiratory distress syndrome, pulmonary hypertension, or pulmonary vasoconstriction. Chemical modifications as shown in Tables III and IV or otherwise described herein can be applied to any siNA construct of the invention. In another embodiment, a siNA molecule of the invention includes nucleotide sequence that can interact with nucleotide sequence of a SARS gene and thereby mediate silencing of SARS gene expression, for example, wherein the siNA mediates regulation of SARS gene expression by cellular processes that modulate the chromatin structure of the SARS gene and prevent transcription of the SARS gene.

In another embodiment, the invention features a siNA molecule comprising nucleotide sequence, for example, nucleotide sequence in the antisense region of the siNA molecule that is complementary to a nucleotide sequence or portion of sequence of

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a SARS gene. In another embodiment, the invention features a siNA molecule comprising a region, for example, the antisense region of the siNA construct, complementary to a sequence comprising a SARS gene sequence or a portion thereof.

In one embodiment, the antisense region of SARS siNA constructs can comprise a sequence complementary to sequence having any of SEQ ID NOs. 1-1651 or 3303-3318. In one embodiment, the antisense region can also comprise sequence having any of SEQ ID NOs. 1652-3302, 3319-3326, 3335-3342, 3351-3358, 3367-3374, 3376, 3378, 3380, 3383, 3385, 3387, 3389, or 3392. In another embodiment, the sense region of the SARS constructs can comprise sequence having any of SEQ ID NOs. 1-1651, 3303-3310, 3311-3318, 3327-3334, 3343-3350, 3359-3366, 3375, 3377, 3379, 3381, 3382, 3384, 3386, 3388, 3390, or 3391.

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In one embodiment, a siNA molecule of the invention comprises any of SEQ ID NOs. 1-3392. The sequences shown in SEQ ID NOs: 1-3392 are not limiting. A siNA molecule of the invention can comprise any contiguous SARS sequence (e.g., about 19 to about 25, or about 19, 20, 21, 22, 23, 24 or 25 contiguous SARS nucleotides).

In yet another embodiment, the invention features a siNA molecule comprising a sequence, for example, the antisense sequence of the siNA construct, complementary to a sequence or portion of sequence comprising sequence represented by GenBank Accession Nos. shown in **Table I**. Chemical modifications in **Tables III and IV** and described herein can be applied to any siNA construct of the invention. siNA molecules of the invention are unmodified or have up to all nucleotides modified with modifications according to Tables III and IV.

In one embodiment of the invention a siNA molecule comprises an antisense strand having about 19 to about 29 (e.g., 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30) nucleotides, wherein the antisense strand is complementary to a RNA sequence encoding a SARS protein, and wherein said siNA further comprises a sense strand having about 19 to about 29 (e.g., 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30) nucleotides, and wherein said sense strand and said antisense strand are distinct nucleotide sequences with at least about 19 complementary nucleotides.

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In another embodiment of the invention a siNA molecule of the invention comprises an antisense region having about 19 to about 29 (e.g., 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30) nucleotides, wherein the antisense region is complementary to a RNA sequence encoding a SARS protein, and wherein said siNA further comprises a sense region having about 19 to about 29 (e.g., 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or more) nucleotides, wherein said sense region and said antisense region comprise a linear molecule with at least about 19 complementary nucleotides.

In one embodiment of the invention a siNA molecule comprises an antisense strand comprising a nucleotide sequence that is complementary to a nucleotide sequence or a portion thereof encoding a SARS protein. The siNA further comprises a sense strand, wherein said sense strand comprises a nucleotide sequence of a SARS gene or a portion thereof.

In another embodiment, a siNA molecule comprises an antisense region comprising a nucleotide sequence that is complementary to a nucleotide sequence encoding a SARS protein or a portion thereof. The siNA molecule further comprises a sense region, wherein said sense region comprises a nucleotide sequence of a SARS gene or a portion thereof.

In one embodiment, a siNA molecule of the invention has RNAi activity that modulates expression of RNA encoded by a SARS gene. Because SARS genes can share some degree of sequence homology with each other, siNA molecules can be designed to target a class of SARS genes or alternately specific SARS genes by selecting sequences that are either shared among different SARS targets (e.g., different viral strains) or alternatively that are unique for a specific SARS target (e.g., a particular viral strain). Therefore, in one embodiment, the siNA molecule can be designed to target conserved regions of SARS RNA sequences having homology among several SARS genes so as to target several SARS genes (e.g., different SARS isoforms, splice variants, mutant genes etc.) with one siNA molecule. In another embodiment, the siNA molecule can be designed to target a sequence that is unique to a specific SARS RNA sequence due to the high degree of specificity that the siNA molecule requires to mediate RNAi activity.

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In one embodiment, nucleic acid molecules of the invention that act as mediators of the RNA interference gene silencing response are double-stranded nucleic acid molecules. In another embodiment, the siNA molecules of the invention consist of duplexes containing about 19 base pairs between oligonucleotides comprising about 19 to about 25 (e.g., 18, 19, 20, 21, 22, 23, 24, 25, or 26) nucleotides. In yet another embodiment, siNA molecules of the invention comprise duplexes with overhanging ends of about about 1 to about 3 (e.g., 1, 2, 3, or 4) nucleotides, for example, about 21-nucleotide duplexes with about 19 base pairs and 3'-terminal mononucleotide, dinucleotide, or trinucleotide overhangs.

In one embodiment, the invention features one or more chemically-modified siNA constructs having specificity for SARS expressing nucleic acid molecules, such as RNA encoding a SARS protein. Non-limiting examples of such chemical modifications include without limitation phosphorothioate internucleotide linkages, 2'deoxyribonucleotides, 2'-O-methyl ribonucleotides, 2'-deoxy-2'-fluoro ribonucleotides, "universal base" nucleotides, "acyclic" nucleotides, 5-C-methyl nucleotides, and terminal glyceryl and/or inverted deoxy abasic residue incorporation. These chemical modifications, when used in various siNA constructs, are shown to preserve RNAi activity in cells while at the same time, dramatically increasing the serum stability of these compounds. Furthermore, contrary to the data published by Parrish et al., supra, applicant demonstrates that multiple (greater than one) phosphorothioate substitutions are well-tolerated and confer substantial increases in serum stability for modified siNA constructs.

In one embodiment, a siNA molecule of the invention comprises modified nucleotides while maintaining the ability to mediate RNAi. The modified nucleotides can be used to improve *in vitro* or *in vivo* characteristics such as stability, activity, and/or bioavailability. For example, a siNA molecule of the invention can comprise modified nucleotides as a percentage of the total number of nucleotides present in the siNA molecule. As such, a siNA molecule of the invention can generally comprise about 5% to about 100% modified nucleotides (e.g., 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95% or 100% modified nucleotides). The actual percentage of modified nucleotides present in a given siNA

molecule will depend on the total number of nucleotides present in the siNA. If the siNA molecule is single stranded, the percent modification can be based upon the total number of nucleotides present in the single stranded siNA molecules. Likewise, if the siNA molecule is double stranded, the percent modification can be based upon the total number of nucleotides present in the sense strand, antisense strand, or both the sense and antisense strands.

One aspect of the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene. In one embodiment, a double stranded siNA molecule comprises one or more chemical modifications and each strand of the double-stranded siNA is about 21 nucleotides long. In one embodiment, the double-stranded siNA molecule does not contain any ribonucleotides. In another embodiment, the double-stranded siNA molecule comprises one or more ribonucleotides. In one embodiment, each strand of the double-stranded siNA molecule comprises about 19 to about 23 (e.g., about 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, or 29) nucleotides, wherein each strand comprises about 19 nucleotides that are complementary to the nucleotides of the other strand. In one embodiment, one of the strands of the double-stranded siNA molecule comprises a nucleotide sequence that is complementary to a nucleotide sequence or a portion thereof of the SARS gene, and the second strand of the double-stranded siNA molecule comprises a nucleotide sequence substantially similar to the nucleotide sequence of the SARS gene or a portion thereof.

In another embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene comprising an antisense region, wherein the antisense region comprises a nucleotide sequence that is complementary to a nucleotide sequence of the SARS gene or a portion thereof, and a sense region, wherein the sense region comprises a nucleotide sequence substantially similar to the nucleotide sequence of the SARS gene or a portion thereof. In one embodiment, the antisense region and the sense region each comprise about 19 to about 23 (e.g. about 19, 20, 21, 22, or 23) nucleotides, wherein the antisense region comprises about 19 nucleotides that are complementary to nucleotides of the sense region.

In another embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene comprising a sense region and an antisense region, wherein the antisense region comprises a nucleotide sequence that is complementary to a nucleotide sequence of RNA encoded by the SARS gene or a portion thereof and the sense region comprises a nucleotide sequence that is complementary to the antisense region.

In one embodiment, the SARS virus RNA comtemplated by the invention comprises SARS virus minus strand RNA. In another embodiment, the SARS virus RNA comtemplated by the invention comprises SARS virus plus strand RNA.

In one embodiment, a siNA molecule of the invention comprises blunt ends, i.e., ends that do not include any overhanging nucleotides. For example, a siNA molecule of the invention comprising modifications described herein (e.g., comprising nucleotides having Formulae I-VII or siNA constructs comprising Stab00-Stab22 or any combination thereof (see **Table IV**)) and/or any length described herein can comprise blunt ends or ends with no overhanging nucleotides.

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In one embodiment, any siNA molecule of the invention can comprise one or more blunt ends, i.e., where a blunt end does not have any overhanging nucleotides. In a nonlimiting example, a blunt ended siNA molecule has a number of base pairs equal to the number of nucleotides present in each strand of the siNA molecule. In another example, a siNA molecule comprises one blunt end, for example wherein the 5'-end of the antisense strand and the 3'-end of the sense strand do not have any overhanging nucleotides. In another example, a siNA molecule comprises one blunt end, for example wherein the 3'-end of the antisense strand and the 5'-end of the sense strand do not have any overhanging nucleotides. In another example, a siNA molecule comprises two blunt ends, for example wherein the 3'-end of the antisense strand and the 5'-end of the sense strand as well as the 5'-end of the antisense strand and 3'-end of the sense strand do not have any overhanging nucleotides. A blunt ended siNA molecule can comprise, for example, from about 18 to about 30 nucleotides (e.g., about 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 nucleotides). Other nucleotides present in a blunt ended siNA molecule can comprise mismatches, bulges, loops, or wobble base pairs, for example, to modulate the activity of the siNA molecule to mediate RNA interference.

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By "blunt ends" is meant symmetric termini or termini of a double stranded siNA molecule having no overhanging nucleotides. The two strands of a double stranded siNA molecule align with each other without over-hanging nucleotides at the termini. For example, a blunt ended siNA construct comprises terminal nucleotides that are complementary between the sense and antisense regions of the siNA molecule.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene, wherein the siNA molecule is assembled from two separate oligonucleotide fragments wherein one fragment comprises the sense region and the second fragment comprises the antisense region of the siNA molecule. The sense region can be connected to the antisense region via a linker molecule, such as a polynucleotide linker or a non-nucleotide linker.

In one embodiment, the invention features double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene, wherein the siNA molecule comprises about 19 to about 21 base pairs, and wherein each strand of the siNA molecule comprises one or more chemical modifications. embodiment, one of the strands of the double-stranded siNA molecule comprises a nucleotide sequence that is complementary to a nucleotide sequence of a SARS gene or a portion thereof, and the second strand of the double-stranded siNA molecule comprises a nucleotide sequence substantially similar to the nucleotide sequence or a portion thereof of the SARS gene. In another embodiment, one of the strands of the double-stranded siNA molecule comprises a nucleotide sequence that is complementary to a nucleotide sequence of a SARS gene or a portion thereof, and the second strand of the doublestranded siNA molecule comprises a nucleotide sequence substantially similar to the nucleotide sequence or a portion thereof of the SARS gene. In another embodiment, each strand of the siNA molecule comprises about 19 to about 23 nucleotides, and each strand comprises at least about 19 nucleotides that are complementary to the nucleotides of the other strand. The SARS gene can comprise, for example, sequences referred to Table I.

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In one embodiment, a siNA molecule of the invention comprises no ribonucleotides. In another embodiment, a siNA molecule of the invention comprises ribonucleotides.

In one embodiment, a siNA molecule of the invention comprises an antisense region comprising a nucleotide sequence that is complementary to a nucleotide sequence of a SARS gene or a portion thereof, and the siNA further comprises a sense region comprising a nucleotide sequence substantially similar to the nucleotide sequence of the SARS gene or a portion thereof. In another embodiment, the antisense region and the sense region each comprise about 19 to about 23 nucleotides and the antisense region comprises at least about 19 nucleotides that are complementary to nucleotides of the sense region. The SARS gene can comprise, for example, sequences referred to Table I.

In one embodiment, a siNA molecule of the invention comprises a sense region and an antisense region, wherein the antisense region comprises a nucleotide sequence that is complementary to a nucleotide sequence of RNA encoded by a SARS gene, or a portion thereof, and the sense region comprises a nucleotide sequence that is complementary to the antisense region. In another embodiment, the siNA molecule is assembled from two separate oligonucleotide fragments, wherein one fragment comprises the sense region and the second fragment comprises the antisense region of the siNA molecule. In another embodiment, the sense region is connected to the antisense region via a linker molecule. In another embodiment, the sense region is connected to the antisense region via a linker molecule, such as a nucleotide or non-nucleotide linker. The SARS gene can comprise, for example, sequences referred to Table I.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene comprising a sense region and an antisense region, wherein the antisense region comprises a nucleotide sequence that is complementary to a nucleotide sequence of RNA encoded by the SARS gene or a portion thereof and the sense region comprises a nucleotide sequence that is complementary to the antisense region, and wherein the siNA molecule has one or more modified pyrimidine and/or purine nucleotides. In one embodiment, the pyrimidine nucleotides in the sense region are 2'-O-methyl pyrimidine

nucleotides or 2'-deoxy-2'-fluoro pyrimidine nucleotides and the purine nucleotides present in the sense region are 2'-deoxy purine nucleotides. In another embodiment, the pyrimidine nucleotides in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides and the purine nucleotides present in the sense region are 2'-O-methyl purine nucleotides. In another embodiment, the pyrimidine nucleotides in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides and the purine nucleotides present in the sense region are 2'-deoxy purine nucleotides. In one embodiment, the pyrimidine nucleotides in the antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides and the purine nucleotides present in the antisense region are 2'-O-methyl or 2'-deoxy purine nucleotides. In another embodiment of any of the above-described siNA molecules, any nucleotides present in a non-complementary region of the sense strand (e.g. overhang region) are 2'-deoxy nucleotides.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene, wherein the siNA molecule is assembled from two separate oligonucleotide fragments wherein one fragment comprises the sense region and the second fragment comprises the antisense region of the siNA molecule, and wherein the fragment comprising the sense region includes a terminal cap moiety at the 5'-end, the 3'-end, or both of the 5' and 3' ends of the fragment. In another embodiment, the terminal cap moiety is an inverted deoxy abasic moiety or glyceryl moiety. In another embodiment, each of the two fragments of the siNA molecule comprise about 21 nucleotides.

In one embodiment, the invention features a siNA molecule comprising at least one modified nucleotide, wherein the modified nucleotide is a 2'-deoxy-2'-fluoro nucleotide. The siNA can be, for example, of length between about 12 and about 36 nucleotides. In another embodiment, all pyrimidine nucleotides present in the siNA are 2'-deoxy-2'-fluoro pyrimidine nucleotides. In another embodiment, the modified nucleotides in the siNA include at least one 2'-deoxy-2'-fluoro cytidine or 2'-deoxy-2'-fluoro uridine nucleotide. In another embodiment, the modified nucleotides in the siNA include at least one 2'-fluoro cytidine and at least one 2'-deoxy-2'-fluoro uridine nucleotides. In another embodiment, all uridine nucleotides present in the siNA are 2'-deoxy-2'-fluoro uridine nucleotides. In another embodiment, all cytidine nucleotides present in the siNA are 2'-

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deoxy-2'-fluoro cytidine nucleotides. In another embodiment, all adenosine nucleotides present in the siNA are 2'-deoxy-2'-fluoro adenosine nucleotides. In another embodiment, all guanosine nucleotides present in the siNA are 2'-deoxy-2'-fluoro guanosine nucleotides. The siNA can further comprise at least one modified internucleotidic linkage, such as phosphorothioate linkage. In another embodiment, the 2'-deoxy-2'-fluoronucleotides are present at specifically selected locations in the siNA that are sensitive to cleavage by ribonucleases, such as locations having pyrimidine nucleotides.

In one embodiment, the invention features a method of increasing the stability of a siNA molecule against cleavage by ribonucleases comprising introducing at least one modified nucleotide into the siNA molecule, wherein the modified nucleotide is a 2'deoxy-2'-fluoro nucleotide. In another embodiment, all pyrimidine nucleotides present in the siNA are 2'-deoxy-2'-fluoro pyrimidine nucleotides. In another embodiment, the modified nucleotides in the siNA include at least one 2'-deoxy-2'-fluoro cytidine or 2'deoxy-2'-fluoro uridine nucleotide. In another embodiment, the modified nucleotides in the siNA include at least one 2'-fluoro cytidine and at least one 2'-deoxy-2'-fluoro uridine nucleotides. In another embodiment, all uridine nucleotides present in the siNA are 2'-deoxy-2'-fluoro uridine nucleotides. In another embodiment, all cytidine nucleotides present in the siNA are 2'-deoxy-2'-fluoro cytidine nucleotides. In another embodiment, all adenosine nucleotides present in the siNA are 2'-deoxy-2'-fluoro adenosine nucleotides. In another embodiment, all guanosine nucleotides present in the siNA are 2'-deoxy-2'-fluoro guanosine nucleotides. The siNA can further comprise at least one modified internucleotidic linkage, such as phosphorothioate linkage. In another embodiment, the 2'-deoxy-2'-fluoronucleotides are present at specifically selected locations in the siNA that are sensitive to cleavage by ribonucleases, such as locations having pyrimidine nucleotides.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene comprising a sense region and an antisense region, wherein the antisense region comprises a nucleotide sequence that is complementary to a nucleotide sequence of RNA encoded by the SARS gene or a portion thereof and the sense region comprises a

nucleotide sequence that is complementary to the antisense region, and wherein the purine nucleotides present in the antisense region comprise 2'-deoxy- purine nucleotides. In an alternative embodiment, the purine nucleotides present in the antisense region comprise 2'-O-methyl purine nucleotides. In either of the above embodiments, the antisense region can comprise a phosphorothioate internucleotide linkage at the 3' end of the antisense region. Alternatively, in either of the above embodiments, the antisense region can comprise a glyceryl modification at the 3' end of the antisense region. In another embodiment of any of the above-described siNA molecules, any nucleotides present in a non-complementary region of the antisense strand (e.g. overhang region) are 2'-deoxy nucleotides.

In one embodiment, the antisense region of a siNA molecule of the invention comprises sequence complementary to a portion of a SARS transcript having sequence unique to a particular SARS disease related allele, such as sequence comprising a SNP associated with the disease specific allele. As such, the antisense region of a siNA molecule of the invention can comprise sequence complementary to sequences that are unique to a particular allele to provide specificity in mediating selective RNAi againt the disease related allele.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that down-regulates expression of a SARS gene, wherein the siNA molecule is assembled from two separate oligonucleotide fragments wherein one fragment comprises the sense region and the second fragment comprises the antisense region of the siNA molecule. In another embodiment about 19 nucleotides of each fragment of the siNA molecule are base-paired to the complementary nucleotides of the other fragment of the siNA molecule and wherein at least two 3' terminal nucleotides of each fragment of the siNA molecule are not base-paired to the nucleotides of the other fragment of the siNA molecule. In one embodiment, each of the two 3' terminal nucleotides of each fragment of the siNA molecule is a 2'-deoxy-pyrimidine nucleotide, such as a 2'-deoxy-thymidine. In another embodiment, all 21 nucleotides of each fragment of the siNA molecule are base-paired to the complementary nucleotides of the other fragment of the siNA molecule. In another embodiment, about 19 nucleotides of the antisense region are base-paired to the nucleotide sequence or a portion thereof of the

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RNA encoded by the SARS gene. In another embodiment, about 21 nucleotides of the antisense region are base-paired to the nucleotide sequence or a portion thereof of the RNA encoded by the SARS gene. In any of the above embodiments, the 5'-end of the fragment comprising said antisense region can optionally includes a phosphate group.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits the expression of a SARS RNA sequence (e.g., wherein said target RNA sequence is encoded by a SARS gene involved in the SARS pathway), wherein the siNA molecule does not contain any ribonucleotides and wherein each strand of the double-stranded siNA molecule is about 21 nucleotides long. Examples of non-ribonucleotide containing siNA constructs are combinations of stabilization chemistries shown in Table IV in any combination of Sense/Antisense chemistries, such as Stab 7/8, Stab 7/11, Stab 8/8, Stab 18/8, Stab 18/11, Stab 12/13, Stab 7/13, Stab 18/13, Stab 7/19, Stab 8/19, Stab 18/19, Stab 7/20, Stab 8/20, or Stab 18/20.

In one embodiment, the invention features a chemically synthesized double stranded RNA molecule that directs cleavage of a SARS RNA via RNA interference, wherein each strand of said RNA molecule is about 21 to about 23 nucleotides in length; one strand of the RNA molecule comprises nucleotide sequence having sufficient complementarity to the SARS RNA for the RNA molecule to direct cleavage of the SARS RNA via RNA interference; and wherein at least one strand of the RNA molecule comprises one or more chemically modified nucleotides described herein, such as deoxynucleotides, 2'-O-methyl nucleotides, 2'-deoxy-2'-fluoro nucloetides, 2'-O-methoxyethyl nucleotides etc.

In one embodiment, the invention features a medicament comprising a siNA molecule of the invention.

In one embodiment, the invention features an active ingredient comprising a siNA molecule of the invention.

In one embodiment, the invention features the use of a double-stranded short interfering nucleic acid (siNA) molecule to down-regulate expression of a SARS gene,

wherein the siNA molecule comprises one or more chemical modifications and each strand of the double-stranded siNA is about 18 to about 28 or more (e.g., 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28 or more) nucleotides long.

In one embodiment, the invention features the use of a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification.

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In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, wherein the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA that encodes a protein or portion thereof, the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification. In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, the other

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strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification. In one embodiment, each strand of the siNA molecule comprises about 18 to about 29 or more (e.g., about 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or more) nucleotides, wherein each strand comprises at least about 18 nucleotides that are complementary to the nucleotides of the other strand. In another embodiment, the siNA molecule is assembled from two oligonucleotide fragments, wherein one fragment comprises the nucleotide sequence of the antisense strand of the siNA molecule and a second fragment comprises nucleotide sequence of the sense region of the siNA molecule. In yet another embodiment, the sense strand is connected to the antisense strand via a linker molecule, such as a polynucleotide linker or a non-nucleotide linker. In a further embodiment, the pyrimidine nucleotides present in the sense strand are 2'deoxy-2'fluoro pyrimidine nucleotides and the purine nucleotides present in the sense region are 2'-deoxy purine nucleotides. In another embodiment, the pyrimidine nucleotides present in the sense strand are 2'-deoxy-2'fluoro pyrimidine nucleotides and the purine nucleotides present in the sense region are 2'-O-methyl purine nucleotides. In still another embodiment, the pyrimidine nucleotides present in the antisense strand are 2'-deoxy-2'-fluoro pyrimidine nucleotides and any purine nucleotides present in the antisense strand are 2'-deoxy purine nucleotides. In another embodiment, the antisense strand comprises one or more 2'-deoxy-2'-fluoro pyrimidine nucleotides and one or more 2'-O-methyl purine nucleotides. In another embodiment, the pyrimidine nucleotides present in the antisense strand are 2'-deoxy-2'-fluoro pyrimidine nucleotides and any purine nucleotides present in the antisense strand are 2'-O-methyl purine nucleotides. In a further embodiment the sense strand comprises a 3'-end and a 5'-end, wherein a terminal cap moiety (e.g., an inverted deoxy abasic moiety or inverted deoxy nucleotide moiety such as inverted thymidine) is present at the 5'-end, the 3'-end, or both of the 5' and 3' ends of the sense strand. In another embodiment, the antisense strand comprises a phosphorothioate internucleotide linkage at the 3' end of the antisense strand. In another embodiment, the antisense strand comprises a glyceryl modification at the 3' end. In another embodiment, the 5'-end of the antisense strand optionally includes a phosphate group.

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In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, wherein the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification, and wherein each of the two strands of the siNA molecule comprises about 21 nucleotides. In one embodiment, about 21 nucleotides of each strand of the siNA molecule are base-paired to the complementary nucleotides of the other strand of the siNA molecule. In another embodiment, about 19 nucleotides of each strand of the siNA molecule are base-paired to the complementary nucleotides of the other strand of the siNA molecule, wherein at least two 3' terminal nucleotides of each strand of the siNA molecule are not base-paired to the nucleotides of the other strand of the siNA molecule. In another embodiment, each of the two 3' terminal nucleotides of each fragment of the siNA molecule is a 2'-deoxy-pyrimidine, such as 2'-deoxy-thymidine. In another embodiment, each strand of the siNA molecule is base-paired to the complementary nucleotides of the other strand of the siNA molecule. In another embodiment, about 19 nucleotides of the antisense strand are base-paired to the nucleotide sequence of the SARS RNA or a portion thereof. In another embodiment, about 21 nucleotides of the antisense strand are base-paired to the nucleotide sequence of the SARS RNA or a portion thereof.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification, and wherein the 5'-end of the antisense strand optionally includes a phosphate group.

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In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand and wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification, and wherein the nucleotide sequence or a portion thereof of the antisense strand is complementary to a nucleotide sequence of the untranslated region or a portion thereof of the SARS RNA.

In one embodiment, the invention features a double-stranded short interfering nucleic acid (siNA) molecule that inhibits expression of a SARS gene, wherein one of the strands of the double-stranded siNA molecule is an antisense strand which comprises nucleotide sequence that is complementary to nucleotide sequence of SARS RNA or a portion thereof, wherein the other strand is a sense strand which comprises nucleotide sequence that is complementary to a nucleotide sequence of the antisense strand, wherein a majority of the pyrimidine nucleotides present in the double-stranded siNA molecule comprises a sugar modification, and wherein the nucleotide sequence of the antisense strand is complementary to a nucleotide sequence of the SARS RNA or a portion thereof that is present in the SARS RNA.

In one embodiment, the invention features a composition comprising a siNA molecule of the invention in a pharmaceutically acceptable carrier or diluent.

In a non-limiting example, the introduction of chemically-modified nucleotides into nucleic acid molecules provides a powerful tool in overcoming potential limitations of *in vivo* stability and bioavailability inherent to native RNA molecules that are delivered exogenously. For example, the use of chemically-modified nucleic acid molecules can enable a lower dose of a particular nucleic acid molecule for a given therapeutic effect since chemically-modified nucleic acid molecules tend to have a longer half-life in serum. Furthermore, certain chemical modifications can improve the bioavailability of nucleic acid molecules by targeting particular cells or tissues and/or improving cellular uptake of the nucleic acid molecule. Therefore, even if the activity of

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a chemically-modified nucleic acid molecule is reduced as compared to a native nucleic acid molecule, for example, when compared to an all-RNA nucleic acid molecule, the overall activity of the modified nucleic acid molecule can be greater than that of the native molecule due to improved stability and/or delivery of the molecule. Unlike native unmodified siNA, chemically-modified siNA can also minimize the possibility of activating interferon activity in humans.

In any of the embodiments of siNA molecules described herein, the antisense region of a siNA molecule of the invention can comprise a phosphorothioate internucleotide linkage at the 3'-end of said antisense region. In any of the embodiments of siNA molecules described herein, the antisense region can comprise about one to about five phosphorothioate internucleotide linkages at the 5'-end of said antisense region. In any of the embodiments of siNA molecules described herein, the 3'-terminal nucleotide overhangs of a siNA molecule of the invention can comprise ribonucleotides or deoxyribonucleotides that are chemically-modified at a nucleic acid sugar, base, or backbone. In any of the embodiments of siNA molecules described herein, the 3'-terminal nucleotide overhangs can comprise one or more universal base ribonucleotides. In any of the embodiments of siNA molecules described herein, the 3'-terminal nucleotide overhangs can comprise one or more acyclic nucleotides.

One embodiment of the invention provides an expression vector comprising a nucleic acid sequence encoding at least one siNA molecule of the invention in a manner that allows expression of the nucleic acid molecule. Another embodiment of the invention provides a mammalian cell comprising such an expression vector. The mammalian cell can be a human cell. The siNA molecule of the expression vector can comprise a sense region and an antisense region. The antisense region can comprise sequence complementary to a RNA or DNA sequence encoding SARS and the sense region can comprise sequence complementary to the antisense region. The siNA molecule can comprise two distinct strands having complementary sense and antisense regions. The siNA molecule can comprise a single strand having complementary sense and antisense regions.

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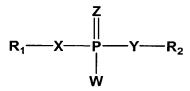
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In one embodiment, the nucleotide sequence of the antisense strand or a portion thereof of a siNA molecule of the invention is complementary to the nucleotide sequence of a SARS RNA or a portion thereof that is present in the RNA of all SARS isolates.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted *in vitro* system, wherein the chemical modification comprises one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) nucleotides comprising a backbone modified internucleotide linkage having Formula I:



wherein each R1 and R2 is independently any nucleotide, non-nucleotide, or polynucleotide which can be naturally-occurring or chemically-modified, each X and Y is independently O, S, N, alkyl, or substituted alkyl, each Z and W is independently O, S, N, alkyl, substituted alkyl, O-alkyl, S-alkyl, alkaryl, aralkyl, or acetyl and wherein W, X, Y, and Z are optionally not all O. In another embodiment, a backbone modification of the invention comprises a phosphonoacetate and/or thiophosphonoacetate internucleotide linkage (see for example Sheehan et al., 2003, Nucleic Acids Research, 31, 4109-4118).

The chemically-modified internucleotide linkages having Formula I, for example, wherein any Z, W, X, and/or Y independently comprises a sulphur atom, can be present in one or both oligonucleotide strands of the siNA duplex, for example, in the sense strand, the antisense strand, or both strands. The siNA molecules of the invention can comprise one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) chemically-modified internucleotide linkages having Formula I at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of the sense strand, the antisense strand, or both strands. For example, an exemplary siNA molecule of the invention can comprise about 1 to about 5 or more (e.g., about 1, 2, 3, 4, 5, or more) chemically-modified internucleotide linkages having Formula I at the 5'-end of the sense strand, the antisense strand, or both strands. In another non-limiting example, an exemplary siNA molecule of the invention can comprise one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) pyrimidine

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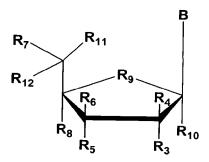
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nucleotides with chemically-modified internucleotide linkages having Formula I in the sense strand, the antisense strand, or both strands. In yet another non-limiting example, an exemplary siNA molecule of the invention can comprise one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) purine nucleotides with chemically-modified internucleotide linkages having Formula I in the sense strand, the antisense strand, or both strands. In another embodiment, a siNA molecule of the invention having internucleotide linkage(s) of Formula I also comprises a chemically-modified nucleotide or non-nucleotide having any of Formulae I-VII.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted *in vitro* system, wherein the chemical modification comprises one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) nucleotides or non-nucleotides having Formula II:

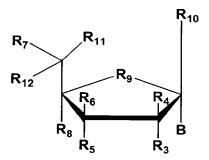


wherein each R3, R4, R5, R6, R7, R8, R10, R11 and R12 is independently H, OH, alkyl, substituted alkyl, alkaryl or aralkyl, F, Cl, Br, CN, CF3, OCF3, OCN, O-alkyl, S-alkyl, N-alkyl, O-alkyl, S-alkyl, N-alkyl, O-alkyl, S-alkyl, S-alkyl, alkyl-OSH, alkyl-OH, O-alkyl-OH, O-alkyl-SH, S-alkyl-OH, S-alkyl-SH, alkyl-S-alkyl, alkyl-O-alkyl, ONO2, NO2, N3, NH2, aminoalkyl, aminoacid, aminoacyl, ONH2, O-aminoalkyl, O-aminoacid, O-aminoacyl, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalklylamino, substituted silyl, or group having Formula I or II; R9 is O, S, CH2, S=O, CHF, or CF2, and B is a nucleosidic base such as adenine, guanine, uracil, cytosine, thymine, 2-aminoadenosine, 5-methylcytosine, 2,6-diaminopurine, or any other non-naturally occurring base that can be complementary or non-complementary to target RNA or a non-nucleosidic base such as phenyl, naphthyl, 3-nitropyrrole, 5-nitroindole, nebularine,

pyridone, pyridinone, or any other non-naturally occurring universal base that can be complementary or non-complementary to target RNA.

The chemically-modified nucleotide or non-nucleotide of Formula II can be present in one or both oligonucleotide strands of the siNA duplex, for example in the sense strand, the antisense strand, or both strands. The siNA molecules of the invention can comprise one or more chemically-modified nucleotide or non-nucleotide of Formula II at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of the sense strand, the antisense strand, or both strands. For example, an exemplary siNA molecule of the invention can comprise about 1 to about 5 or more (e.g., about 1, 2, 3, 4, 5, or more) chemically-modified nucleotides or non-nucleotides of Formula II at the 5'-end of the sense strand, the antisense strand, or both strands. In anther non-limiting example, an exemplary siNA molecule of the invention can comprise about 1 to about 5 or more (e.g., about 1, 2, 3, 4, 5, or more) chemically-modified nucleotides or non-nucleotides of Formula II at the 3'-end of the sense strand, the antisense strand, or both strands.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted *in vitro* system, wherein the chemical modification comprises one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) nucleotides or non-nucleotides having Formula III:



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wherein each R3, R4, R5, R6, R7, R8, R10, R11 and R12 is independently H, OH, alkyl, substituted alkyl, alkaryl or aralkyl, F, Cl, Br, CN, CF3, OCF3, OCN, O-alkyl, S-alkyl, N-alkyl, O-alkenyl, S-alkenyl, N-alkenyl, SO-alkyl, alkyl-OSH, alkyl-OH, O-alkyl-OH, O-alkyl-SH, S-alkyl-OH, S-alkyl-SH, alkyl-S-alkyl, alkyl-O-alkyl, ONO2, NO2, N3, NH2, aminoalkyl, aminoacid, aminoacyl, ONH2, O-aminoalkyl, O-aminoacid, O-

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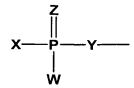
aminoacyl, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalklylamino, substituted silyl, or group having Formula I or II; R9 is O, S, CH2, S=O, CHF, or CF2, and B is a nucleosidic base such as adenine, guanine, uracil, cytosine, thymine, 2-aminoadenosine, 5-methylcytosine, 2,6-diaminopurine, or any other non-naturally occurring base that can be employed to be complementary or non-complementary to target RNA or a non-nucleosidic base such as phenyl, naphthyl, 3-nitropyrrole, 5-nitroindole, nebularine, pyridone, pyridinone, or any other non-naturally occurring universal base that can be complementary or non-complementary to target RNA.

The chemically-modified nucleotide or non-nucleotide of Formula III can be present in one or both oligonucleotide strands of the siNA duplex, for example, in the sense strand, the antisense strand, or both strands. The siNA molecules of the invention can comprise one or more chemically-modified nucleotide or non-nucleotide of Formula III at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of the sense strand, the antisense strand, or both strands. For example, an exemplary siNA molecule of the invention can comprise about 1 to about 5 or more (e.g., about 1, 2, 3, 4, 5, or more) chemically-modified nucleotide(s) or non-nucleotide(s) of Formula III at the 5'-end of the sense strand, the antisense strand, or both strands. In anther non-limiting example, an exemplary siNA molecule of the invention can comprise about 1 to about 5 or more (e.g., about 1, 2, 3, 4, 5, or more) chemically-modified nucleotide or non-nucleotide of Formula III at the 3'-end of the sense strand, the antisense strand, or both strands.

In another embodiment, a siNA molecule of the invention comprises a nucleotide having Formula II or III, wherein the nucleotide having Formula II or III is in an inverted configuration. For example, the nucleotide having Formula II or III is connected to the siNA construct in a 3'-3', 3'-2', 2'-3', or 5'-5' configuration, such as at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of one or both siNA strands.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted *in vitro* system, wherein the chemical modification comprises a 5'-terminal phosphate group having Formula IV:

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wherein each X and Y is independently O, S, N, alkyl, substituted alkyl, or alkylhalo; wherein each Z and W is independently O, S, N, alkyl, substituted alkyl, O-alkyl, S-alkyl, alkaryl, aralkyl, alkylhalo, or acetyl; and wherein W, X, Y and Z are not all O.

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In one embodiment, the invention features a siNA molecule having a 5'-terminal phosphate group having Formula IV on the target-complementary strand, for example, a strand complementary to a target RNA, wherein the siNA molecule comprises an all RNA siNA molecule. In another embodiment, the invention features a siNA molecule having a 5'-terminal phosphate group having Formula IV on the target-complementary strand wherein the siNA molecule also comprises about 1 to about 3 (e.g., about 1, 2, or 3) nucleotide 3'-terminal nucleotide overhangs having about 1 to about 4 (e.g., about 1, 2, 3, or 4) deoxyribonucleotides on the 3'-end of one or both strands. In another embodiment, a 5'-terminal phosphate group having Formula IV is present on the target-complementary strand of a siNA molecule of the invention, for example a siNA molecule having chemical modifications having any of Formulae I-VII.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted *in vitro* system, wherein the chemical modification comprises one or more phosphorothioate internucleotide linkages. For example, in a non-limiting example, the invention features a chemically-modified short interfering nucleic acid (siNA) having about 1, 2, 3, 4, 5, 6, 7, 8 or more phosphorothioate internucleotide linkages in one siNA strand. In yet another embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) individually having about 1, 2, 3, 4, 5, 6, 7, 8 or more phosphorothioate internucleotide linkages in both siNA strands. The phosphorothioate internucleotide linkages can be present in one or both oligonucleotide strands of the siNA duplex, for example in the sense strand, the antisense strand, or both strands. The siNA molecules of the invention can comprise one or more phosphorothioate internucleotide linkages at the 3'-end, or both of the 3'- and

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5'-ends of the sense strand, the antisense strand, or both strands. For example, an exemplary siNA molecule of the invention can comprise about 1 to about 5 or more (e.g., about 1, 2, 3, 4, 5, or more) consecutive phosphorothioate internucleotide linkages at the 5'-end of the sense strand, the antisense strand, or both strands. In another non-limiting example, an exemplary siNA molecule of the invention can comprise one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) pyrimidine phosphorothioate internucleotide linkages in the sense strand, the antisense strand, or both strands. In yet another non-limiting example, an exemplary siNA molecule of the invention can comprise one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) purine phosphorothioate internucleotide linkages in the sense strand, the antisense strand, or both strands.

In one embodiment, the invention features a siNA molecule, wherein the sense strand comprises one or more, for example, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or about one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of the sense strand; and wherein the antisense strand comprises about 1 to about 10 or more, specifically about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of the antisense strand. In another embodiment, one or more, for example about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more, pyrimidine nucleotides of the sense and/or antisense siNA strand are chemically-modified with 2'-deoxy, 2'-O-methyl and/or 2'-deoxy-2'-fluoro nucleotides, with or without one or more, for example about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more, phosphorothioate internucleotide linkages and/or a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends, being present in the same or different strand.

In another embodiment, the invention features a siNA molecule, wherein the sense strand comprises about 1 to about 5, specifically about 1, 2, 3, 4, or 5 phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, or more) 2'-deoxy,

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2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 4, 5, or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3-end, the 5'-end, or both of the 3'- and 5'-ends of the sense strand; and wherein the antisense strand comprises about 1 to about 5 or more, specifically about 1, 2, 3, 4, 5, or more phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of the antisense strand. In another embodiment, one or more, for example about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more, pyrimidine nucleotides of the sense and/or antisense siNA strand are chemically-modified with 2'-deoxy, 2'-O-methyl and/or 2'-deoxy-2'-fluoro nucleotides, with or without about 1 to about 5 or more, for example about 1, 2, 3, 4, 5, or more phosphorothioate internucleotide linkages and/or a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends, being present in the same or different strand.

In one embodiment, the invention features a siNA molecule, wherein the antisense strand comprises one or more, for example, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more phosphorothioate internucleotide linkages, and/or about one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and 20 optionally a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of the sense strand; and wherein the antisense strand comprises about 1 to about 10 or more, specifically about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 25 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of the antisense strand. In another embodiment, one or more, for example about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more pyrimidine nucleotides of the sense and/or antisense siNA strand are chemically-modified with 2'-deoxy, 2'-O-methyl and/or 2'-deoxy-2'-fluoro nucleotides, 30 with or without one or more, for example, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more

phosphorothioate internucleotide linkages and/or a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3' and 5'-ends, being present in the same or different strand.

In another embodiment, the invention features a siNA molecule, wherein the antisense strand comprises about 1 to about 5 or more, specifically about 1, 2, 3, 4, 5 or more phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of the sense strand; and wherein the antisense strand comprises about 1 to about 5 or more, specifically about 1, 2, 3, 4, 5 or more phosphorothioate internucleotide linkages, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) 2'-deoxy, 2'-O-methyl, 2'-deoxy-2'-fluoro, and/or one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more) universal base modified nucleotides, and optionally a terminal cap molecule at the 3'end, the 5'-end, or both of the 3'- and 5'-ends of the antisense strand. In another embodiment, one or more, for example about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more pyrimidine nucleotides of the sense and/or antisense siNA strand are chemicallymodified with 2'-deoxy, 2'-O-methyl and/or 2'-deoxy-2'-fluoro nucleotides, with or without about 1 to about 5, for example about 1, 2, 3, 4, 5 or more phosphorothioate internucleotide linkages and/or a terminal cap molecule at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends, being present in the same or different strand.

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In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule having about 1 to about 5, specifically about 1, 2, 3, 4, 5 or more phosphorothioate internucleotide linkages in each strand of the siNA molecule.

In another embodiment, the invention features a siNA molecule comprising 2'-5' internucleotide linkages. The 2'-5' internucleotide linkage(s) can be at the 3'-end, the 5'-end, or both of the 3'- and 5'-ends of one or both siNA sequence strands. In addition, the 2'-5' internucleotide linkage(s) can be present at various other positions within one or both siNA sequence strands, for example, about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more including every internucleotide linkage of a pyrimidine nucleotide in one or both strands of the siNA molecule can comprise a 2'-5' internucleotide linkage, or about 1, 2, 3, 4, 5,

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6, 7, 8, 9, 10, or more including every internucleotide linkage of a purine nucleotide in one or both strands of the siNA molecule can comprise a 2'-5' internucleotide linkage.

In another embodiment, a chemically-modified siNA molecule of the invention comprises a duplex having two strands, one or both of which can be chemicallymodified, wherein each strand is about 18 to about 27 (e.g., about 18, 19, 20, 21, 22, 23, 24, 25, 26, or 27) nucleotides in length, wherein the duplex has about 18 to about 23 (e.g., about 18, 19, 20, 21, 22, or 23) base pairs, and wherein the chemical modification comprises a structure having any of Formulae I-VII. For example, an exemplary chemically-modified siNA molecule of the invention comprises a duplex having two strands, one or both of which can be chemically-modified with a chemical modification having any of Formulae I-VII or any combination thereof, wherein each strand consists of about 21 nucleotides, each having a 2-nucleotide 3'-terminal nucleotide overhang, and wherein the duplex has about 19 base pairs. In another embodiment, a siNA molecule of the invention comprises a single stranded hairpin structure, wherein the siNA is about 36 to about 70 (e.g., about 36, 40, 45, 50, 55, 60, 65, or 70) nucleotides in length having about 18 to about 23 (e.g., about 18, 19, 20, 21, 22, or 23) base pairs, and wherein the siNA can include a chemical modification comprising a structure having any of Formulae I-VII or any combination thereof. For example, an exemplary chemicallymodified siNA molecule of the invention comprises a linear oligonucleotide having about 42 to about 50 (e.g., about 42, 43, 44, 45, 46, 47, 48, 49, or 50) nucleotides that is chemically-modified with a chemical modification having any of Formulae I-VII or any combination thereof, wherein the linear oligonucleotide forms a hairpin structure having about 19 base pairs and a 2-nucleotide 3'-terminal nucleotide overhang. In another embodiment, a linear hairpin siNA molecule of the invention contains a stem loop motif, wherein the loop portion of the siNA molecule is biodegradable. For example, a linear hairpin siNA molecule of the invention is designed such that degradation of the loop portion of the siNA molecule in vivo can generate a double-stranded siNA molecule with 3'-terminal overhangs, such as 3'-terminal nucleotide overhangs comprising about 2 nucleotides.

In another embodiment, a siNA molecule of the invention comprises a hairpin structure, wherein the siNA is about 25 to about 50 (e.g., about 25, 26, 27, 28, 29, 30, 31,

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32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50) nucleotides in length having about 3 to about 25 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25) base pairs, and wherein the siNA can include one or more chemical modifications comprising a structure having any of Formulae I-VII or any combination thereof. For example, an exemplary chemically-modified siNA molecule of the invention comprises a linear oligonucleotide having about 25 to about 35 (e.g., about 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, or 35) nucleotides that is chemically-modified with one or more chemical modifications having any of Formulae I-VII or any combination thereof, wherein the linear oligonucleotide forms a hairpin structure having about 3 to about 23 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, or 23) base pairs and a 5'-terminal phosphate group that can be chemically modified as described herein (for example a 5'-terminal phosphate group having Formula IV). In another embodiment, a linear hairpin siNA molecule of the invention contains a stem loop motif, wherein the loop portion of the siNA molecule is biodegradable. In another embodiment, a linear hairpin siNA molecule of the invention comprises a loop portion comprising a non-nucleotide linker.

In another embodiment, a siNA molecule of the invention comprises an asymmetric hairpin structure, wherein the siNA is about 25 to about 50 (e.g., about 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50) nucleotides in length having about 3 to about 20 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20) base pairs, and wherein the siNA can include one or more chemical modifications comprising a structure having any of Formulae I-VII or any combination thereof. For example, an exemplary chemically-modified siNA molecule of the invention comprises a linear oligonucleotide having about 25 to about 35 (e.g., about 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, or 35) nucleotides that is chemicallymodified with one or more chemical modifications having any of Formulae I-VII or any combination thereof, wherein the linear oligonucleotide forms an asymmetric hairpin structure having about 3 to about 18 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17 or 18) base pairs and a 5'-terminal phosphate group that can be chemically modified as described herein (for example a 5'-terminal phosphate group having Formula IV). In another embodiment, an asymmetric hairpin siNA molecule of the invention contains a stem loop motif, wherein the loop portion of the siNA molecule is

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biodegradable. In another embodiment, an asymmetric hairpin siNA molecule of the invention comprises a loop portion comprising a non-nucleotide linker.

In another embodiment, a siNA molecule of the invention comprises an asymmetric double stranded structure having separate polynucleotide strands comprising sense and antisense regions, wherein the antisense region is about 16 to about 25 (e.g., about 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25) nucleotides in length, wherein the sense region is about 3 to about 18 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, or 18) nucleotides in length, wherein the sense region and the antisense region have at least 3 complementary nucleotides, and wherein the siNA can include one or more chemical modifications comprising a structure having any of Formulae I-VII or any combination thereof. For example, an exemplary chemically-modified siNA molecule of the invention comprises an asymmetric double stranded structure having separate polynucleotide strands comprising sense and antisense regions, wherein the antisense region is about 18 to about 22 (e.g., about 18, 19, 20, 21, or 22) nucleotides in length and wherein the sense region is about 3 to about 15 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, or 15) nucleotides in length, wherein the sense region the antisense region have at least 3 complementary nucleotides, and wherein the siNA can include one or more chemical modifications comprising a structure having any of Formulae I-VII or any combination thereof. In another embodiment, the asymmetic double stranded siNA molecule can also have a 5'-terminal phosphate group that can be chemically modified as described herein (for example a 5'-terminal phosphate group having Formula IV).

In another embodiment, a siNA molecule of the invention comprises a circular nucleic acid molecule, wherein the siNA is about 38 to about 70 (e.g., about 38, 40, 45, 50, 55, 60, 65, or 70) nucleotides in length having about 18 to about 23 (e.g., about 18, 19, 20, 21, 22, or 23) base pairs, and wherein the siNA can include a chemical modification, which comprises a structure having any of Formulae I-VII or any combination thereof. For example, an exemplary chemically-modified siNA molecule of the invention comprises a circular oligonucleotide having about 42 to about 50 (e.g., about 42, 43, 44, 45, 46, 47, 48, 49, or 50) nucleotides that is chemically-modified with a chemical modification having any of Formulae I-VII or any combination thereof,

wherein the circular oligonucleotide forms a dumbbell shaped structure having about 19 base pairs and 2 loops.

In another embodiment, a circular siNA molecule of the invention contains two loop motifs, wherein one or both loop portions of the siNA molecule is biodegradable. For example, a circular siNA molecule of the invention is designed such that degradation of the loop portions of the siNA molecule *in vivo* can generate a double-stranded siNA molecule with 3'-terminal overhangs, such as 3'-terminal nucleotide overhangs comprising about 2 nucleotides.

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In one embodiment, a siNA molecule of the invention comprises at least one (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) abasic moiety, for example a compound having Formula V:

wherein each R3, R4, R5, R6, R7, R8, R10, R11, R12, and R13 is independently H, OH, alkyl, substituted alkyl, alkaryl or aralkyl, F, Cl, Br, CN, CF3, OCF3, OCN, O-alkyl, S-alkyl, N-alkyl, O-alkyl, S-alkyl, N-alkyl, O-alkyl, S-alkyl-OH, O-alkyl-OH, O-alkyl-SH, S-alkyl-OH, S-alkyl-SH, alkyl-S-alkyl, alkyl-O-alkyl, ONO2, NO2, N3, NH2, aminoalkyl, aminoacid, aminoacyl, ONH2, O-aminoalkyl, O-aminoacid, O-aminoacyl, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalklylamino, substituted silyl, or group having Formula I or II; R9 is O, S, CH2, S=O, CHF, or CF2.

In one embodiment, a siNA molecule of the invention comprises at least one (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) inverted abasic moiety, for example a compound having Formula VI:

wherein each R3, R4, R5, R6, R7, R8, R10, R11, R12, and R13 is independently H, OH, alkyl, substituted alkyl, alkaryl or aralkyl, F, Cl, Br, CN, CF3, OCF3, OCN, O-alkyl, S-alkyl, N-alkyl, O-alkenyl, S-alkenyl, N-alkenyl, SO-alkyl, alkyl-OSH, alkyl-OH, O-alkyl-OH, O-alkyl-SH, S-alkyl-OH, S-alkyl-SH, alkyl-S-alkyl, alkyl-O-alkyl, ONO2, NO2, N3, NH2, aminoalkyl, aminoacid, aminoacyl, ONH2, O-aminoalkyl, O-aminoacid, O-aminoacyl, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalklylamino, substituted silyl, or group having Formula I or II; R9 is O, S, CH2, S=O, CHF, or CF2, and either R2, R3, R8 or R13 serve as points of attachment to the siNA molecule of the invention.

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In another embodiment, a siNA molecule of the invention comprises at least one (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) substituted polyalkyl moieties, for example a compound having Formula VII:

$$R_1$$
 R_2
 R_3

wherein each n is independently an integer from 1 to 12, each R1, R2 and R3 is independently H, OH, alkyl, substituted alkyl, alkaryl or aralkyl, F, Cl, Br, CN, CF3, OCF3, OCN, O-alkyl, S-alkyl, O-alkyl, O-alkenyl, S-alkenyl, N-alkenyl, SO-alkyl, alkyl-OSH, alkyl-OH, O-alkyl-OH, O-alkyl-SH, S-alkyl-OH, S-alkyl-SH, alkyl-S-alkyl, alkyl-O-alkyl, ONO2, NO2, N3, NH2, aminoalkyl, aminoacid, aminoacyl, ONH2, O-aminoacyl, O-aminoacyl, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalklylamino, substituted silyl, or a group having Formula I, and R1, R2 or R3 serves as points of attachment to the siNA molecule of the invention.

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In another embodiment, the invention features a compound having Formula VII, wherein R1 and R2 are hydroxyl (OH) groups, n = 1, and R3 comprises O and is the point of attachment to the 3'-end, the 5'-end, or both of the 3' and 5'-ends of one or both strands of a double-stranded siNA molecule of the invention or to a single-stranded siNA molecule of the invention. This modification is referred to herein as "glyceryl" (for example modification 6 in Figure 10).

In another embodiment, a moiety having any of Formula V, VI or VII of the invention is at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of a siNA molecule of the invention. For example, a moiety having Formula V, VI or VII can be present at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of the antisense strand, the sense strand, or both antisense and sense strands of the siNA molecule. In addition, a moiety having Formula VII can be present at the 3'-end or the 5'-end of a hairpin siNA molecule as described herein.

In another embodiment, a siNA molecule of the invention comprises an abasic residue having Formula V or VI, wherein the abasic residue having Formula VI or VI is connected to the siNA construct in a 3'-3', 3'-2', 2'-3', or 5'-5' configuration, such as at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of one or both siNA strands.

In one embodiment, a siNA molecule of the invention comprises one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) locked nucleic acid (LNA) nucleotides, for example at the 5'-end, the 3'-end, both of the 5' and 3'-ends, or any combination thereof, of the siNA molecule.

In another embodiment, a siNA molecule of the invention comprises one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) acyclic nucleotides, for example at the 5'-end, the 3'-end, both of the 5' and 3'-ends, or any combination thereof, of the siNA molecule.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising a sense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-

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2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any (e.g., one or more or all) purine nucleotides present in the sense region are 2'-deoxy purine nucleotides (e.g., wherein all purine nucleotides are 2'-deoxy purine nucleotides or alternately a plurality of purine nucleotides are 2'-deoxy purine nucleotides).

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising a sense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any (e.g., one or more or all) purine nucleotides present in the sense region are 2'-deoxy purine nucleotides (e.g., wherein all purine nucleotides are 2'-deoxy purine nucleotides or alternately a plurality of purine nucleotides are 2'-deoxy purine nucleotides), wherein any nucleotides comprising a 3'-terminal nucleotide overhang that are present in said sense region are 2'-deoxy nucleotides.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising a sense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any (e.g., one or more or all) purine nucleotides present in the sense region are 2'-O-methyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides).

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising a sense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), wherein any (e.g., one or more or all) purine

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nucleotides present in the sense region are 2'-O-methyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides), and wherein any nucleotides comprising a 3'-terminal nucleotide overhang that are present in said sense region are 2'-deoxy nucleotides.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising an antisense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any (e.g., one or more or all) purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides).

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising an antisense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), wherein any (e.g., one or more or all) purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides), and wherein any nucleotides comprising a 3'-terminal nucleotide overhang that are present in said antisense region are 2'-deoxy nucleotides.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising an antisense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any (e.g., one or

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more or all) purine nucleotides present in the antisense region are 2'-deoxy purine nucleotides (e.g., wherein all purine nucleotides are 2'-deoxy purine nucleotides or alternately a plurality of purine nucleotides are 2'-deoxy purine nucleotides).

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention comprising an antisense region, wherein any (e.g., one or more or all) pyrimidine nucleotides present in the antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any (e.g., one or more or all) purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides).

In one embodiment, the invention features a chemically-modified short interfering nucleic acid (siNA) molecule of the invention capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted in vitro system comprising a sense region, wherein one or more pyrimidine nucleotides present in the sense region are 2'deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and one or more purine nucleotides present in the sense region are 2'-deoxy purine nucleotides (e.g., wherein all purine nucleotides are 2'-deoxy purine nucleotides or alternately a plurality of purine nucleotides are 2'-deoxy purine nucleotides), and an antisense region, wherein one or more pyrimidine nucleotides present in the antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and one or more purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides). The sense region and/or the antisense region can have a terminal cap modification, such as any modification described herein or shown in Figure 10, that is optionally present at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of the

sense and/or antisense sequence. The sense and/or antisense region can optionally further comprise a 3'-terminal nucleotide overhang having about 1 to about 4 (e.g., about 1, 2, 3, or 4) 2'-deoxynucleotides. The overhang nucleotides can further comprise one or more (e.g., about 1, 2, 3, 4 or more) phosphorothioate, phosphonoacetate, and/or thiophosphonoacetate internucleotide linkages. Non-limiting examples of these chemically-modified siNAs are shown in Figures 4 and 5 and Tables III and IV herein. In any of these described embodiments, the purine nucleotides present in the sense region are alternatively 2'-O-methyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides) and one or more purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides). Also, in any of these embodiments, one or more purine nucleotides present in the sense region are alternatively purine ribonucleotides (e.g., wherein all purine nucleotides are purine ribonucleotides or alternately a plurality of purine nucleotides are purine ribonucleotides) and any purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides). Additionally, in any of these embodiments, one or more purine nucleotides present in the sense region and/or present in the antisense region are alternatively selected from the group consisting of 2'deoxy nucleotides, locked nucleic acid (LNA) nucleotides, 2'-methoxyethyl nucleotides, 4'-thionucleotides, and 2'-O-methyl nucleotides (e.g., wherein all purine nucleotides are selected from the group consisting of 2'-deoxy nucleotides, locked nucleic acid (LNA) nucleotides, 2'-methoxyethyl nucleotides, 4'-thionucleotides, and 2'-O-methyl nucleotides or alternately a plurality of purine nucleotides are selected from the group consisting of 2'-deoxy nucleotides, locked nucleic acid (LNA) nucleotides, 2'methoxyethyl nucleotides, 4'-thionucleotides, and 2'-O-methyl nucleotides).

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In another embodiment, any modified nucleotides present in the siNA molecules of the invention, preferably in the antisense strand of the siNA molecules of the invention, but also optionally in the sense and/or both antisense and sense strands, comprise modified nucleotides having properties or characteristics similar to naturally occurring

ribonucleotides. For example, the invention features siNA molecules including modified nucleotides having a Northern conformation (e.g., Northern pseudorotation cycle, see for example Saenger, Principles of Nucleic Acid Structure, Springer-Verlag ed., 1984). As such, chemically modified nucleotides present in the siNA molecules of the invention, preferably in the antisense strand of the siNA molecules of the invention, but also optionally in the sense and/or both antisense and sense strands, are resistant to nuclease degradation while at the same time maintaining the capacity to mediate RNAi. Nonlimiting examples of nucleotides having a northern configuration include locked nucleic acid (LNA) nucleotides (e.g., 2'-O, 4'-C-methylene-(D-ribofuranosyl) nucleotides); 2'methoxyethoxy (MOE) nucleotides: 2'-methyl-thio-ethyl, 2'-deoxy-2'-fluoro nucleotides, 2'-deoxy-2'-chloro nucleotides, 2'-azido nucleotides, and 2'-O-methyl nucleotides.

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In one embodiment, the sense strand of a double stranded siNA molecule of the invention comprises a terminal cap moiety, (see for example **Figure 10**) such as an inverted deoxyabaisc moiety, at the 3'-end, 5'-end, or both 3' and 5'-ends of the sense strand.

In one embodiment, the invention features a chemically-modified short interfering nucleic acid molecule (siNA) capable of mediating RNA interference (RNAi) against SARS inside a cell or reconstituted *in vitro* system, wherein the chemical modification comprises a conjugate covalently attached to the chemically-modified siNA molecule. Non-limiting examples of conjugates contemplated by the invention include conjugates and ligands described in Vargeese *et al.*, USSN 10/427,160, filed April 30, 2003, incorporated by reference herein in its entirety, including the drawings. In another embodiment, the conjugate is covalently attached to the chemically-modified siNA molecule via a biodegradable linker. In one embodiment, the conjugate molecule is attached at the 3'-end of either the sense strand, the antisense strand, or both strands of the chemically-modified siNA molecule. In another embodiment, the conjugate molecule is attached at the 5'-end of either the sense strand, the antisense strand, or both strands of the chemically-modified siNA molecule. In yet another embodiment, the conjugate molecule is attached both the 3'-end and 5'-end of either the sense strand, the antisense strand, or both strands of the chemically-modified siNA molecule. Or any

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combination thereof. In one embodiment, a conjugate molecule of the invention comprises a molecule that facilitates delivery of a chemically-modified siNA molecule into a biological system, such as a cell. In another embodiment, the conjugate molecule attached to the chemically-modified siNA molecule is a polyethylene glycol, human serum albumin, or a ligand for a cellular receptor that can mediate cellular uptake. Examples of specific conjugate molecules contemplated by the instant invention that can be attached to chemically-modified siNA molecules are described in Vargeese *et al.*, U.S. Serial No. 10/201,394, filed July 22, 2002, incorporated by reference herein. The type of conjugates used and the extent of conjugation of siNA molecules of the invention can be evaluated for improved pharmacokinetic profiles, bioavailability, and/or stability of siNA constructs while at the same time maintaining the ability of the siNA to mediate RNAi activity. As such, one skilled in the art can screen siNA constructs that are modified with various conjugates to determine whether the siNA conjugate complex possesses improved properties while maintaining the ability to mediate RNAi, for example in animal models as are generally known in the art.

In one embodiment, the invention features a short interfering nucleic acid (siNA) molecule of the invention, wherein the siNA further comprises a nucleotide, nonnucleotide, or mixed nucleotide/non-nucleotide linker that joins the sense region of the siNA to the antisense region of the siNA. In one embodiment, a nucleotide linker of the invention can be a linker of ≥ 2 nucleotides in length, for example about 3, 4, 5, 6, 7, 8, 9, or 10 nucleotides in length. In another embodiment, the nucleotide linker can be a nucleic acid aptamer. By "aptamer" or "nucleic acid aptamer" as used herein is meant a nucleic acid molecule that binds specifically to a target molecule wherein the nucleic acid molecule has sequence that comprises a sequence recognized by the target molecule in its natural setting. Alternately, an aptamer can be a nucleic acid molecule that binds to a target molecule where the target molecule does not naturally bind to a nucleic acid. The target molecule can be any molecule of interest. For example, the aptamer can be used to bind to a ligand-binding domain of a protein, thereby preventing interaction of the naturally occurring ligand with the protein. This is a non-limiting example and those in the art will recognize that other embodiments can be readily generated using techniques generally known in the art. (See, for example, Gold et al., 1995, Annu. Rev. Biochem., 64, 763; Brody and Gold, 2000, J. Biotechnol., 74, 5; Sun, 2000, Curr. Opin.

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Mol. Ther., 2, 100; Kusser, 2000, J. Biotechnol., 74, 27; Hermann and Patel, 2000, Science, 287, 820; and Jayasena, 1999, Clinical Chemistry, 45, 1628.)

In yet another embodiment, a non-nucleotide linker of the invention comprises abasic nucleotide, polyether, polyamine, polyamide, peptide, carbohydrate, lipid, polyhydrocarbon, or other polymeric compounds (e.g. polyethylene glycols such as those having between 2 and 100 ethylene glycol units). Specific examples include those described by Seela and Kaiser, Nucleic Acids Res. 1990, 18:6353 and Nucleic Acids Res. 1987, 15:3113; Cload and Schepartz, J. Am. Chem. Soc. 1991, 113:6324; Richardson and Schepartz, J. Am. Chem. Soc. 1991, 113:5109; Ma et al., Nucleic Acids Res. 1993, 21:2585 and Biochemistry 1993, 32:1751; Durand et al., Nucleic Acids Res. 1990, 18:6353; McCurdy et al., Nucleosides & Nucleotides 1991, 10:287; Jschke et al., Tetrahedron Lett. 1993, 34:301; Ono et al., Biochemistry 1991, 30:9914; Arnold et al., International Publication No. WO 89/02439; Usman et al., International Publication No. WO 95/06731; Dudycz et al., International Publication No. WO 95/11910 and Ferentz and Verdine, J. Am. Chem. Soc. 1991, 113:4000, all hereby incorporated by reference herein. A "non-nucleotide" further means any group or compound that can be incorporated into a nucleic acid chain in the place of one or more nucleotide units, including either sugar and/or phosphate substitutions, and allows the remaining bases to exhibit their enzymatic activity. The group or compound can be abasic in that it does not contain a commonly recognized nucleotide base, such as adenosine, guanine, cytosine, uracil or thymine, for example at the C1 position of the sugar.

In one embodiment, the invention features a short interfering nucleic acid (siNA) molecule capable of mediating RNA interference (RNAi) inside a cell or reconstituted *in vitro* system, wherein one or both strands of the siNA molecule that are assembled from two separate oligonucleotides do not comprise any ribonucleotides. For example, a siNA molecule can be assembled from a single oligonculeotide where the sense and antisense regions of the siNA comprise separate oligonucleotides not having any ribonucleotides (e.g., nucleotides having a 2'-OH group) present in the oligonucleotides. In another example, a siNA molecule can be assembled from a single oligonculeotide where the sense and antisense regions of the siNA are linked or circularized by a nucleotide or non-nucleotide linker as desreibed herein, wherein the oligonucleotide does not have any

ribonucleotides (e.g., nucleotides having a 2'-OH group) present in the oligonucleotide. Applicant has surprisingly found that the presense of ribonucleotides (e.g., nucleotides having a 2'-hydroxyl group) within the siNA molecule is not required or essential to support RNAi activity. As such, in one embodiment, all positions within the siNA can include chemically modified nucleotides and/or non-nucleotides such as nucleotides and or non-nucleotides having Formula I, II, III, IV, V, VI, or VII or any combination thereof to the extent that the ability of the siNA molecule to support RNAi activity in a cell is maintained.

In one embodiment, a siNA molecule of the invention is a single stranded siNA molecule that mediates RNAi activity in a cell or reconstituted in vitro system comprising a single stranded polynucleotide having complementarity to a target nucleic acid sequence. In another embodiment, the single stranded siNA molecule of the invention comprises a 5'-terminal phosphate group. In another embodiment, the single stranded siNA molecule of the invention comprises a 5'-terminal phosphate group and a 3'-terminal phosphate group (e.g., a 2',3'-cyclic phosphate). In another embodiment, the single stranded siNA molecule of the invention comprises about 19 to about 29 (e.g., about 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, or 29) nucleotides. In yet another embodiment, the single stranded siNA molecule of the invention comprises one or more chemically modified nucleotides or non-nucleotides described herein. For example, all the positions within the siNA molecule can include chemically-modified nucleotides such as nucleotides having any of Formulae I-VII, or any combination thereof to the extent that the ability of the siNA molecule to support RNAi activity in a cell is maintained.

In one embodiment, a siNA molecule of the invention is a single stranded siNA molecule that mediates RNAi activity in a cell or reconstituted in vitro system comprising a single stranded polynucleotide having complementarity to a target nucleic acid sequence, wherein one or more pyrimidine nucleotides present in the siNA are 2'-deoxy-2'-fluoro pyrimidine nucleotides (e.g., wherein all pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides or alternately a plurality of pyrimidine nucleotides are 2'-deoxy-2'-fluoro pyrimidine nucleotides), and wherein any purine nucleotides present in the antisense region are 2'-O-methyl purine nucleotides (e.g., wherein all purine

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nucleotides are 2'-O-methyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-O-methyl purine nucleotides), and a terminal cap modification, such as any modification described herein or shown in Figure 10, that is optionally present at the 3'-end, the 5'-end, or both of the 3' and 5'-ends of the antisense sequence. The siNA optionally further comprises about 1 to about 4 or more (e.g., about 1, 2, 3, 4 or more) terminal 2'-deoxynucleotides at the 3'-end of the siNA molecule, wherein the terminal nucleotides can further comprise one or more (e.g., 1, 2, 3, 4 or more) phosphorothioate, phosphonoacetate, and/or thiophosphonoacetate internucleotide linkages, and wherein the siNA optionally further comprises a terminal phosphate group, such as a 5'-terminal phosphate group. In any of these embodiments, any purine nucleotides present in the antisense region are alternatively 2'-deoxy purine nucleotides (e.g., wherein all purine nucleotides are 2'-deoxy purine nucleotides or alternately a plurality of purine nucleotides are 2'-deoxy purine nucleotides). Also, in any of these embodiments, any purine nucleotides present in the siNA (i.e., purine nucleotides present in the sense and/or antisense region) can alternatively be locked nucleic acid (LNA) nucleotides (e.g., wherein all purine nucleotides are LNA nucleotides or alternately a plurality of purine nucleotides are LNA nucleotides). Also, in any of these embodiments, any purine nucleotides present in the siNA are alternatively 2'-methoxyethyl purine nucleotides (e.g., wherein all purine nucleotides are 2'-methoxyethyl purine nucleotides or alternately a plurality of purine nucleotides are 2'-methoxyethyl purine nucleotides). In another embodiment, any modified nucleotides present in the single stranded siNA molecules of the invention comprise modified nucleotides having properties or characteristics similar to naturally occurring ribonucleotides. For example, the invention features siNA molecules including modified nucleotides having a Northern conformation (e.g., Northern pseudorotation cycle, see for example Saenger, Principles of Nucleic Acid Structure, Springer-Verlag ed., 1984). As such, chemically modified nucleotides present in the single stranded siNA molecules of the invention are preferably resistant to nuclease degradation while at the same time maintaining the capacity to mediate RNAi.

In one embodiment, the invention features a method for modulating the expression of a SARS gene within a cell comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS gene; and (b) introducing the

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siNA molecule into a cell under conditions suitable to modulate the expression of the SARS gene in the cell.

In one embodiment, the invention features a method for modulating the expression of a SARS gene within a cell comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS gene and wherein the sense strand sequence of the siNA comprises a sequence identical or substantially similar to the sequence of the target RNA; and (b) introducing the siNA molecule into a cell under conditions suitable to modulate the expression of the SARS gene in the cell.

In another embodiment, the invention features a method for modulating the expression of more than one SARS gene within a cell comprising: (a) synthesizing siNA molecules of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS genes; and (b) introducing the siNA molecules into a cell under conditions suitable to modulate the expression of the SARS genes in the cell.

In another embodiment, the invention features a method for modulating the expression of two or more SARS genes within a cell comprising: (a) synthesizing one or more siNA molecules of the invention, which can be chemically-modified, wherein the siNA strands comprise sequences complementary to RNA of the SARS genes and wherein the sense strand sequences of the siNAs comprise sequences identical or substantially similar to the sequences of the target RNAs; and (b) introducing the siNA molecules into a cell under conditions suitable to modulate the expression of the SARS genes in the cell.

In another embodiment, the invention features a method for modulating the expression of more than one SARS gene within a cell comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS gene and wherein the sense strand sequence of the siNA comprises a sequence identical or substantially similar to the sequences of the target RNAs; and (b) introducing the siNA

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molecule into a cell under conditions suitable to modulate the expression of the SARS genes in the cell.

In one embodiment, siNA molecules of the invention are used as reagents in ex vivo applications. For example, siNA reagents are intoduced into tissue or cells that are transplanted into a subject for therapeutic effect. The cells and/or tissue can be derived from an organism or subject that later receives the explant, or can be derived from another organism or subject prior to transplantation. The siNA molecules can be used to modulate the expression of one or more genes in the cells or tissue, such that the cells or tissue obtain a desired phenotype or are able to perform a function when transplanted in vivo. In one embodiment, certain target cells from a patient are extracted. These extracted cells are contacted with siNAs targeteing a specific nucleotide sequence within the cells under conditions suitable for uptake of the siNAs by these cells (e.g. using delivery reagents such as cationic lipids, liposomes and the like or using techniques such as electroporation to facilitate the delivery of siNAs into cells). The cells are then reintroduced back into the same patient or other patients. In one embodiment, the invention features a method of modulating the expression of a SARS gene in a tissue explant comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS gene; and (b) introducing the siNA molecule into a cell of the tissue explant derived from a particular organism under conditions suitable to modulate the expression of the SARS gene in the tissue explant. In another embodiment, the method further comprises introducing the tissue explant back into the organism the tissue was derived from or into another organism under conditions suitable to modulate the expression of the SARS gene in that organism.

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In one embodiment, the invention features a method of modulating the expression of a SARS gene in a tissue explant comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS gene and wherein the sense strand sequence of the siNA comprises a sequence identical or substantially similar to the sequence of the target RNA; and (b) introducing the siNA molecule into a cell of the tissue explant derived from a particular organism under conditions suitable to modulate

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the expression of the SARS gene in the tissue explant. In another embodiment, the method further comprises introducing the tissue explant back into the organism the tissue was derived from or into another organism under conditions suitable to modulate the expression of the SARS gene in that organism.

In another embodiment, the invention features a method of modulating the expression of more than one SARS gene in a tissue explant comprising: (a) synthesizing siNA molecules of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS genes; and (b) introducing the siNA molecules into a cell of the tissue explant derived from a particular organism under conditions suitable to modulate the expression of the SARS genes in the tissue explant. In another embodiment, the method further comprises introducing the tissue explant back into the organism the tissue was derived from or into another organism under conditions suitable to modulate the expression of the SARS genes in that organism.

In one embodiment, the invention features a method of modulating the expression of a SARS gene in an organism comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS gene; and (b) introducing the siNA molecule into the organism under conditions suitable to modulate the expression of the SARS gene in the organism. The level of SARS protein or RNA can be determined as is known in the art.

In another embodiment, the invention features a method of modulating the expression of more than one SARS gene in an organism comprising: (a) synthesizing siNA molecules of the invention, which can be chemically-modified, wherein one of the siNA strands comprises a sequence complementary to RNA of the SARS genes; and (b) introducing the siNA molecules into the organism under conditions suitable to modulate the expression of the SARS genes in the organism. The level of SARS protein or RNA can be determined as is known in the art.

In one embodiment, the invention features a method for modulating the expression of a SARS gene within a cell comprising: (a) synthesizing a siNA molecule of the

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invention, which can be chemically-modified, wherein the siNA comprises a single stranded sequence having complementarity to RNA of the SARS gene; and (b) introducing the siNA molecule into a cell under conditions suitable to modulate the expression of the SARS gene in the cell.

In another embodiment, the invention features a method for modulating the expression of more than one SARS gene within a cell comprising: (a) synthesizing siNA molecules of the invention, which can be chemically-modified, wherein the siNA comprises a single stranded sequence having complementarity to RNA of the SARS gene; and (b) contacting the cell in vitro or in vivo with the siNA molecule under conditions suitable to modulate the expression of the SARS genes in the cell.

In one embodiment, the invention features a method of modulating the expression of a SARS gene in a tissue explant comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein the siNA comprises a single stranded sequence having complementarity to RNA of the SARS gene; and (b) contacting the cell of the tissue explant derived from a particular organism with the siNA molecule under conditions suitable to modulate the expression of the SARS gene in the tissue explant. In another embodiment, the method further comprises introducing the tissue explant back into the organism the tissue was derived from or into another organism under conditions suitable to modulate the expression of the SARS gene in that organism.

In another embodiment, the invention features a method of modulating the expression of more than one SARS gene in a tissue explant comprising: (a) synthesizing siNA molecules of the invention, which can be chemically-modified, wherein the siNA comprises a single stranded sequence having complementarity to RNA of the SARS gene; and (b) introducing the siNA molecules into a cell of the tissue explant derived from a particular organism under conditions suitable to modulate the expression of the SARS genes in the tissue explant. In another embodiment, the method further comprises introducing the tissue explant back into the organism the tissue was derived from or into another organism under conditions suitable to modulate the expression of the SARS genes in that organism.

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In one embodiment, the invention features a method of modulating the expression of a SARS gene in an organism comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein the siNA comprises a single stranded sequence having complementarity to RNA of the SARS gene; and (b) introducing the siNA molecule into the organism under conditions suitable to modulate the expression of the SARS gene in the organism.

In another embodiment, the invention features a method of modulating the expression of more than one SARS gene in an organism comprising: (a) synthesizing siNA molecules of the invention, which can be chemically-modified, wherein the siNA comprises a single stranded sequence having complementarity to RNA of the SARS gene; and (b) introducing the siNA molecules into the organism under conditions suitable to modulate the expression of the SARS genes in the organism.

In one embodiment, the invention features a method of modulating the expression of a SARS gene in an organism comprising contacting the organism with a siNA molecule of the invention under conditions suitable to modulate the expression of the SARS gene in the organism.

In another embodiment, the invention features a method of modulating the expression of more than one SARS gene in an organism comprising contacting the organism with one or more siNA molecules of the invention under conditions suitable to modulate the expression of the SARS genes in the organism.

The siNA molecules of the invention can be designed to down regulate or inhibit target (e.g., SARS) gene expression through RNAi targeting of a variety of RNA molecules. In one embodiment, the siNA molecules of the invention are used to target various RNAs corresponding to a target gene. Non-limiting examples of such RNAs include messenger RNA (mRNA), alternate RNA splice variants of target gene(s), post-transcriptionally modified RNA of target gene(s), pre-mRNA of target gene(s), and/or RNA templates. If alternate splicing produces a family of transcripts that are distinguished by usage of appropriate exons, the instant invention can be used to inhibit gene expression through the appropriate exons to specifically inhibit or to distinguish among the functions of gene family members. For example, a protein that contains an

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alternatively spliced transmembrane domain can be expressed in both membrane bound and secreted forms. Use of the invention to target the exon containing the transmembrane domain can be used to determine the functional consequences of pharmaceutical targeting of membrane bound as opposed to the secreted form of the protein. Non-limiting examples of applications of the invention relating to targeting these RNA molecules include therapeutic pharmaceutical applications, pharmaceutical discovery applications, molecular diagnostic and gene function applications, and gene mapping, for example using single nucleotide polymorphism mapping with siNA molecules of the invention. Such applications can be implemented using known gene sequences or from partial sequences available from an expressed sequence tag (EST).

In another embodiment, the siNA molecules of the invention are used to target conserved sequences corresponding to a gene family or gene families such as SARS family genes. As such, siNA molecules targeting multiple SARS targets can provide increased therapeutic effect. In addition, siNA can be used to characterize pathways of gene function in a variety of applications. For example, the present invention can be used to inhibit the activity of target gene(s) in a pathway to determine the function of uncharacterized gene(s) in gene function analysis, mRNA function analysis, or translational analysis. The invention can be used to determine potential target gene pathways involved in various diseases and conditions toward pharmaceutical development. The invention can be used to understand pathways of gene expression involved in, for example, the progression and/or maintenance of SARS virus infection, acute respiratory failure, viral pneumonia, and other indications that can respond to the level of SARS in a cell or tissue.

In one embodiment, siNA molecule(s) and/or methods of the invention are used to down regulate the expression of gene(s) that encode RNA referred to by Genbank Accession, for example SARS genes encoding RNA sequence(s) referred to herein by Genbank Accession number, for example, Genbank Accession Nos. shown in **Table I**.

In one embodiment, the invention features a method comprising: (a) generating a library of siNA constructs having a predetermined complexity; and (b) assaying the siNA constructs of (a) above, under conditions suitable to determine RNAi target sites within the target RNA sequence. In one embodiment, the siNA molecules of (a) have strands of

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a fixed length, for example, about 23 nucleotides in length. In another embodiment, the siNA molecules of (a) are of differing length, for example having strands of about 19 to about 25 (e.g., about 19, 20, 21, 22, 23, 24, or 25) nucleotides in length. In one embodiment, the assay can comprise a reconstituted *in vitro* siNA assay as described herein. In another embodiment, the assay can comprise a cell culture system in which target RNA is expressed. In another embodiment, fragments of target RNA are analyzed for detectable levels of cleavage, for example by gel electrophoresis, northern blot analysis, or RNAse protection assays, to determine the most suitable target site(s) within the target RNA sequence. The target RNA sequence can be obtained as is known in the art, for example, by cloning and/or transcription for *in vitro* systems, and by cellular expression in *in vivo* systems.

In one embodiment, the invention features a method comprising: (a) generating a randomized library of siNA constructs having a predetermined complexity, such as of 4^N, where N represents the number of base paired nucleotides in each of the siNA construct strands (eg. for a siNA construct having 21 nucleotide sense and antisense strands with 19 base pairs, the complexity would be 419); and (b) assaying the siNA constructs of (a) above, under conditions suitable to determine RNAi target sites within the target SARS RNA sequence. In another embodiment, the siNA molecules of (a) have strands of a fixed length, for example about 23 nucleotides in length. In yet another embodiment, the siNA molecules of (a) are of differing length, for example having strands of about 19 to about 25 (e.g., about 19, 20, 21, 22, 23, 24, or 25) nucleotides in length. In one embodiment, the assay can comprise a reconstituted in vitro siNA assay as described in Example 7 herein. In another embodiment, the assay can comprise a cell culture system in which target RNA is expressed. In another embodiment, fragments of SARS RNA are analyzed for detectable levels of cleavage, for example by gel electrophoresis, northern blot analysis, or RNAse protection assays, to determine the most suitable target site(s) within the target SARS RNA sequence. The target SARS RNA sequence can be obtained as is known in the art, for example, by cloning and/or transcription for in vitro systems, and by cellular expression in in vivo systems.

In another embodiment, the invention features a method comprising: (a) analyzing the sequence of a RNA target encoded by a target gene; (b) synthesizing one or more sets

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of siNA molecules having sequence complementary to one or more regions of the RNA of (a); and (c) assaying the siNA molecules of (b) under conditions suitable to determine RNAi targets within the target RNA sequence. In one embodiment, the siNA molecules of (b) have strands of a fixed length, for example about 23 nucleotides in length. In another embodiment, the siNA molecules of (b) are of differing length, for example having strands of about 19 to about 25 (e.g., about 19, 20, 21, 22, 23, 24, or 25) nucleotides in length. In one embodiment, the assay can comprise a reconstituted in vitro siNA assay as described herein. In another embodiment, the assay can comprise a cell culture system in which target RNA is expressed. Fragments of target RNA are analyzed for detectable levels of cleavage, for example by gel electrophoresis, northern blot analysis, or RNAse protection assays, to determine the most suitable target site(s) within the target RNA sequence. The target RNA sequence can be obtained as is known in the art, for example, by cloning and/or transcription for in vitro systems, and by expression in in vivo systems.

By "target site" is meant a sequence within a target RNA that is "targeted" for cleavage mediated by a siNA construct which contains sequences within its antisense region that are complementary to the target sequence.

By "detectable level of cleavage" is meant cleavage of target RNA (and formation of cleaved product RNAs) to an extent sufficient to discern cleavage products above the background of RNAs produced by random degradation of the target RNA. Production of cleavage products from 1-5% of the target RNA is sufficient to detect above the background for most methods of detection.

In one embodiment, the invention features a composition comprising a siNA molecule of the invention, which can be chemically-modified, in a pharmaceutically acceptable carrier or diluent. In another embodiment, the invention features a pharmaceutical composition comprising siNA molecules of the invention, which can be chemically-modified, targeting one or more genes in a pharmaceutically acceptable carrier or diluent. In another embodiment, the invention features a method for diagnosing a disease or condition in a subject comprising administering to the subject a composition of the invention under conditions suitable for the diagnosis of the disease or condition in the subject. In another embodiment, the invention features a method for

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treating or preventing a disease or condition in a subject, comprising administering to the subject a composition of the invention under conditions suitable for the treatment or prevention of the disease or condition in the subject, alone or in conjunction with one or more other therapeutic compounds. In yet another embodiment, the invention features a method for reducing or preventing tissue rejection in a subject comprising administering to the subject a composition of the invention under conditions suitable for the reduction or prevention of tissue rejection in the subject.

In another embodiment, the invention features a method for validating a SARS gene target, comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands includes a sequence complementary to RNA of a SARS target gene; (b) introducing the siNA molecule into a cell, tissue, or organism under conditions suitable for modulating expression of the SARS target gene in the cell, tissue, or organism; and (c) determining the function of the gene by assaying for any phenotypic change in the cell, tissue, or organism.

In another embodiment, the invention features a method for validating a SARS target comprising: (a) synthesizing a siNA molecule of the invention, which can be chemically-modified, wherein one of the siNA strands includes a sequence complementary to RNA of a SARS target gene; (b) introducing the siNA molecule into a biological system under conditions suitable for modulating expression of the SARS target gene in the biological system; and (c) determining the function of the gene by assaying for any phenotypic change in the biological system.

By "biological system" is meant, material, in a purified or unpurified form, from biological sources, including but not limited to human or animal, wherein the system comprises the components required for RNAi acitivity. The term "biological system" includes, for example, a cell, tissue, or organism, or extract thereof. The term biological system also includes reconstituted RNAi systems that can be used in an *in vitro* setting.

By "phenotypic change" is meant any detectable change to a cell that occurs in response to contact or treatment with a nucleic acid molecule of the invention (e.g., siNA). Such detectable changes include, but are not limited to, changes in shape, size, proliferation, motility, protein expression or RNA expression or other physical or

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chemical changes as can be assayed by methods known in the art. The detectable change can also include expression of reporter genes/molecules such as Green Florescent Protein (GFP) or various tags that are used to identify an expressed protein or any other cellular component that can be assayed.

In one embodiment, the invention features a kit containing a siNA molecule of the invention, which can be chemically-modified, that can be used to modulate the expression of a SARS target gene in a biological system, including, for example, in a cell, tissue, or organism. In another embodiment, the invention features a kit containing more than one siNA molecule of the invention, which can be chemically-modified, that can be used to modulate the expression of more than one SARS target gene in a biological system, including, for example, in a cell, tissue, or organism.

In one embodiment, the invention features a cell containing one or more siNA molecules of the invention, which can be chemically-modified. In another embodiment, the cell containing a siNA molecule of the invention is a mammalian cell. In yet another embodiment, the cell containing a siNA molecule of the invention is a human cell.

In one embodiment, the synthesis of a siNA molecule of the invention, which can be chemically-modified, comprises: (a) synthesis of two complementary strands of the siNA molecule; (b) annealing the two complementary strands together under conditions suitable to obtain a double-stranded siNA molecule. In another embodiment, synthesis of the two complementary strands of the siNA molecule is by solid phase oligonucleotide synthesis. In yet another embodiment, synthesis of the two complementary strands of the siNA molecule is by solid phase tandem oligonucleotide synthesis.

In one embodiment, the invention features a method for synthesizing a siNA duplex molecule comprising: (a) synthesizing a first oligonucleotide sequence strand of the siNA molecule, wherein the first oligonucleotide sequence strand comprises a cleavable linker molecule that can be used as a scaffold for the synthesis of the second oligonucleotide sequence strand of the siNA; (b) synthesizing the second oligonucleotide sequence strand of siNA on the scaffold of the first oligonucleotide sequence strand, wherein the second oligonucleotide sequence strand further comprises a chemical moiety than can be used to purify the siNA duplex; (c) cleaving the linker molecule of (a) under

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conditions suitable for the two siNA oligonucleotide strands to hybridize and form a stable duplex; and (d) purifying the siNA duplex utilizing the chemical moiety of the second oligonucleotide sequence strand. In one embodiment, cleavage of the linker molecule in (c) above takes place during deprotection of the oligonucleotide, for example under hydrolysis conditions using an alkylamine base such as methylamine. In one embodiment, the method of synthesis comprises solid phase synthesis on a solid support such as controlled pore glass (CPG) or polystyrene, wherein the first sequence of (a) is synthesized on a cleavable linker, such as a succinyl linker, using the solid support as a scaffold. The cleavable linker in (a) used as a scaffold for synthesizing the second strand can comprise similar reactivity as the solid support derivatized linker, such that cleavage of the solid support derivatized linker and the cleavable linker of (a) takes place concomitantly. In another embodiment, the chemical moiety of (b) that can be used to isolate the attached oligonucleotide sequence comprises a trityl group, for example a dimethoxytrityl group, which can be employed in a trityl-on synthesis strategy as described herein. In yet another embodiment, the chemical moiety, such as a dimethoxytrityl group, is removed during purification, for example, using acidic conditions.

In a further embodiment, the method for siNA synthesis is a solution phase synthesis or hybrid phase synthesis wherein both strands of the siNA duplex are synthesized in tandem using a cleavable linker attached to the first sequence which acts a scaffold for synthesis of the second sequence. Cleavage of the linker under conditions suitable for hybridization of the separate siNA sequence strands results in formation of the double-stranded siNA molecule.

In another embodiment, the invention features a method for synthesizing a siNA duplex molecule comprising: (a) synthesizing one oligonucleotide sequence strand of the siNA molecule, wherein the sequence comprises a cleavable linker molecule that can be used as a scaffold for the synthesis of another oligonucleotide sequence; (b) synthesizing a second oligonucleotide sequence having complementarity to the first sequence strand on the scaffold of (a), wherein the second sequence comprises the other strand of the double-stranded siNA molecule and wherein the second sequence further comprises a chemical moiety than can be used to isolate the attached oligonucleotide

sequence; (c) purifying the product of (b) utilizing the chemical moiety of the second oligonucleotide sequence strand under conditions suitable for isolating the full-length sequence comprising both siNA oligonucleotide strands connected by the cleavable linker and under conditions suitable for the two siNA oligonucleotide strands to hybridize and form a stable duplex. In one embodiment, cleavage of the linker molecule in (c) above takes place during deprotection of the oligonucleotide, for example under hydrolysis conditions. In another embodiment, cleavage of the linker molecule in (c) above takes place after deprotection of the oligonucleotide. In another embodiment, the method of synthesis comprises solid phase synthesis on a solid support such as controlled pore glass (CPG) or polystyrene, wherein the first sequence of (a) is synthesized on a cleavable linker, such as a succinyl linker, using the solid support as a scaffold. The cleavable linker in (a) used as a scaffold for synthesizing the second strand can comprise similar reactivity or differing reactivity as the solid support derivatized linker, such that cleavage of the solid support derivatized linker and the cleavable linker of (a) takes place either concomitantly or sequentially. In one embodiment, the chemical moiety of (b) that can be used to isolate the attached oligonucleotide sequence comprises a trityl group, for example a dimethoxytrityl group.

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In another embodiment, the invention features a method for making a double-stranded siNA molecule in a single synthetic process comprising: (a) synthesizing an oligonucleotide having a first and a second sequence, wherein the first sequence is complementary to the second sequence, and the first oligonucleotide sequence is linked to the second sequence via a cleavable linker, and wherein a terminal 5'-protecting group, for example, a 5'-O-dimethoxytrityl group (5'-O-DMT) remains on the oligonucleotide having the second sequence; (b) deprotecting the oligonucleotide whereby the deprotection results in the cleavage of the linker joining the two oligonucleotide sequences; and (c) purifying the product of (b) under conditions suitable for isolating the double-stranded siNA molecule, for example using a trityl-on synthesis strategy as described herein.

In another embodiment, the method of synthesis of siNA molecules of the invention comprises the teachings of Scaringe *et al.*, US Patent Nos. 5,889,136; 6,008,400; and 6,111,086, incorporated by reference herein in their entirety.

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In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications, for example, one or more chemical modifications having any of Formulae I-VII or any combination thereof that increases the nuclease resistance of the siNA construct.

In another embodiment, the invention features a method for generating siNA molecules with increased nuclease resistance comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having increased nuclease resistance.

In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications described herein that modulates the binding affinity between the sense and antisense strands of the siNA construct.

In another embodiment, the invention features a method for generating siNA molecules with increased binding affinity between the sense and antisense strands of the siNA molecule comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having increased binding affinity between the sense and antisense strands of the siNA molecule.

In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications described herein that modulates the binding affinity between the antisense strand of the siNA construct and a complementary target RNA sequence within a cell.

In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications described herein that modulates the binding affinity between the antisense strand of the siNA construct and a complementary target DNA sequence within a cell.

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In another embodiment, the invention features a method for generating siNA molecules with increased binding affinity between the antisense strand of the siNA molecule and a complementary target RNA sequence comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having increased binding affinity between the antisense strand of the siNA molecule and a complementary target RNA sequence.

In another embodiment, the invention features a method for generating siNA molecules with increased binding affinity between the antisense strand of the siNA molecule and a complementary target DNA sequence comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having increased binding affinity between the antisense strand of the siNA molecule and a complementary target DNA sequence.

In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications described herein that modulate the polymerase activity of a cellular polymerase capable of generating additional endogenous siNA molecules having sequence homology to the chemically-modified siNA construct.

In another embodiment, the invention features a method for generating siNA molecules capable of mediating increased polymerase activity of a cellular polymerase capable of generating additional endogenous siNA molecules having sequence homology to a chemically-modified siNA molecule comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules capable of mediating increased polymerase activity of a cellular polymerase capable of generating additional endogenous siNA molecules having sequence homology to the chemically-modified siNA molecule.

In one embodiment, the invention features chemically-modified siNA constructs that mediate RNAi against SARS in a cell, wherein the chemical modifications do not

significantly effect the interaction of siNA with a target RNA molecule, DNA molecule and/or proteins or other factors that are essential for RNAi in a manner that would decrease the efficacy of RNAi mediated by such siNA constructs.

In another embodiment, the invention features a method for generating siNA molecules with improved RNAi activity against SARS comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved RNAi activity.

In yet another embodiment, the invention features a method for generating siNA molecules with improved RNAi activity against SARS target RNA comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved RNAi activity against the target RNA.

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In yet another embodiment, the invention features a method for generating siNA molecules with improved RNAi activity against SARS target DNA comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved RNAi activity against the target DNA.

In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications described herein that modulates the cellular uptake of the siNA construct.

In another embodiment, the invention features a method for generating siNA molecules against SARS with improved cellular uptake comprising (a) introducing nucleotides having any of Formula I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved cellular uptake.

In one embodiment, the invention features siNA constructs that mediate RNAi against SARS, wherein the siNA construct comprises one or more chemical modifications described herein that increases the bioavailability of the siNA construct,

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for example, by attaching polymeric conjugates such as polyethyleneglycol or equivalent conjugates that improve the pharmacokinetics of the siNA construct, or by attaching conjugates that target specific tissue types or cell types *in vivo*. Non-limiting examples of such conjugates are described in Vargeese *et al.*, U.S. Serial No. 10/201,394 incorporated by reference herein.

In one embodiment, the invention features a method for generating siNA molecules of the invention with improved bioavailability, comprising (a) introducing a conjugate into the structure of a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved bioavailability. Such conjugates can include ligands for cellular receptors, such as peptides derived from naturally occurring protein ligands; protein localization sequences, including cellular ZIP code sequences; antibodies; nucleic acid aptamers; vitamins and other co-factors, such as folate and N-acetylgalactosamine; polymers, such as polyethyleneglycol (PEG); phospholipids; cholesterol; polyamines, such as spermine or spermidine; and others.

In one embodiment, the invention features a double stranded short interfering nucleic acid (siNA) molecule that comprises a first nucleotide sequence complementary to a target RNA sequence or a portion thereof, and a second sequence having complementarity to said first sequence, wherein said second sequence is chemically modified in a manner that it can no longer act as a guide sequence for efficiently mediating RNA interference and/or be recognized by cellular proteins that facilitate RNAi.

In one embodiment, the invention features a double stranded short interfering nucleic acid (siNA) molecule that comprises a first nucleotide sequence complementary to a target RNA sequence or a portion thereof, and a second sequence having complementarity to said first sequence, wherein the second sequence is designed or modified in a manner that prevents its entry into the RNAi pathway as a guide sequence or as a sequence that is complementary to a target nucleic acid (e.g., RNA) sequence. Such design or modifications are expected to enhance the activity of siNA and/or improve the specificity of siNA molecules of the invention. These modifications are also expected to minimize any off-target effects and/or associated toxicity.

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In one embodiment, the invention features a double stranded short interfering nucleic acid (siNA) molecule that comprises a first nucleotide sequence complementary to a target RNA sequence or a portion thereof, and a second sequence having complementarity to said first sequence, wherein said second sequence is incapable of acting as a guide sequence for mediating RNA interference.

In one embodiment, the invention features a double stranded short interfering nucleic acid (siNA) molecule that comprises a first nucleotide sequence complementary to a target RNA sequence or a portion thereof, and a second sequence having complementarity to said first sequence, wherein said second sequence does not have a terminal 5'-hydroxyl (5'-OH) or 5'-phosphate group.

In one embodiment, the invention features a double stranded short interfering nucleic acid (siNA) molecule that comprises a first nucleotide sequence complementary to a target RNA sequence or a portion thereof, and a second sequence having complementarity to said first sequence, wherein said second sequence comprises a terminal cap moiety at the 5'-end of said second sequence. In one embodiment, the terminal cap moiety comprises an inverted abasic, inverted deoxy abasic, inverted nucleotide moiety, a group shown in **Figure 10**, an alkyl or cycloalkyl group, a heterocycle, or any other group that prevents RNAi activity in which the second sequence serves as a guide sequence or template for RNAi.

In one embodiment, the invention features a double stranded short interfering nucleic acid (siNA) molecule that comprises a first nucleotide sequence complementary to a target RNA sequence or a portion thereof, and a second sequence having complementarity to said first sequence, wherein said second sequence comprises a terminal cap moiety at the 5'-end and 3'-end of said second sequence. In one embodiment, each terminal cap moiety individually comprises an inverted abasic, inverted deoxy abasic, inverted nucleotide moiety, a group shown in **Figure 10**, an alkyl or cycloalkyl group, a heterocycle, or any other group that prevents RNAi activity in which the second sequence serves as a guide sequence or template for RNAi.

In one embodiment, the invention features a method for generating siNA molecules of the invention with improved specificity for down regulating or inhibiting

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the expression of a target nucleic acid (e.g., a DNA or RNA such as a gene or its corresponding RNA), comprising (a) introducing one or more chemical modifications into the structure of a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved specificity. In another embodiment, the chemical modification used to improve specificity comprises terminal cap modifications at the 5'-end, 3'-end, or both 5' and 3'-ends of the siNA molecule. The terminal cap modifications can comprise, for example, structures shown in Figure 10 (e.g. inverted deoxyabasic moieties) or any other chemical modification that renders a portion of the siNA molecule (e.g. the sense strand) incapable of mediating RNA interference against an off target nucleic acid sequence. In a non-limiting example, a siNA molecule is designed such that only the antisense sequence of the siNA molecule can serve as a guide sequence for RISC mediated degradation of a corresponding target RNA sequence. This can be accomplished by rendering the sense sequence of the siNA inactive by introducing chemical modifications to the sense strand that preclude recognition of the sense strand as a guide sequence by RNAi machinery. In one embodiment, such chemical modifications comprise any chemical group at the 5'-end of the sense strand of the siNA, or any other group that serves to render the sense strand inactive as a guide sequence for mediating RNA interference. These modifications, for example, can result in a molecule where the 5'-end of the sense strand no longer has a free 5'-hydroxyl (5'-OH) or a free 5'-phosphate group (e.g., phosphate, diphosphate, triphosphate, cyclic phosphate etc.). Non-limiting examples of such siNA constructs are described herein, such as "Stab 9/10", "Stab 7/8", "Stab 7/19" and "Stab 17/22" chemistries and variants thereof (see Table IV) wherein the 5'-end and 3'-end of the sense strand of the siNA do not comprise a hydroxyl group or phosphate group.

In one embodiment, the invention features a method for generating siNA molecules of the invention with improved specificity for down regulating or inhibiting the expression of a target nucleic acid (e.g., a DNA or RNA such as a gene or its corresponding RNA), comprising introducing one or more chemical modifications into the structure of a siNA molecule that prevent a strand or portion of the siNA molecule from acting as a template or guide sequence for RNAi activity. In one embodiment, the inactive strand or sense region of the siNA molecule is the sense strand or sense region of the siNA molecule, i.e. the strand or region of the siNA that does not have

complementarity to the target nucleic acid sequence. In one embodiment, such chemical modifications comprise any chemical group at the 5'-end of the sense strand or region of the siNA that does not comprise a 5'-hydroxyl (5'-OH) or 5'-phosphate group, or any other group that serves to render the sense strand or sense region inactive as a guide sequence for mediating RNA interference. Non-limiting examples of such siNA constructs are described herein, such as "Stab 9/10", "Stab 7/8", "Stab 7/19" and "Stab 17/22" chemistries and variants thereof (see Table IV) wherein the 5'-end and 3'-end of the sense strand of the siNA do not comprise a hydroxyl group or phosphate group.

In one embodiment, the invention features a method for screening siNA molecules that are active in mediating RNA interference against a target nucleic acid sequence comprising (a) generating a plurality of unmodified siNA molecules, (b) screening the siNA molecules of step (a) under conditions suitable for isolating siNA molecules that are active in mediating RNA interference against the target nucleic acid sequence, and (c) introducing chemical modifications (e.g. chemical modifications as described herein or as otherwise known in the art) into the active siNA molecules of (b). In one embodiment, the method further comprises re-screening the chemically modified siNA molecules of step (c) under conditions suitable for isolating chemically modified siNA molecules that are active in mediating RNA interference against the target nucleic acid sequence.

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In one embodiment, the invention features a method for screening chemically modified siNA molecules that are active in mediating RNA interference against a target nucleic acid sequence comprising (a) generating a plurality of chemically modified siNA molecules (e.g. siNA molecules as described herein or as otherwise known in the art), and (b) screening the siNA molecules of step (a) under conditions suitable for isolating chemically modified siNA molecules that are active in mediating RNA interference against the target nucleic acid sequence.

The term "ligand" refers to any compound or molecule, such as a drug, peptide, hormone, or neurotransmitter, that is capable of interacting with another compound, such as a receptor, either directly or indirectly. The receptor that interacts with a ligand can be present on the surface of a cell or can alternately be an intercullular receptor. Interaction

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of the ligand with the receptor can result in a biochemical reaction, or can simply be a physical interaction or association.

In another embodiment, the invention features a method for generating siNA molecules of the invention with improved bioavailability comprising (a) introducing an excipient formulation to a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved bioavailability. Such excipients include polymers such as cyclodextrins, lipids, cationic lipids, polyamines, phospholipids, nanoparticles, receptors, ligands, and others.

In another embodiment, the invention features a method for generating siNA molecules of the invention with improved bioavailability comprising (a) introducing nucleotides having any of Formulae I-VII or any combination thereof into a siNA molecule, and (b) assaying the siNA molecule of step (a) under conditions suitable for isolating siNA molecules having improved bioavailability.

In another embodiment, polyethylene glycol (PEG) can be covalently attached to siNA compounds of the present invention. The attached PEG can be any molecular weight, preferably from about 2,000 to about 50,000 daltons (Da).

The present invention can be used alone or as a component of a kit having at least one of the reagents necessary to carry out the *in vitro* or *in vivo* introduction of RNA to test samples and/or subjects. For example, preferred components of the kit include a siNA molecule of the invention and a vehicle that promotes introduction of the siNA into cells of interest as described herein (e.g., using lipids and other methods of transfection known in the art, see for example Beigelman *et al*, US 6,395,713). The kit can be used for target validation, such as in determining gene function and/or activity, or in drug optimization, and in drug discovery (see for example Usman et al., USSN 60/402,996). Such a kit can also include instructions to allow a user of the kit to practice the invention.

The term "short interfering nucleic acid", "siNA", "short interfering RNA", "siRNA", "short interfering nucleic acid molecule", "short interfering oligonucleotide molecule", or "chemically-modified short interfering nucleic acid molecule" as used herein refers to any nucleic acid molecule capable of inhibiting or down regulating gene

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expression or viral replication, for example by mediating RNA interference "RNAi" or gene silencing in a sequence-specific manner; see for example Zamore et al., 2000, Cell, 101, 25-33; Bass, 2001, Nature, 411, 428-429; Elbashir et al., 2001, Nature, 411, 494-498; and Kreutzer et al., International PCT Publication No. WO 00/44895; Zernicka-Goetz et al., International PCT Publication No. WO 01/36646; Fire, International PCT Publication No. WO 99/32619; Plaetinck et al., International PCT Publication No. WO 00/01846; Mello and Fire, International PCT Publication No. WO 01/29058; Deschamps-Depaillette, International PCT Publication No. WO 99/07409; and Li et al., International PCT Publication No. WO 00/44914; Allshire, 2002, Science, 297, 1818-1819; Volpe et al., 2002, Science, 297, 1833-1837; Jenuwein, 2002, Science, 297, 2215-2218; and Hall et al., 2002, Science, 297, 2232-2237; Hutvagner and Zamore, 2002, Science, 297, 2056-60; McManus et al., 2002, RNA, 8, 842-850; Reinhart et al., 2002, Gene & Dev., 16, 1616-1626; and Reinhart & Bartel, 2002, Science, 297, 1831). Non limiting examples of siNA molecules of the invention are shown in Figures 4-6, and Tables II and III herein. For example the siNA can be a double-stranded polynucleotide molecule comprising self-complementary sense and antisense regions, wherein the antisense region comprises nucleotide sequence that is complementary to nucleotide sequence in a target nucleic acid molecule or a portion thereof and the sense region having nucleotide sequence corresponding to the target nucleic acid sequence or a portion thereof. The siNA can be assembled from two separate oligonucleotides, where one strand is the sense strand and the other is the antisense strand, wherein the antisense and sense strands are self-complementary (i.e. each strand comprises nucleotide sequence that is complementary to nucleotide sequence in the other strand; such as where the antisense strand and sense strand form a duplex or double stranded structure, for example wherein the double stranded region is about 19 base pairs); the antisense strand comprises nucleotide sequence that is complementary to nucleotide sequence in a target nucleic acid molecule or a portion thereof and the sense strand comprises nucleotide sequence corresponding to the target nucleic acid sequence or a portion thereof. Alternatively, the siNA is assembled from a single oligonucleotide, where the selfcomplementary sense and antisense regions of the siNA are linked by means of a nucleic acid based or non-nucleic acid-based linker(s). The siNA can be a polynucleotide with a duplex, asymmetric duplex, hairpin or asymmetric hairpin secondary structure, having

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self-complementary sense and antisense regions, wherein the antisense region comprises nucleotide sequence that is complementary to nucleotide sequence in a separate target nucleic acid molecule or a portion thereof and the sense region having nucleotide sequence corresponding to the target nucleic acid sequence or a portion thereof. The siNA can be a circular single-stranded polynucleotide having two or more loop structures and a stem comprising self-complementary sense and antisense regions, wherein the antisense region comprises nucleotide sequence that is complementary to nucleotide sequence in a target nucleic acid molecule or a portion thereof and the sense region having nucleotide sequence corresponding to the target nucleic acid sequence or a portion thereof, and wherein the circular polynucleotide can be processed either in vivo or in vitro to generate an active siNA molecule capable of mediating RNAi. The siNA can also comprise a single stranded polynucleotide having nucleotide sequence complementary to nucleotide sequence in a target nucleic acid molecule or a portion thereof (for example, where such siNA molecule does not require the presence within the siNA molecule of nucleotide sequence corresponding to the target nucleic acid sequence or a portion thereof), wherein the single stranded polynucleotide can further comprise a terminal phosphate group, such as a 5'-phosphate (see for example Martinez et al., 2002, Cell., 110, 563-574 and Schwarz et al., 2002, Molecular Cell, 10, 537-568), or 5',3'diphosphate. In certain embodiments, the siNA molecule of the invention comprises separate sense and antisense sequences or regions, wherein the sense and antisense regions are covalently linked by nucleotide or non-nucleotide linkers molecules as is known in the art, or are alternately non-covalently linked by ionic interactions, hydrogen bonding, van der waals interactions, hydrophobic intercations, and/or stacking interactions. In certain embodiments, the siNA molecules of the invention comprise nucleotide sequence that is complementary to nucleotide sequence of a target gene. In another embodiment, the siNA molecule of the invention interacts with nucleotide sequence of a target gene in a manner that causes inhibition of expression of the target gene. As used herein, siNA molecules need not be limited to those molecules containing only RNA, but further encompasses chemically-modified nucleotides and nonnucleotides. In certain embodiments, the short interfering nucleic acid molecules of the invention lack 2'-hydroxy (2'-OH) containing nucleotides. Applicant describes in certain embodiments short interfering nucleic acids that do not require the presence of

nucleotides having a 2'-hydroxy group for mediating RNAi and as such, short interfering nucleic acid molecules of the invention optionally do not include any ribonucleotides (e.g., nucleotides having a 2'-OH group). Such siNA molecules that do not require the presence of ribonucleotides within the siNA molecule to support RNAi can however have an attached linker or linkers or other attached or associated groups, moieties, or 5 chains containing one or more nucleotides with 2'-OH groups. Optionally, siNA molecules can comprise ribonucleotides at about 5, 10, 20, 30, 40, or 50% of the nucleotide positions. The modified short interfering nucleic acid molecules of the invention can also be referred to as short interfering modified oligonucleotides "siMON." As used herein, the term siNA is meant to be equivalent to other terms used to describe 10 nucleic acid molecules that are capable of mediating sequence specific RNAi, for example short interfering RNA (siRNA), double-stranded RNA (dsRNA), micro-RNA (miRNA), short hairpin RNA (shRNA), short interfering oligonucleotide, short interfering nucleic acid, short interfering modified oligonucleotide, chemically-modified siRNA, post-transcriptional gene silencing RNA (ptgsRNA), and others. In addition, as 15 used herein, the term RNAi is meant to be equivalent to other terms used to describe sequence specific RNA interference, such as post transcriptional gene silencing, translational inhibition, or epigenetics. For example, siNA molecules of the invention can be used to epigenetically silence genes at both the post-transcriptional level or the pre-transcriptional level. In a non-limiting example, epigenetic regulation of gene 20 expression by siNA molecules of the invention can result from siNA mediated modification of chromatin structure to alter gene expression (see, for example, Verdel et al., 2004, Science, 303, 672-676; Pal-Bhadra et al., 2004, Science, 303, 669-672; Allshire, 2002, Science, 297, 1818-1819; Volpe et al., 2002, Science, 297, 1833-1837; Jenuwein, 2002, Science, 297, 2215-2218; and Hall et al., 2002, Science, 297, 2232-25 2237).

In one embodiment, a siNA molecule of the invention is a duplex forming oligonucleotide "DFO", (see for example **Figures 14-15** and Vaish *et al.*, USSN 10/727,780 filed December 3, 2003).

In one embodiment, a siNA molecule of the invention is a multifunctional siNA, (see for example Figures 16-22 and Jadhav et al., USSN 60/543,480, filed February 10,

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2004). The multifunctional siNA of the invention can comprise sequence targeting, for example, two regions of SARS RNA (see for example target sequences in **Tables II and III**) or alternately, SARS RNA and cellular RNA involved in SARS virus infection or replication. In another embodiment, a multifunctional siNA of the invention can comprise sequence targeting for example both viral genes encoding RNAi inhibitory factors and viral genes encoding viral structural proteins.

By "asymmetric hairpin" as used herein is meant a linear siNA molecule comprising an antisense region, a loop portion that can comprise nucleotides or non-nucleotides, and a sense region that comprises fewer nucleotides than the antisense region to the extent that the sense region has enough complementary nucleotides to base pair with the antisense region and form a duplex with loop. For example, an asymmetric hairpin siNA molecule of the invention can comprise an antisense region having length sufficient to mediate RNAi in a cell or in vitro system (e.g. about 19 to about 22 (e.g., about 19, 20, 21, or 22) nucleotides) and a loop region comprising about 4 to about 8 (e.g., about 4, 5, 6, 7, or 8) nucleotides, and a sense region having about 3 to about 18 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, or 18) nucleotides that are complementary to the antisense region. The asymmetric hairpin siNA molecule can also comprise a 5'-terminal phosphate group that can be chemically modified. The loop portion of the asymmetric hairpin siNA molecule can comprise nucleotides, non-nucleotides, linker molecules, or conjugate molecules as described herein.

By "asymmetric duplex" as used herein is meant a siNA molecule having two separate strands comprising a sense region and an antisense region, wherein the sense region comprises fewer nucleotides than the antisense region to the extent that the sense region has enough complementary nucleotides to base pair with the antisense region and form a duplex. For example, an asymmetric duplex siNA molecule of the invention can comprise an antisense region having length sufficient to mediate RNAi in a cell or in vitro system (e.g. about 19 to about 22 (e.g. about 19, 20, 21, or 22) nucleotides) and a sense region having about 3 to about 18 (e.g., about 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, or 18) nucleotides that are complementary to the antisense region.

By "modulate" is meant that the expression of the gene, or level of RNA molecule or equivalent RNA molecules encoding one or more proteins or protein subunits, or

activity of one or more proteins or protein subunits is up regulated or down regulated, such that expression, level, or activity is greater than or less than that observed in the absence of the modulator. For example, the term "modulate" can mean "inhibit," but the use of the word "modulate" is not limited to this definition.

By "inhibit", "down-regulate", or "reduce", it is meant that the expression of the gene, or level of RNA molecules or equivalent RNA molecules encoding one or more proteins or protein subunits, or activity of one or more proteins or protein subunits, is reduced below that observed in the absence of the nucleic acid molecules (e.g., siNA) of the invention. In one embodiment, inhibition, down-regulation or reduction with an siNA molecule is below that level observed in the presence of an inactive or attenuated molecule. In another embodiment, inhibition, down-regulation, or reduction with siNA molecules is below that level observed in the presence of, for example, an siNA molecule with scrambled sequence or with mismatches. In another embodiment, inhibition, down-regulation, or reduction of gene expression with a nucleic acid molecule of the instant invention is greater in the presence of the nucleic acid molecule than in its absence.

By "gene", or "target gene", is meant, a nucleic acid that encodes an RNA, for example, nucleic acid sequences including, but not limited to, structural genes encoding a polypeptide. A gene or target gene can also encode a functional RNA (fRNA) or noncoding RNA (ncRNA), such as small temporal RNA (stRNA), micro RNA (miRNA), small nuclear RNA (snRNA), short interfering RNA (siRNA), small nucleolar RNA (snRNA), ribosomal RNA (rRNA), transfer RNA (tRNA) and precursor RNAs thereof. Such non-coding RNAs can serve as target nucleic acid molecules for siNA mediated RNA interference in modulating the activity of fRNA or ncRNA involved in functional or regulatory cellular processes. Abberant fRNA or ncRNA activity leading to disease can therefore be modulated by siNA molecules of the invention. siNA molecules targeting fRNA and ncRNA can also be used to manipulate or alter the genotype or phenotype of an organism or cell, by intervening in cellular processes such as genetic imprinting, transcription, translation, or nucleic acid processing (e.g., transamination, methylation etc.). The target gene can be a gene derived from a cell, an endogenous gene, a transgene, or exogenous genes such as genes of a pathogen, for example a virus,

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which is present in the cell after infection thereof. The cell containing the target gene can be derived from or contained in any organism, for example a plant, animal, protozoan, virus, bacterium, or fungus. Non-limiting examples of plants include monocots, dicots, or gymnosperms. Non-limiting examples of animals include vertebrates or invertebrates. Non-limiting examples of fungi include molds or yeasts.

By "SARS" or "SARS virus" as used herein is meant the SARS virus or any protein, peptide, or polypeptide, having SARS virus activity or encoded by the SARS genome. The term "SARS" also includes nucleic acid molecules encoding RNA or protein(s) associated with the development and/or maintenance of SARS virus infection, such as nucleic acid molecules which encode SARS RNA or polypeptides (such as polynucleotides having Genbank Accession numbers shown in Table I), including polypeptides of different strains of SARS, mutant SARS genes, and splice variants of SARS genes, as well as genes involved in SARS pathways of gene expression and/or SARS activity. Also, the term "SARS" is meant to encompass SARS viral gene products and genes that modulate cellular targets for SARS virus infection, such as those described herein.

By "SARS protein" or "SARS virus protein" is meant, protein, peptide, or polypeptide, having SARS virus activity or encoded by the SARS genome or alternately, cellular proteins involved in SARS virus infection and/or replication.

By "homologous sequence" is meant, a nucleotide sequence that is shared by one or more polynucleotide sequences, such as genes, gene transcripts and/or non-coding polynucleotides. For example, a homologous sequence can be a nucleotide sequence that is shared by two or more genes encoding related but different proteins, such as different members of a gene family, different protein epitopes, different protein isoforms or completely divergent genes, such as a cytokine and its corresponding receptors. A homologous sequence can be a nucleotide sequence that is shared by two or more non-coding polynucleotides, such as noncoding DNA or RNA, regulatory sequences, introns, and sites of transcriptional control or regulation. Homologous sequences can also include conserved sequence regions shared by more than one polynucleotide sequence. Homology does not need to be perfect homology (e.g., 100%), as partially homologous sequences are also contemplated by the instant invention (e.g., 99%, 98%, 97%, 96%,

95%, 94%, 93%, 92%, 91%, 90%, 89%, 88%, 87%, 86%, 85%, 84%, 83%, 82%, 81%, 80% etc.).

By "conserved sequence region" is meant, a nucleotide sequence of one or more regions in a polynucleotide does not vary significantly between generations or from one biological system or organism to another biological system or organism. The polynucleotide can include both coding and non-coding DNA and RNA.

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By "sense region" is meant a nucleotide sequence of a siNA molecule having complementarity to an antisense region of the siNA molecule. In addition, the sense region of a siNA molecule can comprise a nucleic acid sequence having homology with a target nucleic acid sequence.

By "antisense region" is meant a nucleotide sequence of a siNA molecule having complementarity to a target nucleic acid sequence. In addition, the antisense region of a siNA molecule can optionally comprise a nucleic acid sequence having complementarity to a sense region of the siNA molecule.

By "target nucleic acid" is meant any nucleic acid sequence whose expression or activity is to be modulated. The target nucleic acid can be DNA or RNA.

By "complementarity" is meant that a nucleic acid can form hydrogen bond(s) with another nucleic acid sequence by either traditional Watson-Crick or other non-traditional types. In reference to the nucleic molecules of the present invention, the binding free energy for a nucleic acid molecule with its complementary sequence is sufficient to allow the relevant function of the nucleic acid to proceed, e.g., RNAi activity. Determination of binding free energies for nucleic acid molecules is well known in the art (see, e.g., Turner et al., 1987, CSH Symp. Quant. Biol. LII pp.123-133; Frier et al., 1986, Proc. Nat. Acad. Sci. USA 83:9373-9377; Turner et al., 1987, J. Am. Chem. Soc. 109:3783-3785). A percent complementarity indicates the percentage of contiguous residues in a nucleic acid molecule that can form hydrogen bonds (e.g., Watson-Crick base pairing) with a second nucleic acid sequence (e.g., 5, 6, 7, 8, 9, or 10 nucleotides out of a total of 10 nucleotides in the first oligonuelcotide being based paired to a second nucleic acid sequence having 10 nucleotides represents 50%, 60%, 70%, 80%, 90%, and

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100% complementary respectively). "Perfectly complementary" means that all the contiguous residues of a nucleic acid sequence will hydrogen bond with the same number of contiguous residues in a second nucleic acid sequence.

The siNA molecules of the invention represent a novel therapeutic approach to treat various diseases and conditions, including SARS virus infection, acute respiratory failure, viral pneumonia, and any other indications that can respond to the level of SARS in a cell or tissue. The reduction of SARS expression and thus reduction in the level of the respective protein relieves, to some extent, the symptoms of the disease or condition.

In one embodiment of the present invention, each sequence of a siNA molecule of the invention is independently about 18 to about 24 nucleotides in length, in specific embodiments about 18, 19, 20, 21, 22, 23, or 24 nucleotides in length. In another embodiment, the siNA duplexes of the invention independently comprise about 17 to about 23 base pairs (e.g., about 17, 18, 19, 20, 21, 22 or 23). In yet another embodiment, siNA molecules of the invention comprising hairpin or circular structures are about 35 to about 55 (e.g., about 35, 40, 45, 50 or 55) nucleotides in length, or about 38 to about 44 (e.g., 38, 39, 40, 41, 42, 43 or 44) nucleotides in length and comprising about 16 to about 22 (e.g., about 16, 17, 18, 19, 20, 21 or 22) base pairs. Exemplary siNA molecules of the invention are shown in **Table II**. Exemplary synthetic siNA molecules of the invention are shown in **Table II** and/or **Figures 4-5**.

As used herein "cell" is used in its usual biological sense, and does not refer to an entire multicellular organism, e.g., specifically does not refer to a human. The cell can be present in an organism, e.g., birds, plants and mammals such as humans, cows, sheep, apes, monkeys, swine, dogs, and cats. The cell can be prokaryotic (e.g., bacterial cell) or eukaryotic (e.g., mammalian or plant cell). The cell can be of somatic or germ line origin, totipotent or pluripotent, dividing or non-dividing. The cell can also be derived from or can comprise a gamete or embryo, a stem cell, or a fully differentiated cell.

The siNA molecules of the invention are added directly, or can be complexed with cationic lipids, packaged within liposomes, or otherwise delivered to target cells or tissues. The nucleic acid or nucleic acid complexes can be locally administered to relevant tissues *ex vivo*, or *in vivo* through injection, infusion pump or stent, with or

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without their incorporation in biopolymers. In particular embodiments, the nucleic acid molecules of the invention comprise sequences shown in **Tables II-III** and/or **Figures 4-5**. Examples of such nucleic acid molecules consist essentially of sequences defined in these tables and figures. Furthermore, the chemically modified constructs described in **Table IV** can be applied to any siNA sequence of the invention.

In another aspect, the invention provides mammalian cells containing one or more siNA molecules of this invention. The one or more siNA molecules can independently be targeted to the same or different sites.

By "RNA" is meant a molecule comprising at least one ribonucleotide residue. By "ribonucleotide" is meant a nucleotide with a hydroxyl group at the 2' position of a β-D-ribo-furanose moiety. The terms include double-stranded RNA, single-stranded RNA, isolated RNA such as partially purified RNA, essentially pure RNA, synthetic RNA, recombinantly produced RNA, as well as altered RNA that differs from naturally occurring RNA by the addition, deletion, substitution and/or alteration of one or more nucleotides. Such alterations can include addition of non-nucleotide material, such as to the end(s) of the siNA or internally, for example at one or more nucleotides of the RNA. Nucleotides in the RNA molecules of the instant invention can also comprise non-standard nucleotides, such as non-naturally occurring nucleotides or chemically synthesized nucleotides or deoxynucleotides. These altered RNAs can be referred to as analogs or analogs of naturally-occurring RNA.

By "subject" is meant an organism, which is a donor or recipient of explanted cells or the cells themselves. "Subject" also refers to an organism to which the nucleic acid molecules of the invention can be administered. A subject can be a mammal or mammalian cells, including a human or human cells.

The term "phosphorothioate" as used herein refers to an internucleotide linkage having Formula I, wherein Z and/or W comprise a sulfur atom. Hence, the term phosphorothioate refers to both phosphorothioate and phosphorodithioate internucleotide linkages.

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The term "phosphonoacetate" as used herein refers to an internucleotide linkage having Formula I, wherein Z and/or W comprise an acetyl or protected acetyl group.

The term "thiophosphonoacetate" as used herein refers to an internucleotide linkage having Formula I, wherein Z comprises an acetyl or protected acetyl group and W comprises a sulfur atom or alternately W comprises an acetyl or protected acetyl group and Z comprises a sulfur atom.

The term "universal base" as used herein refers to nucleotide base analogs that form base pairs with each of the natural DNA/RNA bases with little discrimination between them. Non-limiting examples of universal bases include C-phenyl, C-naphthyl and other aromatic derivatives, inosine, azole carboxamides, and nitroazole derivatives such as 3-nitropyrrole, 4-nitroindole, 5-nitroindole, and 6-nitroindole as known in the art (see for example Loakes, 2001, *Nucleic Acids Research*, 29, 2437-2447).

The term "acyclic nucleotide" as used herein refers to any nucleotide having an acyclic ribose sugar, for example where any of the ribose carbons (C1, C2, C3, C4, or C5), are independently or in combination absent from the nucleotide.

The nucleic acid molecules of the instant invention, individually, or in combination or in conjunction with other drugs, can be used to treat diseases or conditions discussed herein (e.g., cancers and othe proliferative conditions). For example, to treat a particular disease or condition, the siNA molecules can be administered to a subject or can be administered to other appropriate cells evident to those skilled in the art, individually or in combination with one or more drugs under conditions suitable for the treatment.

In a further embodiment, the siNA molecules can be used in combination with other known treatments to treat conditions or diseases discussed above. For example, the described molecules could be used in combination with one or more known therapeutic agents to treat a disease or condition. Non-limiting examples of other therapeutic agents that can be readily combined with a siNA molecule of the invention are enzymatic nucleic acid molecules, allosteric nucleic acid molecules, antisense, decoy, or aptamer nucleic acid molecules, antibodies such as monoclonal antibodies, small molecules, and other organic and/or inorganic compounds including metals, salts and ions.

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In one embodiment, the invention features an expression vector comprising a nucleic acid sequence encoding at least one siNA molecule of the invention, in a manner which allows expression of the siNA molecule. For example, the vector can contain sequence(s) encoding both strands of a siNA molecule comprising a duplex. The vector can also contain sequence(s) encoding a single nucleic acid molecule that is self-complementary and thus forms a siNA molecule. Non-limiting examples of such expression vectors are described in Paul et al., 2002, Nature Biotechnology, 19, 505; Miyagishi and Taira, 2002, Nature Biotechnology, 19, 497; Lee et al., 2002, Nature Biotechnology, 19, 500; and Novina et al., 2002, Nature Medicine, advance online publication doi:10.1038/nm725.

In another embodiment, the invention features a mammalian cell, for example, a human cell, including an expression vector of the invention.

In yet another embodiment, the expression vector of the invention comprises a sequence for a siNA molecule having complementarity to a RNA molecule referred to by a Genbank Accession numbers, for example Genbank Accession Nos. shown in **Table I**.

In one embodiment, an expression vector of the invention comprises a nucleic acid sequence encoding two or more siNA molecules, which can be the same or different.

In another aspect of the invention, siNA molecules that interact with target RNA molecules and down-regulate gene encoding target RNA molecules (for example target RNA molecules referred to by Genbank Accession numbers herein) are expressed from transcription units inserted into DNA or RNA vectors. The recombinant vectors can be DNA plasmids or viral vectors. siNA expressing viral vectors can be constructed based on, but not limited to, adeno-associated virus, retrovirus, adenovirus, or alphavirus. The recombinant vectors capable of expressing the siNA molecules can be delivered as described herein, and persist in target cells. Alternatively, viral vectors can be used that provide for transient expression of siNA molecules. Such vectors can be repeatedly administered as necessary. Once expressed, the siNA molecules bind and down-regulate gene function or expression via RNA interference (RNAi). Delivery of siNA expressing vectors can be systemic, such as by intravenous or intramuscular administration, by administration to target cells ex-planted from a subject followed by reintroduction into

the subject, or by any other means that would allow for introduction into the desired target cell.

By "vectors" is meant any nucleic acid- and/or viral-based technique used to deliver a desired nucleic acid.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

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BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows a non-limiting example of a scheme for the synthesis of siNA molecules. The complementary siNA sequence strands, strand 1 and strand 2, are synthesized in tandem and are connected by a cleavable linkage, such as a nucleotide succinate or abasic succinate, which can be the same or different from the cleavable linker used for solid phase synthesis on a solid support. The synthesis can be either solid phase or solution phase, in the example shown, the synthesis is a solid phase synthesis. The synthesis is performed such that a protecting group, such as a dimethoxytrityl group, remains intact on the terminal nucleotide of the tandem oligonucleotide. Upon cleavage and deprotection of the oligonucleotide, the two siNA strands spontaneously hybridize to form a siNA duplex, which allows the purification of the duplex by utilizing the properties of the terminal protecting group, for example by applying a trityl on purification method wherein only duplexes/oligonucleotides with the terminal protecting group are isolated.

Figure 2 shows a MALDI-TOF mass spectrum of a purified siNA duplex synthesized by a method of the invention. The two peaks shown correspond to the predicted mass of the separate siNA sequence strands. This result demonstrates that the siNA duplex generated from tandem synthesis can be purified as a single entity using a simple trityl-on purification methodology.

Figure 3 shows a non-limiting proposed mechanistic representation of target RNA degradation involved in RNAi. Double-stranded RNA (dsRNA), which is generated by RNA-dependent RNA polymerase (RdRP) from foreign single-stranded RNA, for example viral, transposon, or other exogenous RNA, activates the DICER enzyme that in

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turn generates siNA duplexes. Alternately, synthetic or expressed siNA can be introduced directly into a cell by appropriate means. An active siNA complex forms which recognizes a target RNA, resulting in degradation of the target RNA by the RISC endonuclease complex or in the synthesis of additional RNA by RNA-dependent RNA polymerase (RdRP), which can activate DICER and result in additional siNA molecules, thereby amplifying the RNAi response.

Figure 4A-F shows non-limiting examples of chemically-modified siNA constructs of the present invention. In the figure, N stands for any nucleotide (adenosine, guanosine, cytosine, uridine, or optionally thymidine, for example thymidine can be substituted in the overhanging regions designated by parenthesis (N N). Various modifications are shown for the sense and antisense strands of the siNA constructs.

Figure 4A: The sense strand comprises 21 nucleotides wherein the two terminal 3'-nucleotides are optionally base paired and wherein all nucleotides present are ribonucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. The antisense strand comprises 21 nucleotides, optionally having a 3'-terminal glyceryl moiety wherein the two terminal 3'-nucleotides are optionally complementary to the target RNA sequence, and wherein all nucleotides present are ribonucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. A modified internucleotide linkage, such as a phosphorothioate, phosphorodithioate or other modified internucleotide linkage as described herein, shown as "s", optionally connects the (N N) nucleotides in the antisense strand.

Figure 4B: The sense strand comprises 21 nucleotides wherein the two terminal 3'-nucleotides are optionally base paired and wherein all pyrimidine nucleotides that may be present are 2'deoxy-2'-fluoro modified nucleotides and all purine nucleotides that may be present are 2'-O-methyl modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. The antisense strand comprises 21 nucleotides, optionally having a 3'-terminal glyceryl moiety and wherein the two terminal 3'-nucleotides are optionally complementary to the target RNA sequence, and wherein all

pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides and all purine nucleotides that may be present are 2'-O-methyl modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. A modified internucleotide linkage, such as a phosphorothioate, phosphorodithioate or other modified internucleotide linkage as described herein, shown as "s", optionally connects the (N N) nucleotides in the sense and antisense strand.

Figure 4C: The sense strand comprises 21 nucleotides having 5'- and 3'- terminal cap moieties wherein the two terminal 3'-nucleotides are optionally base paired and wherein all pyrimidine nucleotides that may be present are 2'-O-methyl or 2'-deoxy-2'-fluoro modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. The antisense strand comprises 21 nucleotides, optionally having a 3'-terminal glyceryl moiety and wherein the two terminal 3'-nucleotides are optionally complementary to the target RNA sequence, and wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. A modified internucleotide linkage, such as a phosphorothioate, phosphorodithioate or other modified internucleotide linkage as described herein, shown as "s", optionally connects the (N N) nucleotides in the antisense strand.

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Figure 4D: The sense strand comprises 21 nucleotides having 5'- and 3'- terminal cap moieties wherein the two terminal 3'-nucleotides are optionally base paired and wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein and wherein and all purine nucleotides that may be present are 2'-deoxy nucleotides. The antisense strand comprises 21 nucleotides, optionally having a 3'-terminal glyceryl moiety and wherein the two terminal 3'-nucleotides are optionally complementary to the target RNA sequence, wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides and all purine nucleotides that may be present are

2'-O-methyl modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. A modified internucleotide linkage, such as a phosphorothioate, phosphorodithioate or other modified internucleotide linkage as described herein, shown as "s", optionally connects the (N N) nucleotides in the antisense strand.

Figure 4E: The sense strand comprises 21 nucleotides having 5'- and 3'- terminal cap moieties wherein the two terminal 3'-nucleotides are optionally base paired and wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. The antisense strand comprises 21 nucleotides, optionally having a 3'-terminal glyceryl moiety and wherein the two terminal 3'-nucleotides are optionally complementary to the target RNA sequence, and wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides and all purine nucleotides that may be present are 2'-O-methyl modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein. A modified internucleotide linkage, such as a phosphorothioate, phosphorodithioate or other modified internucleotide linkage as described herein, shown as "s", optionally connects the (N N) nucleotides in the antisense strand.

Figure 4F: The sense strand comprises 21 nucleotides having 5'- and 3'- terminal cap moieties wherein the two terminal 3'-nucleotides are optionally base paired and wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides except for (N N) nucleotides, which can comprise ribonucleotides, deoxynucleotides, universal bases, or other chemical modifications described herein and wherein and all purine nucleotides that may be present are 2'-deoxy nucleotides. The antisense strand comprises 21 nucleotides, optionally having a 3'-terminal glyceryl moiety and wherein the two terminal 3'-nucleotides are optionally complementary to the target RNA sequence, and having one 3'-terminal phosphorothioate internucleotide linkage and wherein all pyrimidine nucleotides that may be present are 2'-deoxy-2'-fluoro modified nucleotides and all purine nucleotides that may be present are 2'-deoxy nucleotides except for (N N) nucleotides, which can comprise ribonucleotides,

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deoxynucleotides, universal bases, or other chemical modifications described herein. A modified internucleotide linkage, such as a phosphorothioate, phosphorodithioate or other modified internucleotide linkage as described herein, shown as "s", optionally connects the (N N) nucleotides in the antisense strand. The antisense strand of constructs A-F comprise sequence complementary to any target nucleic acid sequence of the invention. Furthermore, when a glyceryl moiety (L) is present at the 3'-end of the antisense strand for any construct shown in Figure 4 A-F, the modified internucleotide linkage is optional.

Figure 5A-F shows non-limiting examples of specific chemically-modified siNA sequences of the invention. A-F applies the chemical modifications described in Figure 4A-F to a SARS virus siNA sequence. Such chemical modifications can be applied to any SARS sequence and/or SARS polymorphism sequence.

Figure 6 shows non-limiting examples of different siNA constructs of the invention. The examples shown (constructs 1, 2, and 3) have 19 representative base pairs; however, different embodiments of the invention include any number of base pairs described herein. Bracketed regions represent nucleotide overhangs, for example comprising about 1, 2, 3, or 4 nucleotides in length, preferably about 2 nucleotides. Constructs 1 and 2 can be used independently for RNAi activity. Construct 2 can comprise a polynucleotide or non-nucleotide linker, which can optionally be designed as a biodegradable linker. In one embodiment, the loop structure shown in construct 2 can comprise a biodegradable linker that results in the formation of construct 1 in vivo and/or in vitro. In another example, construct 3 can be used to generate construct 2 under the same principle wherein a linker is used to generate the active siNA construct 2 in vivo and/or in vitro, which can optionally utilize another biodegradable linker to generate the active siNA construct 1 in vivo and/or in vitro. As such, the stability and/or activity of the siNA constructs can be modulated based on the design of the siNA construct for use in vivo or in vitro and/or in vitro.

Figure 7A-C is a diagrammatic representation of a scheme utilized in generating an expression cassette to generate siNA hairpin constructs.

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Figure 7A: A DNA oligomer is synthesized with a 5'-restriction site (R1) sequence followed by a region having sequence identical (sense region of siNA) to a predetermined SARS target sequence, wherein the sense region comprises, for example, about 19, 20, 21, or 22 nucleotides (N) in length, which is followed by a loop sequence of defined sequence (X), comprising, for example, about 3 to about 10 nucleotides.

- Figure 7B: The synthetic construct is then extended by DNA polymerase to generate a hairpin structure having self-complementary sequence that will result in a siNA transcript having specificity for a SARS target sequence and having self-complementary sense and antisense regions.
- Figure 7C: The construct is heated (for example to about 95°C) to linearize the sequence, thus allowing extension of a complementary second DNA strand using a primer to the 3'-restriction sequence of the first strand. The double-stranded DNA is then inserted into an appropriate vector for expression in cells. The construct can be designed such that a 3'-terminal nucleotide overhang results from the transcription, for example by engineering restriction sites and/or utilizing a poly-U termination region as described in Paul et al., 2002, Nature Biotechnology, 29, 505-508.
 - Figure 8A-C is a diagrammatic representation of a scheme utilized in generating an expression cassette to generate double-stranded siNA constructs.
- Figure 8A: A DNA oligomer is synthesized with a 5'-restriction (R1) site sequence followed by a region having sequence identical (sense region of siNA) to a predetermined SARS target sequence, wherein the sense region comprises, for example, about 19, 20, 21, or 22 nucleotides (N) in length, and which is followed by a 3'-restriction site (R2) which is adjacent to a loop sequence of defined sequence (X).
- Figure 8B: The synthetic construct is then extended by DNA polymerase to generate a hairpin structure having self-complementary sequence.
 - Figure 8C: The construct is processed by restriction enzymes specific to R1 and R2 to generate a double-stranded DNA which is then inserted into an appropriate vector for expression in cells. The transcription cassette is designed such that a U6 promoter region flanks each side of the dsDNA which generates the separate sense and antisense

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strands of the siNA. Poly T termination sequences can be added to the constructs to generate U overhangs in the resulting transcript.

Figure 9A-E is a diagrammatic representation of a method used to determine target sites for siNA mediated RNAi within a particular target nucleic acid sequence, such as messenger RNA.

Figure 9A: A pool of siNA oligonucleotides are synthesized wherein the antisense region of the siNA constructs has complementarity to target sites across the target nucleic acid sequence, and wherein the sense region comprises sequence complementary to the antisense region of the siNA.

10 Figure 9B&C: (Figure 9B) The sequences are pooled and are inserted into vectors such that (Figure 9C) transfection of a vector into cells results in the expression of the siNA.

Figure 9D: Cells are sorted based on phenotypic change that is associated with modulation of the target nucleic acid sequence.

Figure 9E: The siNA is isolated from the sorted cells and is sequenced to identify efficacious target sites within the target nucleic acid sequence.

Figure 10 shows non-limiting examples of different stabilization chemistries (1-10) that can be used, for example, to stabilize the 3'-end of siNA sequences of the invention, including (1) [3-3']-inverted deoxyribose; (2) deoxyribonucleotide; (3) [5'-3']-3'-deoxyribonucleotide; (4) [5'-3']-ribonucleotide; (5) [5'-3']-3'-O-methyl ribonucleotide; (6) 3'-glyceryl; (7) [3'-5']-3'-deoxyribonucleotide; (8) [3'-3']-deoxyribonucleotide; (9) [5'-2']-deoxyribonucleotide; and (10) [5-3']-dideoxyribonucleotide. In addition to modified and unmodified backbone chemistries indicated in the figure, these chemistries can be combined with different backbone modifications as described herein, for example, backbone modifications having Formula I. In addition, the 2'-deoxy nucleotide shown 5' to the terminal modifications shown can be another modified or unmodified nucleotide or non-nucleotide described herein, for example modifications having any of Formulae I-VII or any combination thereof.

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Figure 11 shows a non-limiting example of a strategy used to identify chemically modified siNA constructs of the invention that are nuclease resistance while preserving the ability to mediate RNAi activity. Chemical modifications are introduced into the siNA construct based on educated design parameters (e.g. introducing 2'-mofications, base modifications, backbone modifications, terminal cap modifications etc). The modified construct in tested in an appropriate system (e.g. human serum for nuclease resistance, shown, or an animal model for PK/delivery parameters). In parallel, the siNA construct is tested for RNAi activity, for example in a cell culture system such as a luciferase reporter assay). Lead siNA constructs are then identified which possess a particular characteristic while maintaining RNAi activity, and can be further modified and assayed once again. This same approach can be used to identify siNA-conjugate molecules with improved pharmacokinetic profiles, delivery, and RNAi activity.

Figure 12 shows non-limiting examples of phosphorylated siNA molecules of the invention, including linear and duplex constructs and asymmetric derivatives thereof.

Figure 13 shows non-limiting examples of chemically modified terminal phosphate groups of the invention.

Figure 14A shows a non-limiting example of methodology used to design self complementary DFO constructs utilizing palidrome and/or repeat nucleic acid sequences that are identified in a target nucleic acid sequence. (i) A palindrome or repeat sequence is identified in a nucleic acid target sequence. (ii) A sequence is designed that is complementary to the target nucleic acid sequence and the palindrome sequence. (iii) An inverse repeat sequence of the non-palindrome/repeat portion of the complementary sequence is appended to the 3'-end of the complementary sequence to generate a self complementary DFO molecule comprising sequence complementary to the nucleic acid target. (iv) The DFO molecule can self-assemble to form a double stranded oligonucleotide. Figure 14B shows a non-limiting representative example of a duplex forming oligonucleotide sequence. Figure 14C shows a non-limiting example of the self assembly schematic of a representative duplex forming oligonucleotide sequence. Figure 14D shows a non-limiting example of the self assembly schematic of a representative duplex forming oligonucleotide sequence followed by interaction with a target nucleic acid sequence resulting in modulation of gene expression.

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Figure 15 shows a non-limiting example of the design of self complementary DFO constructs utilizing palidrome and/or repeat nucleic acid sequences that are incorporated into the DFO constructs that have sequence complementary to any target nucleic acid sequence of interest. Incorporation of these palindrome/repeat sequences allow the design of DFO constructs that form duplexes in which each strand is capable of mediating modulation of target gene expression, for example by RNAi. First, the target sequence is identified. A complementary sequence is then generated in which nucleotide or non-nucleotide modifications (shown as X or Y) are introduced into the complementary sequence that generate an artificial palindrome (shown as XYXYXY in the Figure). An inverse repeat of the non-palindrome/repeat complementary sequence is appended to the 3'-end of the complementary sequence to generate a self complementary DFO comprising sequence complementary to the nucleic acid target. The DFO can self-assemble to form a double stranded oligonucleotide.

Figure 16 shows non-limiting examples of multifunctional siNA molecules of the invention comprising two separate polynucleotide sequences that are each capable of mediating RNAi directed cleavage of differing target nucleic acid sequences. Figure 16A shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the first and second complementary regions are situated at the 3'-ends of each polynucleotide sequence in the multifunctional siNA. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences. Figure 16B shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the first and second complementary regions are situated at the 5'-ends of each polynucleotide sequence in the multifunctional siNA. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences.

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Figure 17 shows non-limiting examples of multifunctional siNA molecules of the invention comprising a single polynucleotide sequence comprising distinct regions that are each capable of mediating RNAi directed cleavage of differing target nucleic acid sequences. Figure 17A shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the second complementary region is situated at the 3'-end of the polynucleotide sequence in the multifunctional siNA. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences. Figure 17B shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the first complementary region is situated at the 5'-end of the polynucleotide sequence in the multifunctional siNA. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences. In one embodiment, these multifunctional siNA constructs are processed in vivo or in vitro to generate multifunctional siNA constructs as shown in Figure 16.

Figure 18 shows non-limiting examples of multifunctional siNA molecules of the invention comprising two separate polynucleotide sequences that are each capable of mediating RNAi directed cleavage of differing target nucleic acid sequences and wherein the multifunctional siNA construct further comprises a self complementary, palindrome, or repeat region, thus enabling shorter bifuctional siNA constructs that can mediate RNA interference against differing target nucleic acid sequences. Figure 18A shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the first and second complementary regions are situated at the 3'-ends of each polynucleotide sequence in the multifunctional siNA, and

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wherein the first and second complementary regions further comprise a self complementary, palindrome, or repeat region. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences. **Figure 18B** shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the first and second complementary regions are situated at the 5'-ends of each polynucleotide sequence in the multifunctional siNA, and wherein the first and second complementary regions further comprise a self complementary, palindrome, or repeat region. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences.

Figure 19 shows non-limiting examples of multifunctional siNA molecules of the invention comprising a single polynucleotide sequence comprising distinct regions that are each capable of mediating RNAi directed cleavage of differing target nucleic acid sequences and wherein the multifunctional siNA construct further comprises a self complementary, palindrome, or repeat region, thus enabling shorter bifuctional siNA constructs that can mediate RNA interference against differing target nucleic acid sequences. Figure 19A shows a non-limiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the second complementary region is situated at the 3'-end of the polynucleotide sequence in the multifunctional siNA, and wherein the first and second complementary regions further comprise a self complementary, palindrome, or repeat region. The dashed portions of each polynucleotide sequence of the multifunctional siNA construct have complementarity with regard to corresponding portions of the siNA duplex, but do not have complementarity to the target nucleic acid sequences. Figure 19B shows a nonlimiting example of a multifunctional siNA molecule having a first region that is complementary to a first target nucleic acid sequence (complementary region 1) and a

second region that is complementary to a second target nucleic acid sequence (complementary region 2), wherein the first complementary region is situated at the 5'end of the polynucleotide sequence in the multifunctional siNA, and wherein the first
and second complementary regions further comprise a self complementary, palindrome,
or repeat region. The dashed portions of each polynucleotide sequence of the
multifunctional siNA construct have complementarity with regard to corresponding
portions of the siNA duplex, but do not have complementarity to the target nucleic acid
sequences. In one embodiment, these multifunctional siNA constructs are processed in
vivo or in vitro to generate multifunctional siNA constructs as shown in Figure 18.

Figure 20 shows a non-limiting example of how multifunctional siNA molecules of the invention can target two separate target nucleic acid molecules, such as separate RNA molecules encoding differing proteins, for example, differing viral strains, a virus and a cellular protein involved in viral infection or replication, or differing proteins involved in a common or divergent biologic pathway that is implicated in the maintenance of progression of disease. Each strand of the multifunctional siNA construct comprises a region having complementarity to separate target nucleic acid molecules. The multifunctional siNA molecule is designed such that each strand of the siNA can be utilized by the RISC complex to initiate RNA interferance mediated cleavage of its corresponding target. These design parameters can include destabilization of each end of the siNA construct (see for example Schwarz et al., 2003, Cell, 115, 199-208). Such destabilization can be accomplished for example by using guanosine-cytidine base pairs, alternate base pairs (e.g., wobbles), or destabilizing chemically modified nucleotides at terminal nucleotide positions as is known in the art.

Figure 21 shows a non-limiting example of how multifunctional siNA molecules of the invention can target two separate target nucleic acid sequences within the same target nucleic acid molecule, such as alternate coding regions of a RNA, coding and non-coding regions of a RNA, or alternate splice variant regions of a RNA. Each strand of the multifunctional siNA construct comprises a region having complementarity to the separate regions of the target nucleic acid molecule. The multifunctional siNA molecule is designed such that each strand of the siNA can be utilized by the RISC complex to initiate RNA interferance mediated cleavage of its corresponding target region. These

design parameters can include destabilization of each end of the siNA construct (see for example Schwarz et al., 2003, Cell, 115, 199-208). Such destabilization can be accomplished for example by using guanosine-cytidine base pairs, alternate base pairs (e.g., wobbles), or destabilizing chemically modified nucleotides at terminal nucleotide positions as is known in the art.

DETAILED DESCRIPTION OF THE INVENTION

Mechanism of Action of Nucleic Acid Molecules of the Invention

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The discussion that follows discusses the proposed mechanism of RNA interference mediated by short interfering RNA as is presently known, and is not meant to be limiting and is not an admission of prior art. Applicant demonstrates herein that chemically-modified short interfering nucleic acids possess similar or improved capacity to mediate RNAi as do siRNA molecules and are expected to possess improved stability and activity *in vivo*; therefore, this discussion is not meant to be limiting only to siRNA and can be applied to siNA as a whole. By "improved capacity to mediate RNAi" or "improved RNAi activity" is meant to include RNAi activity measured *in vitro* and/or *in vivo* where the RNAi activity is a reflection of both the ability of the siNA to mediate RNAi and the stability of the siNAs of the invention. In this invention, the product of these activities can be increased *in vitro* and/or *in vivo* compared to an all RNA siRNA or a siNA containing a plurality of ribonucleotides. In some cases, the activity or stability of the siNA molecule can be decreased (i.e., less than ten-fold), but the overall activity of the siNA molecule is enhanced *in vitro* and/or *in vivo*.

RNA interference refers to the process of sequence specific post-transcriptional gene silencing in animals mediated by short interfering RNAs (siRNAs) (Fire *et al.*, 1998, *Nature*, 391, 806). The corresponding process in plants is commonly referred to as post-transcriptional gene silencing or RNA silencing and is also referred to as quelling in fungi. The process of post-transcriptional gene silencing is thought to be an evolutionarily-conserved cellular defense mechanism used to prevent the expression of foreign genes which is commonly shared by diverse flora and phyla (Fire *et al.*, 1999,

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Trends Genet., 15, 358). Such protection from foreign gene expression may have evolved in response to the production of double-stranded RNAs (dsRNAs) derived from viral infection or the random integration of transposon elements into a host genome via a cellular response that specifically destroys homologous single-stranded RNA or viral genomic RNA. The presence of dsRNA in cells triggers the RNAi response though a mechanism that has yet to be fully characterized. This mechanism appears to be different from the interferon response that results from dsRNA-mediated activation of protein kinase PKR and 2', 5'-oligoadenylate synthetase resulting in non-specific cleavage of mRNA by ribonuclease L.

The presence of long dsRNAs in cells stimulates the activity of a ribonuclease III enzyme referred to as Dicer. Dicer is involved in the processing of the dsRNA into short pieces of dsRNA known as short interfering RNAs (siRNAs) (Berstein et al., 2001, Nature, 409, 363). Short interfering RNAs derived from Dicer activity are typically about 21 to about 23 nucleotides in length and comprise about 19 base pair duplexes. Dicer has also been implicated in the excision of 21- and 22-nucleotide small temporal RNAs (stRNAs) from precursor RNA of conserved structure that are implicated in translational control (Hutvagner et al., 2001, Science, 293, 834). The RNAi response also features an endonuclease complex containing a siRNA, commonly referred to as an RNA-induced silencing complex (RISC), which mediates cleavage of single-stranded RNA having sequence homologous to the siRNA. Cleavage of the target RNA takes place in the middle of the region complementary to the guide sequence of the siRNA duplex (Elbashir et al., 2001, Genes Dev., 15, 188). In addition, RNA interference can also involve small RNA (e.g., micro-RNA or miRNA) mediated gene silencing, presumably though cellular mechanisms that regulate chromatin structure and thereby prevent transcription of target gene sequences (see for example Allshire, 2002, Science, 297, 1818-1819; Volpe et al., 2002, Science, 297, 1833-1837; Jenuwein, 2002, Science, 297, 2215-2218; and Hall et al., 2002, Science, 297, 2232-2237). As such, siNA molecules of the invention can be used to mediate gene silencing via interaction with RNA transcripts or alternately by interaction with particular gene sequences, wherein such interaction results in gene silencing either at the transcriptional level or posttranscriptional level.

RNAi has been studied in a variety of systems. Fire et al., 1998, Nature, 391, 806, were the first to observe RNAi in C. elegans. Wianny and Goetz, 1999, Nature Cell Biol., 2, 70, describe RNAi mediated by dsRNA in mouse embryos. Hammond et al., 2000, Nature, 404, 293, describe RNAi in Drosophila cells transfected with dsRNA. Elbashir et al., 2001, Nature, 411, 494, describe RNAi induced by introduction of duplexes of synthetic 21-nucleotide RNAs in cultured mammalian cells including human embryonic kidney and HeLa cells. Recent work in Drosophila embryonic lysates has revealed certain requirements for siRNA length, structure, chemical composition, and sequence that are essential to mediate efficient RNAi activity. These studies have shown that 21 nucleotide siRNA duplexes are most active when containing two 2-nucleotide 3'terminal nucleotide overhangs. Furthermore, substitution of one or both siRNA strands with 2'-deoxy or 2'-O-methyl nucleotides abolishes RNAi activity, whereas substitution of 3'-terminal siRNA nucleotides with deoxy nucleotides was shown to be tolerated. Mismatch sequences in the center of the siRNA duplex were also shown to abolish RNAi activity. In addition, these studies also indicate that the position of the cleavage site in the target RNA is defined by the 5'-end of the siRNA guide sequence rather than the 3'end (Elbashir et al., 2001, EMBO J., 20, 6877). Other studies have indicated that a 5'phosphate on the target-complementary strand of a siRNA duplex is required for siRNA activity and that ATP is utilized to maintain the 5'-phosphate moiety on the siRNA (Nykanen et al., 2001, Cell, 107, 309); however, siRNA molecules lacking a 5'phosphate are active when introduced exogenously, suggesting that 5'-phosphorylation of siRNA constructs may occur in vivo.

Synthesis of Nucleic acid Molecules

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Synthesis of nucleic acids greater than 100 nucleotides in length is difficult using automated methods, and the therapeutic cost of such molecules is prohibitive. In this invention, small nucleic acid motifs ("small" refers to nucleic acid motifs no more than 100 nucleotides in length, preferably no more than 80 nucleotides in length, and most preferably no more than 50 nucleotides in length; *e.g.*, individual siNA oligonucleotide sequences or siNA sequences synthesized in tandem) are preferably used for exogenous delivery. The simple structure of these molecules increases the ability of the nucleic acid

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to invade targeted regions of protein and/or RNA structure. Exemplary molecules of the instant invention are chemically synthesized, and others can similarly be synthesized.

Oligonucleotides (e.g., certain modified oligonucleotides or portions of oligonucleotides lacking ribonucleotides) are synthesized using protocols known in the art, for example as described in Caruthers et al., 1992, Methods in Enzymology 211, 3-19, Thompson et al., International PCT Publication No. WO 99/54459, Wincott et al., 1995, Nucleic Acids Res. 23, 2677-2684, Wincott et al., 1997, Methods Mol. Bio., 74, 59, Brennan et al., 1998, Biotechnol Bioeng., 61, 33-45, and Brennan, U.S. Pat. No. 6,001,311. All of these references are incorporated herein by reference. The synthesis of oligonucleotides makes use of common nucleic acid protecting and coupling groups, such as dimethoxytrityl at the 5'-end, and phosphoramidites at the 3'-end. In a nonlimiting example, small scale syntheses are conducted on a 394 Applied Biosystems, Inc. synthesizer using a 0.2 µmol scale protocol with a 2.5 min coupling step for 2'-Omethylated nucleotides and a 45 second coupling step for 2'-deoxy nucleotides or 2'deoxy-2'-fluoro nucleotides. Table V outlines the amounts and the contact times of the reagents used in the synthesis cycle. Alternatively, syntheses at the 0.2 µmol scale can be performed on a 96-well plate synthesizer, such as the instrument produced by Protogene (Palo Alto, CA) with minimal modification to the cycle. A 33-fold excess (60 μ L of 0.11 M = 6.6 μ mol) of 2'-O-methyl phosphoramidite and a 105-fold excess of Sethyl tetrazole (60 μ L of 0.25 M = 15 μ mol) can be used in each coupling cycle of 2'-Omethyl residues relative to polymer-bound 5'-hydroxyl. A 22-fold excess (40 µL of 0.11 $M = 4.4 \mu mol$) of deoxy phosphoramidite and a 70-fold excess of S-ethyl tetrazole (40 μ L of 0.25 M = 10 μ mol) can be used in each coupling cycle of deoxy residues relative to polymer-bound 5'-hydroxyl. Average coupling yields on the 394 Applied Biosystems, Inc. synthesizer, determined by colorimetric quantitation of the trityl fractions, are typically 97.5-99%. Other oligonucleotide synthesis reagents for the 394 Applied Biosystems, Inc. synthesizer include the following: detritylation solution is 3% TCA in methylene chloride (ABI); capping is performed with 16% N-methyl imidazole in THF (ABI) and 10% acetic anhydride/10% 2,6-lutidine in THF (ABI); and oxidation solution is 16.9 mM I₂, 49 mM pyridine, 9% water in THF (PerSeptive Biosystems, Inc.). Burdick & Jackson Synthesis Grade acetonitrile is used directly from the reagent bottle. S-Ethyltetrazole solution (0.25 M in acetonitrile) is made up from the solid obtained

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from American International Chemical, Inc. Alternately, for the introduction of phosphorothioate linkages, Beaucage reagent (3H-1,2-Benzodithiol-3-one 1,1-dioxide, 0.05 M in acetonitrile) is used.

Deprotection of the DNA-based oligonucleotides is performed as follows: the polymer-bound trityl-on oligoribonucleotide is transferred to a 4 mL glass screw top vial and suspended in a solution of 40% aqueous methylamine (1 mL) at 65 °C for 10 minutes. After cooling to -20 °C, the supernatant is removed from the polymer support. The support is washed three times with 1.0 mL of EtOH:MeCN:H2O/3:1:1, vortexed and the supernatant is then added to the first supernatant. The combined supernatants, containing the oligoribonucleotide, are dried to a white powder.

The method of synthesis used for RNA including certain siNA molecules of the invention follows the procedure as described in Usman et al., 1987, J. Am. Chem. Soc., 109, 7845; Scaringe et al., 1990, Nucleic Acids Res., 18, 5433; and Wincott et al., 1995, Nucleic Acids Res. 23, 2677-2684 Wincott et al., 1997, Methods Mol. Bio., 74, 59, and makes use of common nucleic acid protecting and coupling groups, such as dimethoxytrityl at the 5'-end, and phosphoramidites at the 3'-end. In a non-limiting example, small scale syntheses are conducted on a 394 Applied Biosystems, Inc. synthesizer using a 0.2 µmol scale protocol with a 7.5 min coupling step for alkylsilyl protected nucleotides and a 2.5 min coupling step for 2'-O-methylated nucleotides. Table V outlines the amounts and the contact times of the reagents used in the synthesis cycle. Alternatively, syntheses at the 0.2 µmol scale can be done on a 96-well plate synthesizer, such as the instrument produced by Protogene (Palo Alto, CA) with minimal modification to the cycle. A 33-fold excess (60 μ L of 0.11 M = 6.6 μ mol) of 2'-Omethyl phosphoramidite and a 75-fold excess of S-ethyl tetrazole (60 μ L of 0.25 M = 15 µmol) can be used in each coupling cycle of 2'-O-methyl residues relative to polymerbound 5'-hydroxyl. A 66-fold excess (120 μ L of 0.11 M = 13.2 μ mol) of alkylsilyl (ribo) protected phosphoramidite and a 150-fold excess of S-ethyl tetrazole (120 µL of 0.25 M = 30 µmol) can be used in each coupling cycle of ribo residues relative to polymerbound 5'-hydroxyl. Average coupling yields on the 394 Applied Biosystems, Inc. synthesizer, determined by colorimetric quantitation of the trityl fractions, are typically 97.5-99%. Other oligonucleotide synthesis reagents for the 394 Applied Biosystems,

Inc. synthesizer include the following: detritylation solution is 3% TCA in methylene chloride (ABI); capping is performed with 16% N-methyl imidazole in THF (ABI) and 10% acetic anhydride/10% 2,6-lutidine in THF (ABI); oxidation solution is 16.9 mM I₂, 49 mM pyridine, 9% water in THF (PerSeptive Biosystems, Inc.). Burdick & Jackson Synthesis Grade acetonitrile is used directly from the reagent bottle. S-Ethyltetrazole solution (0.25 M in acetonitrile) is made up from the solid obtained from American International Chemical, Inc. Alternately, for the introduction of phosphorothioate linkages, Beaucage reagent (3H-1,2-Benzodithiol-3-one 1,1-dioxide0.05 M in acetonitrile) is used.

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Deprotection of the RNA is performed using either a two-pot or one-pot protocol. For the two-pot protocol, the polymer-bound trityl-on oligoribonucleotide is transferred to a 4 mL glass screw top vial and suspended in a solution of 40% aq. methylamine (1 mL) at 65 °C for 10 min. After cooling to -20 °C, the supernatant is removed from the polymer support. The support is washed three times with 1.0 mL of EtOH:MeCN:H2O/3:1:1, vortexed and the supernatant is then added to the first supernatant. The combined supernatants, containing the oligoribonucleotide, are dried to a white powder. The base deprotected oligoribonucleotide is resuspended in anhydrous TEA/HF/NMP solution (300 μ L of a solution of 1.5 mL N-methylpyrrolidinone, 750 μ L TEA and 1 mL TEA•3HF to provide a 1.4 M HF concentration) and heated to 65 °C. After 1.5 h, the oligomer is quenched with 1.5 M NH₄HCO₃.

Alternatively, for the one-pot protocol, the polymer-bound trityl-on oligoribonucleotide is transferred to a 4 mL glass screw top vial and suspended in a solution of 33% ethanolic methylamine/DMSO: 1/1 (0.8 mL) at 65 °C for 15 minutes. The vial is brought to room temperature TEA•3HF (0.1 mL) is added and the vial is heated at 65 °C for 15 minutes. The sample is cooled at -20 °C and then quenched with $1.5 \, \mathrm{M} \, \mathrm{NH_4HCO_3}$.

For purification of the trityl-on oligomers, the quenched NH_4HCO_3 solution is loaded onto a C-18 containing cartridge that had been prewashed with acetonitrile followed by 50 mM TEAA. After washing the loaded cartridge with water, the RNA is detritylated with 0.5% TFA for 13 minutes. The cartridge is then washed again with

water, salt exchanged with 1 M NaCl and washed with water again. The oligonucleotide is then eluted with 30% acetonitrile.

The average stepwise coupling yields are typically >98% (Wincott *et al.*, 1995 *Nucleic Acids Res.* 23, 2677-2684). Those of ordinary skill in the art will recognize that the scale of synthesis can be adapted to be larger or smaller than the example described above including but not limited to 96-well format.

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Alternatively, the nucleic acid molecules of the present invention can be synthesized separately and joined together post-synthetically, for example, by ligation (Moore et al., 1992, Science 256, 9923; Draper et al., International PCT publication No. WO 93/23569; Shabarova et al., 1991, Nucleic Acids Research 19, 4247; Bellon et al., 1997, Nucleosides & Nucleotides, 16, 951; Bellon et al., 1997, Bioconjugate Chem. 8, 204), or by hybridization following synthesis and/or deprotection.

The siNA molecules of the invention can also be synthesized via a tandem synthesis methodology as described in Example 1 herein, wherein both siNA strands are synthesized as a single contiguous oligonucleotide fragment or strand separated by a cleavable linker which is subsequently cleaved to provide separate siNA fragments or strands that hybridize and permit purification of the siNA duplex. The linker can be a polynucleotide linker or a non-nucleotide linker. The tandem synthesis of siNA as described herein can be readily adapted to both multiwell/multiplate synthesis platforms such as 96 well or similarly larger multi-well platforms. The tandem synthesis of siNA as described herein can also be readily adapted to large scale synthesis platforms employing batch reactors, synthesis columns and the like.

A siNA molecule can also be assembled from two distinct nucleic acid strands or fragments wherein one fragment includes the sense region and the second fragment includes the antisense region of the RNA molecule.

The nucleic acid molecules of the present invention can be modified extensively to enhance stability by modification with nuclease resistant groups, for example, 2'-amino, 2'-C-allyl, 2'-fluoro, 2'-O-methyl, 2'-H (for a review see Usman and Cedergren, 1992, TIBS 17, 34; Usman et al., 1994, Nucleic Acids Symp. Ser. 31, 163). siNA constructs can

be purified by gel electrophoresis using general methods or can be purified by high pressure liquid chromatography (HPLC; see Wincott *et al.*, *supra*, the totality of which is hereby incorporated herein by reference) and re-suspended in water.

In another aspect of the invention, siNA molecules of the invention are expressed from transcription units inserted into DNA or RNA vectors. The recombinant vectors can be DNA plasmids or viral vectors. siNA expressing viral vectors can be constructed based on, but not limited to, adeno-associated virus, retrovirus, adenovirus, or alphavirus. The recombinant vectors capable of expressing the siNA molecules can be delivered as described herein, and persist in target cells. Alternatively, viral vectors can be used that provide for transient expression of siNA molecules.

Optimizing Activity of the nucleic acid molecule of the invention.

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Chemically synthesizing nucleic acid molecules with modifications (base, sugar and/or phosphate) can prevent their degradation by serum ribonucleases, which can increase their potency (see *e.g.*, Eckstein *et al.*, International Publication No. WO 92/07065; Perrault *et al.*, 1990 *Nature* 344, 565; Pieken *et al.*, 1991, *Science* 253, 314; Usman and Cedergren, 1992, *Trends in Biochem. Sci.* 17, 334; Usman *et al.*, International Publication No. WO 93/15187; and Rossi *et al.*, International Publication No. WO 91/03162; Sproat, U.S. Pat. No. 5,334,711; Gold *et al.*, U.S. Pat. No. 6,300,074; and Burgin *et al.*, *supra*; all of which are incorporated by reference herein). All of the above references describe various chemical modifications that can be made to the base, phosphate and/or sugar moieties of the nucleic acid molecules described herein. Modifications that enhance their efficacy in cells, and removal of bases from nucleic acid molecules to shorten oligonucleotide synthesis times and reduce chemical requirements are desired.

There are several examples in the art describing sugar, base and phosphate modifications that can be introduced into nucleic acid molecules with significant enhancement in their nuclease stability and efficacy. For example, oligonucleotides are modified to enhance stability and/or enhance biological activity by modification with nuclease resistant groups, for example, 2'-amino, 2'-C-allyl, 2'-fluoro, 2'-O-methyl, 2'-O-allyl, 2'-H, nucleotide base modifications (for a review see Usman and Cedergren, 1992,

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TIBS. 17, 34; Usman et al., 1994, Nucleic Acids Symp. Ser. 31, 163; Burgin et al., 1996, Biochemistry, 35, 14090). Sugar modification of nucleic acid molecules have been extensively described in the art (see Eckstein et al., International Publication PCT No. WO 92/07065; Perrault et al. Nature, 1990, 344, 565-568; Pieken et al. Science, 1991, 253, 314-317; Usman and Cedergren, Trends in Biochem. Sci., 1992, 17, 334-339; Usman et al. International Publication PCT No. WO 93/15187; Sproat, U.S. Pat. No. 5,334,711 and Beigelman et al., 1995, J. Biol. Chem., 270, 25702; Beigelman et al., International PCT publication No. WO 97/26270; Beigelman et al., U.S. Pat. No. 5,716,824; Usman et al., U.S. Pat. No. 5,627,053; Woolf et al., International PCT Publication No. WO 98/13526; Thompson et al., USSN 60/082,404 which was filed on April 20, 1998; Karpeisky et al., 1998, Tetrahedron Lett., 39, 1131; Earnshaw and Gait, 1998, Biopolymers (Nucleic Acid Sciences), 48, 39-55; Verma and Eckstein, 1998, Annu. Rev. Biochem., 67, 99-134; and Burlina et al., 1997, Bioorg. Med. Chem., 5, 1999-2010; all of the references are hereby incorporated in their totality by reference herein). Such publications describe general methods and strategies to determine the location of incorporation of sugar, base and/or phosphate modifications and the like into nucleic acid molecules without modulating catalysis, and are incorporated by reference herein. In view of such teachings, similar modifications can be used as described herein to modify the siNA nucleic acid molecules of the instant invention so long as the ability of siNA to promote RNAi is cells is not significantly inhibited.

While chemical modification of oligonucleotide internucleotide linkages with phosphorothioate, phosphorodithioate, and/or 5'-methylphosphonate linkages improves stability, excessive modifications can cause some toxicity or decreased activity. Therefore, when designing nucleic acid molecules, the amount of these internucleotide linkages should be minimized. The reduction in the concentration of these linkages should lower toxicity, resulting in increased efficacy and higher specificity of these molecules.

Short interfering nucleic acid (siNA) molecules having chemical modifications that maintain or enhance activity are provided. Such a nucleic acid is also generally more resistant to nucleases than an unmodified nucleic acid. Accordingly, the *in vitro* and/or *in vivo* activity should not be significantly lowered. In cases in which modulation is the

goal, therapeutic nucleic acid molecules delivered exogenously should optimally be stable within cells until translation of the target RNA has been modulated long enough to reduce the levels of the undesirable protein. This period of time varies between hours to days depending upon the disease state. Improvements in the chemical synthesis of RNA and DNA (Wincott et al., 1995, Nucleic Acids Res. 23, 2677; Caruthers et al., 1992, Methods in Enzymology 211, 3-19 (incorporated by reference herein)) have expanded the ability to modify nucleic acid molecules by introducing nucleotide modifications to enhance their nuclease stability, as described above.

In one embodiment, nucleic acid molecules of the invention include one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) G-clamp nucleotides. A G-clamp nucleotide is a modified cytosine analog wherein the modifications confer the ability to hydrogen bond both Watson-Crick and Hoogsteen faces of a complementary guanine within a duplex, see for example Lin and Matteucci, 1998, *J. Am. Chem. Soc.*, 120, 8531-8532. A single G-clamp analog substitution within an oligonucleotide can result in substantially enhanced helical thermal stability and mismatch discrimination when hybridized to complementary oligonucleotides. The inclusion of such nucleotides in nucleic acid molecules of the invention results in both enhanced affinity and specificity to nucleic acid targets, complementary sequences, or template strands. In another embodiment, nucleic acid molecules of the invention include one or more (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) LNA "locked nucleic acid" nucleotides such as a 2', 4'-C methylene bicyclo nucleotide (see for example Wengel et al., International PCT Publication No. WO 00/66604 and WO 99/14226).

In another embodiment, the invention features conjugates and/or complexes of siNA molecules of the invention. Such conjugates and/or complexes can be used to facilitate delivery of siNA molecules into a biological system, such as a cell. The conjugates and complexes provided by the instant invention can impart therapeutic activity by transferring therapeutic compounds across cellular membranes, altering the pharmacokinetics, and/or modulating the localization of nucleic acid molecules of the invention. The present invention encompasses the design and synthesis of novel conjugates and complexes for the delivery of molecules, including, but not limited to, small molecules, lipids, cholesterol, phospholipids, nucleosides, nucleotides, nucleic

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acids, antibodies, toxins, negatively charged polymers and other polymers, for example proteins, peptides, hormones, carbohydrates, polyethylene glycols, or polyamines, across cellular membranes. In general, the transporters described are designed to be used either individually or as part of a multi-component system, with or without degradable linkers. These compounds are expected to improve delivery and/or localization of nucleic acid molecules of the invention into a number of cell types originating from different tissues, in the presence or absence of serum (see Sullenger and Cech, U.S. Pat. No. 5,854,038). Conjugates of the molecules described herein can be attached to biologically active molecules via linkers that are biodegradable, such as biodegradable nucleic acid linker molecules.

The term "biodegradable linker" as used herein, refers to a nucleic acid or nonnucleic acid linker molecule that is designed as a biodegradable linker to connect one molecule to another molecule, for example, a biologically active molecule to a siNA molecule of the invention or the sense and antisense strands of a siNA molecule of the invention. The biodegradable linker is designed such that its stability can be modulated for a particular purpose, such as delivery to a particular tissue or cell type. The stability of a nucleic acid-based biodegradable linker molecule can be modulated by using various chemistries, for example combinations of ribonucleotides, deoxyribonucleotides, and chemically-modified nucleotides, such as 2'-O-methyl, 2'-fluoro, 2'-amino, 2'-O-amino, 2'-C-allyl, 2'-O-allyl, and other 2'-modified or base modified nucleotides. The biodegradable nucleic acid linker molecule can be a dimer, trimer, tetramer or longer nucleic acid molecule, for example, an oligonucleotide of about 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 nucleotides in length, or can comprise a single nucleotide with a phosphorus-based linkage, for example, a phosphoramidate or phosphodiester linkage. The biodegradable nucleic acid linker molecule can also comprise nucleic acid backbone, nucleic acid sugar, or nucleic acid base modifications.

The term "biodegradable" as used herein, refers to degradation in a biological system, for example enzymatic degradation or chemical degradation.

The term "biologically active molecule" as used herein, refers to compounds or molecules that are capable of eliciting or modifying a biological response in a system.

Non-limiting examples of biologically active siNA molecules either alone or in 104

combination with other molecules contemplated by the instant invention include therapeutically active molecules such as antibodies, cholesterol, hormones, antivirals, peptides, proteins, chemotherapeutics, small molecules, vitamins, co-factors, nucleosides, nucleotides, oligonucleotides, enzymatic nucleic acids, antisense nucleic acids, triplex forming oligonucleotides, 2,5-A chimeras, siNA, dsRNA, allozymes, aptamers, decoys and analogs thereof. Biologically active molecules of the invention also include molecules capable of modulating the pharmacokinetics and/or pharmacodynamics of other biologically active molecules, for example, lipids and polymers such as polyamines, polyamides, polyethylene glycol and other polyethers.

The term "phospholipid" as used herein, refers to a hydrophobic molecule comprising at least one phosphorus group. For example, a phospholipid can comprise a phosphorus-containing group and saturated or unsaturated alkyl group, optionally substituted with OH, COOH, oxo, amine, or substituted or unsubstituted aryl groups.

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Therapeutic nucleic acid molecules (e.g., siNA molecules) delivered exogenously optimally are stable within cells until reverse transcription of the RNA has been modulated long enough to reduce the levels of the RNA transcript. The nucleic acid molecules are resistant to nucleases in order to function as effective intracellular therapeutic agents. Improvements in the chemical synthesis of nucleic acid molecules described in the instant invention and in the art have expanded the ability to modify nucleic acid molecules by introducing nucleotide modifications to enhance their nuclease stability as described above.

In yet another embodiment, siNA molecules having chemical modifications that maintain or enhance enzymatic activity of proteins involved in RNAi are provided. Such nucleic acids are also generally more resistant to nucleases than unmodified nucleic acids. Thus, *in vitro* and/or *in vivo* the activity should not be significantly lowered.

Use of the nucleic acid-based molecules of the invention will lead to better treatment of the disease progression by affording the possibility of combination therapies (e.g., multiple siNA molecules targeted to different genes; nucleic acid molecules coupled with known small molecule modulators; or intermittent treatment with combinations of molecules, including different motifs and/or other chemical or

biological molecules). The treatment of subjects with siNA molecules can also include combinations of different types of nucleic acid molecules, such as enzymatic nucleic acid molecules (ribozymes), allozymes, antisense, 2,5-A oligoadenylate, decoys, and aptamers.

In another aspect a siNA molecule of the invention comprises one or more 5' and/or a 3'- cap structure, for example on only the sense siNA strand, the antisense siNA strand, or both siNA strands.

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By "cap structure" is meant chemical modifications, which have been incorporated at either terminus of the oligonucleotide (see, for example, Adamic et al., U.S. Pat. No. 5,998,203, incorporated by reference herein). These terminal modifications protect the nucleic acid molecule from exonuclease degradation, and may help in delivery and/or localization within a cell. The cap may be present at the 5'-terminus (5'-cap) or at the 3'terminal (3'-cap) or may be present on both termini. In non-limiting examples, the 5'-cap includes, but is not limited to, glyceryl, inverted deoxy abasic residue (moiety); 4',5'methylene nucleotide; 1-(beta-D-erythrofuranosyl) nucleotide, 4'-thio nucleotide; carbocyclic nucleotide; 1,5-anhydrohexitol nucleotide; L-nucleotides; alpha-nucleotides; modified base nucleotide; phosphorodithioate linkage; threo-pentofuranosyl nucleotide; acyclic 3',4'-seco nucleotide; acyclic 3,4-dihydroxybutyl nucleotide; acyclic 3,5dihydroxypentyl nucleotide, 3'-3'-inverted nucleotide moiety; 3'-3'-inverted abasic moiety; 3'-2'-inverted nucleotide moiety; 3'-2'-inverted abasic moiety; 1,4-butanediol phosphate; 3'-phosphoramidate; hexylphosphate; aminohexyl phosphate; 3'-phosphate; 3'-phosphorothioate; phosphorodithioate; or bridging or non-bridging methylphosphonate moiety.

Non-limiting examples of the 3'-cap include, but are not limited to, glyceryl, inverted deoxy abasic residue (moiety), 4', 5'-methylene nucleotide; 1-(beta-Derythrofuranosyl) nucleotide; 4'-thio nucleotide, carbocyclic nucleotide; 5'-amino-alkyl phosphate; 1,3-diamino-2-propyl phosphate; 3-aminopropyl phosphate; 6-aminohexyl phosphate; 1,2-aminododecyl phosphate; hydroxypropyl phosphate; 1,5-anhydrohexitol nucleotide; L-nucleotide; alpha-nucleotide; modified base nucleotide; phosphorodithioate; *threo*-pentofuranosyl nucleotide; acyclic 3',4'-seco nucleotide; 3,4-dihydroxybutyl nucleotide; 3,5-dihydroxypentyl nucleotide, 5'-5'-inverted nucleotide

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moiety; 5'-5'-inverted abasic moiety; 5'-phosphoramidate; 5'-phosphorothioate; 1,4-butanediol phosphate; 5'-amino; bridging and/or non-bridging 5'-phosphoramidate, phosphorothioate and/or phosphorodithioate, bridging or non bridging methylphosphonate and 5'-mercapto moieties (for more details see Beaucage and Iyer, 1993, *Tetrahedron* 49, 1925; incorporated by reference herein).

By the term "non-nucleotide" is meant any group or compound which can be incorporated into a nucleic acid chain in the place of one or more nucleotide units, including either sugar and/or phosphate substitutions, and allows the remaining bases to exhibit their enzymatic activity. The group or compound is abasic in that it does not contain a commonly recognized nucleotide base, such as adenosine, guanine, cytosine, uracil or thymine and therefore lacks a base at the 1'-position.

An "alkyl" group refers to a saturated aliphatic hydrocarbon, including straightchain, branched-chain, and cyclic alkyl groups. Preferably, the alkyl group has 1 to 12 carbons. More preferably, it is a lower alkyl of from 1 to 7 carbons, more preferably 1 to 4 carbons. The alkyl group can be substituted or unsubstituted. When substituted the substituted group(s) is preferably, hydroxyl, cyano, alkoxy, =O, =S, NO2 or N(CH3)2, amino, or SH. The term also includes alkenyl groups that are unsaturated hydrocarbon groups containing at least one carbon-carbon double bond, including straight-chain, branched-chain, and cyclic groups. Preferably, the alkenyl group has 1 to 12 carbons. More preferably, it is a lower alkenyl of from 1 to 7 carbons, more preferably 1 to 4 carbons. The alkenyl group may be substituted or unsubstituted. When substituted the substituted group(s) is preferably, hydroxyl, cyano, alkoxy, =0, =S, NO2, halogen, N(CH₃)₂, amino, or SH. The term "alkyl" also includes alkynyl groups that have an unsaturated hydrocarbon group containing at least one carbon-carbon triple bond, including straight-chain, branched-chain, and cyclic groups. Preferably, the alkynyl group has 1 to 12 carbons. More preferably, it is a lower alkynyl of from 1 to 7 carbons, more preferably 1 to 4 carbons. The alkynyl group may be substituted or unsubstituted. When substituted the substituted group(s) is preferably, hydroxyl, cyano, alkoxy, =O, =S, NO₂ or N(CH₃)₂, amino or SH.

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Such alkyl groups can also include aryl, alkylaryl, carbocyclic aryl, heterocyclic aryl, amide and ester groups. An "aryl" group refers to an aromatic group that has at least one ring having a conjugated pi electron system and includes carbocyclic aryl, heterocyclic aryl and biaryl groups, all of which may be optionally substituted. The preferred substituent(s) of aryl groups are halogen, trihalomethyl, hydroxyl, SH, OH, cyano, alkoxy, alkyl, alkenyl, alkynyl, and amino groups. An "alkylaryl" group refers to an alkyl group (as described above) covalently joined to an aryl group (as described above). Carbocyclic aryl groups are groups wherein the ring atoms on the aromatic ring are all carbon atoms. The carbon atoms are optionally substituted. Heterocyclic aryl groups are groups having from 1 to 3 heteroatoms as ring atoms in the aromatic ring and the remainder of the ring atoms are carbon atoms. Suitable heteroatoms include oxygen, sulfur, and nitrogen, and include furanyl, thienyl, pyridyl, pyrrolyl, N-lower alkyl pyrrolo, pyrimidyl, pyrazinyl, imidazolyl and the like, all optionally substituted. An "amide" refers to an -C(O)-NH-R, where R is either alkyl, aryl, alkylaryl or hydrogen. An "ester" refers to an -C(O)-OR', where R is either alkyl, aryl, alkylaryl or hydrogen.

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By "nucleotide" as used herein is as recognized in the art to include natural bases (standard), and modified bases well known in the art. Such bases are generally located at the 1' position of a nucleotide sugar moiety. Nucleotides generally comprise a base, sugar and a phosphate group. The nucleotides can be unmodified or modified at the sugar, phosphate and/or base moiety, (also referred to interchangeably as nucleotide analogs, modified nucleotides, non-natural nucleotides, non-standard nucleotides and other; see, for example, Usman and McSwiggen, supra; Eckstein et al., International PCT Publication No. WO 92/07065; Usman et al., International PCT Publication No. WO 93/15187; Uhlman & Peyman, supra, all are hereby incorporated by reference herein). There are several examples of modified nucleic acid bases known in the art as summarized by Limbach et al., 1994, Nucleic Acids Res. 22, 2183. Some of the nonlimiting examples of base modifications that can be introduced into nucleic acid molecules include, inosine, purine, pyridin-4-one, pyridin-2-one, phenyl, pseudouracil, 2, 4, 6-trimethoxy benzene, 3-methyl uracil, dihydrouridine, naphthyl, aminophenyl, 5-alkylcytidines (e.g., 5-methylcytidine), 5-alkyluridines (e.g., ribothymidine), 5-halouridine (e.g., 5-bromouridine) or 6-azapyrimidines or 6-alkylpyrimidines (e.g. 6methyluridine), propyne, and others (Burgin et al., 1996, Biochemistry, 35, 14090;

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Uhlman & Peyman, *supra*). By "modified bases" in this aspect is meant nucleotide bases other than adenine, guanine, cytosine and uracil at 1' position or their equivalents.

In one embodiment, the invention features modified siNA molecules, with phosphate backbone modifications comprising one or more phosphorothioate, phosphorodithioate, methylphosphonate, phosphotriester, morpholino, amidate carbamate, carboxymethyl, acetamidate, polyamide, sulfonate, sulfonamide, sulfamate, formacetal, thioformacetal, and/or alkylsilyl, substitutions. For a review of oligonucleotide backbone modifications, see Hunziker and Leumann, 1995, *Nucleic Acid Analogues: Synthesis and Properties*, in *Modern Synthetic Methods*, VCH, 331-417, and Mesmaeker et al., 1994, *Novel Backbone Replacements for Oligonucleotides*, in *Carbohydrate Modifications in Antisense Research*, ACS, 24-39.

By "abasic" is meant sugar moieties lacking a base or having other chemical groups in place of a base at the 1' position, see for example Adamic *et al.*, U.S. Pat. No. 5,998,203.

By "unmodified nucleoside" is meant one of the bases adenine, cytosine, guanine, thymine, or uracil joined to the 1' carbon of β -D-ribo-furanose.

By "modified nucleoside" is meant any nucleotide base which contains a modification in the chemical structure of an unmodified nucleotide base, sugar and/or phosphate. Non-limiting examples of modified nucleotides are shown by Formulae I-VII and/or other modifications described herein.

In connection with 2'-modified nucleotides as described for the present invention, by "amino" is meant 2'-NH₂ or 2'-O- NH₂, which can be modified or unmodified. Such modified groups are described, for example, in Eckstein *et al.*, U.S. Pat. No. 5,672,695 and Matulic-Adamic *et al.*, U.S. Pat. No. 6,248,878, which are both incorporated by reference in their entireties.

Various modifications to nucleic acid siNA structure can be made to enhance the utility of these molecules. Such modifications will enhance shelf-life, half-life *in vitro*, stability, and ease of introduction of such oligonucleotides to the target site, *e.g.*, to

enhance penetration of cellular membranes, and confer the ability to recognize and bind to targeted cells.

Administration of Nucleic Acid Molecules

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A siRNA molecule of the invention can be adapted for use to treat for example SARS virus infection, acute respiratory failure, viral pneumonia, and other indications that can respond to the level of SARS in a cell or tissue, alone or in combination with other therapies. For example, a siNA molecule can comprise a delivery vehicle. including liposomes, for administration to a subject, carriers and diluents and their salts, and/or can be present in pharmaceutically acceptable formulations. Methods for the delivery of nucleic acid molecules are described in Akhtar et al., 1992, Trends Cell Bio., 2, 139; Delivery Strategies for Antisense Oligonucleotide Therapeutics, ed. Akhtar, 1995, Maurer et al., 1999, Mol. Membr. Biol., 16, 129-140; Hofland and Huang, 1999, Handb. Exp. Pharmacol., 137, 165-192; and Lee et al., 2000, ACS Symp. Ser., 752, 184-192, all of which are incorporated herein by reference. Beigelman et al., U.S. Pat. No. 6,395,713 and Sullivan et al., PCT WO 94/02595 further describe the general methods for delivery of nucleic acid molecules. These protocols can be utilized for the delivery of virtually any nucleic acid molecule. Nucleic acid molecules can be administered to cells by a variety of methods known to those of skill in the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as biodegradable polymers, hydrogels, cyclodextrins (see for example Gonzalez et al., 1999, Bioconjugate Chem., 10, 1068-1074; Wang et al., International PCT publication Nos. WO 03/47518 and WO 03/46185), poly(lactic-co-glycolic)acid (PLGA) and PLCA microspheres (see for example US Patent 6,447,796 and US Patent Application Publication No. US 2002130430), biodegradable nanocapsules, and bioadhesive microspheres, or by proteinaceous vectors (O'Hare and Normand, International PCT Publication No. WO 00/53722). In another embodiment, the nucleic acid molecules of the invention can also be formulated or complexed with polyethyleneimine and derivatives thereof, such as polyethyleneiminepolyethyleneglycol-N-acetylgalactosamine (PEI-PEG-GAL) or polyethyleneiminepolyethyleneglycol-tri-N-acetylgalactosamine (PEI-PEG-triGAL) derivatives.

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Alternatively, the nucleic acid/vehicle combination is locally delivered by direct injection or by use of an infusion pump.

In one embodiment, the nucleic acid molecules or the invention are administered via pulmonary delivery, such as by inhalation of an aerosol or spray dried formulation administered by an inhalation device or nebulizer, providing rapid local uptake of the nucleic acid molecules into relevant pulmonary tissues. Solid particulate compositions containing respirable dry particles of micronized nucleic acid compositions can be prepared by grinding dried or lyophilized nucleic acid compositions, and then passing the micronized composition through, for example, a 400 mesh screen to break up or separate out large agglomerates. A solid particulate composition comprising the nucleic acid compositions of the invention can optionally contain a dispersant which serves to facilitate the formation of an aerosol as well as other therapeutic compounds. A suitable dispersant is lactose, which can be blended with the nucleic acid compound in any suitable ratio, such as a 1 to 1 ratio by weight.

Aerosols of liquid particles comprising a nucleic acid composition of the invention can be produced by any suitable means, such as with a nebulizer (see for example US 4,501,729). Nebulizers are commercially available devices which transform solutions or suspensions of an active ingredient into a therapeutic aerosol mist either by means of acceleration of a compressed gas, typically air or oxygen, through a narrow venturi orifice or by means of ultrasonic agitation. Suitable formulations for use in nebulizers comprise the active ingredient in a liquid carrier in an amount of up to 40% w/w preferably less than 20% w/w of the formulation. The carrier is typically water or a dilute aqueous alcoholic solution, preferably made isotonic with body fluids by the addition of, for example, sodium chloride or other suitable salts. Optional additives include preservatives if the formulation is not prepared sterile, for example, methyl hydroxybenzoate, anti-oxidants, flavorings, volatile oils, buffering agents and emulsifiers and other formulation surfactants. The aerosols of solid particles comprising the active composition and surfactant can likewise be produced with any solid particulate aerosol generator. Aerosol generators for administering solid particulate therapeutics to a subject produce particles which are respirable, as explained above, and generate a volume of aerosol containing a predetermined metered dose of a therapeutic composition at a rate

suitable for human administration. One illustrative type of solid particulate aerosol generator is an insufflator. Suitable formulations for administration by insufflation include finely comminuted powders which can be delivered by means of an insufflator. In the insufflator, the powder, e.g., a metered dose thereof effective to carry out the treatments described herein, is contained in capsules or cartridges, typically made of gelatin or plastic, which are either pierced or opened in situ and the powder delivered by air drawn through the device upon inhalation or by means of a manually-operated pump. The powder employed in the insufflator consists either solely of the active ingredient or of a powder blend comprising the active ingredient, a suitable powder diluent, such as lactose, and an optional surfactant. The active ingredient typically comprises from 0.1 to 100 w/w of the formulation. A second type of illustrative aerosol generator comprises a metered dose inhaler. Metered dose inhalers are pressurized aerosol dispensers, typically containing a suspension or solution formulation of the active ingredient in a liquified propellant. During use these devices discharge the formulation through a valve adapted to deliver a metered volume to produce a fine particle spray containing the active ingredient. Suitable propellants include certain chlorofluorocarbon compounds, for example, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane and mixtures thereof. The formulation can additionally contain one or more co-solvents, for example, ethanol, emulsifiers and other formulation surfactants, such as oleic acid or sorbitan trioleate, anti-oxidants and suitable flavoring agents. Other methods for pulmonary delivery are described in, for example US Patent Application No. 20040037780, and US Patent Nos. 6,592,904; 6,582,728; 6,565,885.

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In one embodiment, a siNA molecule of the invention is complexed with membrane disruptive agents such as those described in U.S. Patent Application Publication No. 20010007666, incorporated by reference herein in its entirety including the drawings. In another embodiment, the membrane disruptive agent or agents and the siNA molecule are also complexed with a cationic lipid or helper lipid molecule, such as those lipids described in U.S. Patent No. 6,235,310, incorporated by reference herein in its entirety including the drawings.

Thus, the invention features a pharmaceutical composition comprising one or more nucleic acid(s) of the invention in an acceptable carrier, such as a stabilizer, buffer, and

the like. The polynucleotides of the invention can be administered (e.g., RNA, DNA or protein) and introduced into a subject by any standard means, with or without stabilizers, buffers, and the like, to form a pharmaceutical composition. When it is desired to use a liposome delivery mechanism, standard protocols for formation of liposomes can be followed. The compositions of the present invention can also be formulated and used as tablets, capsules or elixirs for oral administration, suppositories for rectal administration, sterile solutions, suspensions for injectable administration, and the other compositions known in the art.

The present invention also includes pharmaceutically acceptable formulations of the compounds described. These formulations include salts of the above compounds, e.g., acid addition salts, for example, salts of hydrochloric, hydrobromic, acetic acid, and benzene sulfonic acid.

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A pharmacological composition or formulation refers to a composition or formulation in a form suitable for administration, e.g., systemic administration, into a cell or subject, including for example a human. Suitable forms, in part, depend upon the use or the route of entry, for example oral, transdermal, or by injection. Such forms should not prevent the composition or formulation from reaching a target cell (i.e., a cell to which the negatively charged nucleic acid is desirable for delivery). For example, pharmacological compositions injected into the blood stream should be soluble. Other factors are known in the art, and include considerations such as toxicity and forms that prevent the composition or formulation from exerting its effect.

By "systemic administration" is meant *in vivo* systemic absorption or accumulation of drugs in the blood stream followed by distribution throughout the entire body. Administration routes that lead to systemic absorption include, without limitation: intravenous, subcutaneous, intraperitoneal, inhalation, oral, intrapulmonary and intramuscular. Each of these administration routes exposes the siNA molecules of the invention to an accessible diseased tissue. The rate of entry of a drug into the circulation has been shown to be a function of molecular weight or size. The use of a liposome or other drug carrier comprising the compounds of the instant invention can potentially localize the drug, for example, in certain tissue types, such as the tissues of the reticular endothelial system (RES). A liposome formulation that can facilitate the

association of drug with the surface of cells, such as, lymphocytes and macrophages is also useful. This approach can provide enhanced delivery of the drug to target cells by taking advantage of the specificity of macrophage and lymphocyte immune recognition of abnormal cells, such as cells producing excess repeat expansion genes.

By "pharmaceutically acceptable formulation" is meant, a composition or formulation that allows for the effective distribution of the nucleic acid molecules of the instant invention in the physical location most suitable for their desired activity. Non-limiting examples of agents suitable for formulation with the nucleic acid molecules of the instant invention include: P-glycoprotein inhibitors (such as Pluronic P85),; biodegradable polymers, such as poly (DL-lactide-coglycolide) microspheres for sustained release delivery (Emerich, DF et al, 1999, Cell Transplant, 8, 47-58); and loaded nanoparticles, such as those made of polybutylcyanoacrylate. Other non-limiting examples of delivery strategies for the nucleic acid molecules of the instant invention include material described in Boado et al., 1998, J. Pharm. Sci., 87, 1308-1315; Tyler et al., 1999, FEBS Lett., 421, 280-284; Pardridge et al., 1995, PNAS USA., 92, 5592-5596; Boado, 1995, Adv. Drug Delivery Rev., 15, 73-107; Aldrian-Herrada et al., 1998, Nucleic Acids Res., 26, 4910-4916; and Tyler et al., 1999, PNAS USA., 96, 7053-7058.

The invention also features the use of the composition comprising surface-modified liposomes containing poly (ethylene glycol) lipids (PEG-modified, or long-circulating liposomes or stealth liposomes). These formulations offer a method for increasing the accumulation of drugs in target tissues. This class of drug carriers resists opsonization and elimination by the mononuclear phagocytic system (MPS or RES), thereby enabling longer blood circulation times and enhanced tissue exposure for the encapsulated drug (Lasic et al. Chem. Rev. 1995, 95, 2601-2627; Ishiwata et al., Chem. Pharm. Bull. 1995, 43, 1005-1011). Such liposomes have been shown to accumulate selectively in tumors, presumably by extravasation and capture in the neovascularized target tissues (Lasic et al., Science 1995, 267, 1275-1276; Oku et al., 1995, Biochim. Biophys. Acta, 1238, 86-90). The long-circulating liposomes enhance the pharmacokinetics and pharmacodynamics of DNA and RNA, particularly compared to conventional cationic liposomes which are known to accumulate in tissues of the MPS (Liu et al., J. Biol. Chem. 1995, 42, 24864-24870; Choi et al., International PCT

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Publication No. WO 96/10391; Ansell et al., International PCT Publication No. WO 96/10390; Holland et al., International PCT Publication No. WO 96/10392). Long-circulating liposomes are also likely to protect drugs from nuclease degradation to a greater extent compared to cationic liposomes, based on their ability to avoid accumulation in metabolically aggressive MPS tissues such as the liver and spleen.

The present invention also includes compositions prepared for storage or administration that include a pharmaceutically effective amount of the desired compounds in a pharmaceutically acceptable carrier or diluent. Acceptable carriers or diluents for therapeutic use are well known in the pharmaceutical art, and are described, for example, in *Remington's Pharmaceutical Sciences*, Mack Publishing Co. (A.R. Gennaro edit. 1985), hereby incorporated by reference herein. For example, preservatives, stabilizers, dyes and flavoring agents can be provided. These include sodium benzoate, sorbic acid and esters of *p*-hydroxybenzoic acid. In addition, antioxidants and suspending agents can be used.

A pharmaceutically effective dose is that dose required to prevent, inhibit the occurrence, or treat (alleviate a symptom to some extent, preferably all of the symptoms) of a disease state. The pharmaceutically effective dose depends on the type of disease, the composition used, the route of administration, the type of mammal being treated, the physical characteristics of the specific mammal under consideration, concurrent medication, and other factors that those skilled in the medical arts will recognize. Generally, an amount between 0.1 mg/kg and 100 mg/kg body weight/day of active ingredients is administered dependent upon potency of the negatively charged polymer.

The nucleic acid molecules of the invention and formulations thereof can be administered orally, topically, parenterally, by inhalation or spray, or rectally in dosage unit formulations containing conventional non-toxic pharmaceutically acceptable carriers, adjuvants and/or vehicles. The term parenteral as used herein includes percutaneous, subcutaneous, intravascular (e.g., intravenous), intramuscular, or intrathecal injection or infusion techniques and the like. In addition, there is provided a pharmaceutical formulation comprising a nucleic acid molecule of the invention and a pharmaceutically acceptable carrier. One or more nucleic acid molecules of the invention can be present in association with one or more non-toxic pharmaceutically

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acceptable carriers and/or diluents and/or adjuvants, and if desired other active ingredients. The pharmaceutical compositions containing nucleic acid molecules of the invention can be in a form suitable for oral use, for example, as tablets, troches, lozenges, aqueous or oily suspensions, dispersible powders or granules, emulsion, hard or soft capsules, or syrups or elixirs.

Compositions intended for oral use can be prepared according to any method known to the art for the manufacture of pharmaceutical compositions and such compositions can contain one or more such sweetening agents, flavoring agents, coloring agents or preservative agents in order to provide pharmaceutically elegant and palatable preparations. Tablets contain the active ingredient in admixture with non-toxic pharmaceutically acceptable excipients that are suitable for the manufacture of tablets. These excipients can be, for example, inert diluents; such as calcium carbonate, sodium carbonate, lactose, calcium phosphate or sodium phosphate; granulating and disintegrating agents, for example, corn starch, or alginic acid; binding agents, for example starch, gelatin or acacia; and lubricating agents, for example magnesium stearate, stearic acid or tale. The tablets can be uncoated or they can be coated by known techniques. In some cases such coatings can be prepared by known techniques to delay disintegration and absorption in the gastrointestinal tract and thereby provide a sustained action over a longer period. For example, a time delay material such as glyceryl monosterate or glyceryl distearate can be employed.

Formulations for oral use can also be presented as hard gelatin capsules wherein the active ingredient is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin, or as soft gelatin capsules wherein the active ingredient is mixed with water or an oil medium, for example peanut oil, liquid paraffin or olive oil.

Aqueous suspensions contain the active materials in a mixture with excipients suitable for the manufacture of aqueous suspensions. Such excipients are suspending agents, for example sodium carboxymethylcellulose, methylcellulose, hydropropylmethylcellulose, sodium alginate, polyvinylpyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents can be a naturally-occurring phosphatide, for example, lecithin, or condensation products of an alkylene oxide with fatty acids, for example polyoxyethylene stearate, or condensation products of ethylene oxide with long chain

aliphatic alcohols, for example heptadecaethyleneoxycetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with partial esters derived from fatty acids and hexitol anhydrides, for example polyethylene sorbitan monooleate. The aqueous suspensions can also contain one or more preservatives, for example ethyl, or n-propyl p-hydroxybenzoate, one or more coloring agents, one or more flavoring agents, and one or more sweetening agents, such as sucrose or saccharin.

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Oily suspensions can be formulated by suspending the active ingredients in a vegetable oil, for example arachis oil, olive oil, sesame oil or coconut oil, or in a mineral oil such as liquid paraffin. The oily suspensions can contain a thickening agent, for example beeswax, hard paraffin or cetyl alcohol. Sweetening agents and flavoring agents can be added to provide palatable oral preparations. These compositions can be preserved by the addition of an anti-oxidant such as ascorbic acid

Dispersible powders and granules suitable for preparation of an aqueous suspension by the addition of water provide the active ingredient in admixture with a dispersing or wetting agent, suspending agent and one or more preservatives. Suitable dispersing or wetting agents or suspending agents are exemplified by those already mentioned above. Additional excipients, for example sweetening, flavoring and coloring agents, can also be present.

Pharmaceutical compositions of the invention can also be in the form of oil-in-water emulsions. The oily phase can be a vegetable oil or a mineral oil or mixtures of these. Suitable emulsifying agents can be naturally-occurring gums, for example gum acacia or gum tragacanth, naturally-occurring phosphatides, for example soy bean, lecithin, and esters or partial esters derived from fatty acids and hexitol, anhydrides, for example sorbitan monooleate, and condensation products of the said partial esters with ethylene oxide, for example polyoxyethylene sorbitan monooleate. The emulsions can also contain sweetening and flavoring agents.

Syrups and elixirs can be formulated with sweetening agents, for example glycerol, propylene glycol, sorbitol, glucose or sucrose. Such formulations can also contain a

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demulcent, a preservative and flavoring and coloring agents. The pharmaceutical compositions can be in the form of a sterile injectable aqueous or oleaginous suspension. This suspension can be formulated according to the known art using those suitable dispersing or wetting agents and suspending agents that have been mentioned above. The sterile injectable preparation can also be a sterile injectable solution or suspension in a non-toxic parentally acceptable diluent or solvent, for example as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that can be employed are water, Ringer's solution and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose, any bland fixed oil can be employed including synthetic mono-or diglycerides. In addition, fatty acids such as oleic acid find use in the preparation of injectables.

The nucleic acid molecules of the invention can also be administered in the form of suppositories, e.g., for rectal administration of the drug. These compositions can be prepared by mixing the drug with a suitable non-irritating excipient that is solid at ordinary temperatures but liquid at the rectal temperature and will therefore melt in the rectum to release the drug. Such materials include cocoa butter and polyethylene glycols.

Nucleic acid molecules of the invention can be administered parenterally in a sterile medium. The drug, depending on the vehicle and concentration used, can either be suspended or dissolved in the vehicle. Advantageously, adjuvants such as local anesthetics, preservatives and buffering agents can be dissolved in the vehicle.

Dosage levels of the order of from about 0.1 mg to about 140 mg per kilogram of body weight per day are useful in the treatment of the above-indicated conditions (about 0.5 mg to about 7 g per subject per day). The amount of active ingredient that can be combined with the carrier materials to produce a single dosage form varies depending upon the host treated and the particular mode of administration. Dosage unit forms generally contain between from about 1 mg to about 500 mg of an active ingredient.

It is understood that the specific dose level for any particular subject depends upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, sex, diet, time of administration, route of administration,

and rate of excretion, drug combination and the severity of the particular disease undergoing therapy.

For administration to non-human animals, the composition can also be added to the animal feed or drinking water. It can be convenient to formulate the animal feed and drinking water compositions so that the animal takes in a therapeutically appropriate quantity of the composition along with its diet. It can also be convenient to present the composition as a premix for addition to the feed or drinking water.

The nucleic acid molecules of the present invention can also be administered to a subject in combination with other therapeutic compounds to increase the overall therapeutic effect. The use of multiple compounds to treat an indication can increase the beneficial effects while reducing the presence of side effects.

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In one embodiment, the invention comprises compositions suitable for administering nucleic acid molecules of the invention to specific cell types. example, the asialoglycoprotein receptor (ASGPr) (Wu and Wu, 1987, J. Biol. Chem. 262, 4429-4432) is unique to hepatocytes and binds branched galactose-terminal glycoproteins, such as asialoorosomucoid (ASOR). In another example, the folate receptor is overexpressed in many cancer cells. Binding of such glycoproteins, synthetic glycoconjugates, or folates to the receptor takes place with an affinity that strongly depends on the degree of branching of the oligosaccharide chain, for example, triatennary structures are bound with greater affinity than biatenarry or monoatennary chains (Baenziger and Fiete, 1980, Cell, 22, 611-620; Connolly et al., 1982, J. Biol. Chem., 257, 939-945). Lee and Lee, 1987, Glycoconjugate J., 4, 317-328, obtained this high specificity through the use of N-acetyl-D-galactosamine as the carbohydrate moiety, which has higher affinity for the receptor, compared to galactose. This "clustering effect" has also been described for the binding and uptake of mannosyl-terminating glycoproteins or glycoconjugates (Ponpipom et al., 1981, J. Med. Chem., 24, 1388-1395). The use of galactose, galactosamine, or folate based conjugates to transport exogenous compounds across cell membranes can provide a targeted delivery approach to, for example, the treatment of liver disease, cancers of the liver, or other cancers. The use of bioconjugates can also provide a reduction in the required dose of therapeutic compounds required for treatment. Furthermore, therapeutic bioavialability,

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pharmacodynamics, and pharmacokinetic parameters can be modulated through the use of nucleic acid bioconjugates of the invention. Non-limiting examples of such bioconjugates are described in Vargeese et al., USSN 10/201,394, filed August 13, 2001; and Matulic-Adamic et al., USSN 10/151,116, filed May 17, 2002. In one embodiment, nucleic acid molecules of the invention are complexed with or covalently attached to nanoparticles, such as Hepatitis B virus S, M, or L evelope proteins (see for example Yamado et al., 2003, Nature Biotechnology, 21, 885). In one embodiment, nucleic acid molecules of the invention are delivered with specificity for human tumor cells, specifically non-apoptotic human tumor cells including for example T-cells, hepatocytes, breast carcinoma cells, ovarian carcinoma cells, melanoma cells, intestinal epithelial cells, prostate cells, testicular cells, non-small cell lung cancers, small cell lung cancers, etc.

Alternatively, certain siNA molecules of the instant invention can be expressed within cells from eukaryotic promoters (e.g., Izant and Weintraub, 1985, Science, 229, 15 345; McGarry and Lindquist, 1986, Proc. Natl. Acad. Sci., USA 83, 399; Scanlon et al., 1991, Proc. Natl. Acad. Sci. USA, 88, 10591-5; Kashani-Sabet et al., 1992, Antisense Res. Dev., 2, 3-15; Dropulic et al., 1992, J. Virol., 66, 1432-41; Weerasinghe et al., 1991, J. Virol., 65, 5531-4; Ojwang et al., 1992, Proc. Natl. Acad. Sci. USA, 89, 10802-6; Chen et al., 1992, Nucleic Acids Res., 20, 4581-9; Sarver et al., 1990 Science, 247, 1222-1225; Thompson et al., 1995, Nucleic Acids Res., 23, 2259; Good et al., 1997, 20 Gene Therapy, 4, 45. Those skilled in the art realize that any nucleic acid can be expressed in eukaryotic cells from the appropriate DNA/RNA vector. The activity of such nucleic acids can be augmented by their release from the primary transcript by a enzymatic nucleic acid (Draper et al., PCT WO 93/23569, and Sullivan et al., PCT WO 94/02595; Ohkawa et al., 1992, Nucleic Acids Symp. Ser., 27, 15-6; Taira et al., 1991, 25 Nucleic Acids Res., 19, 5125-30; Ventura et al., 1993, Nucleic Acids Res., 21, 3249-55; Chowrira et al., 1994, J. Biol. Chem., 269, 25856.

In another aspect of the invention, RNA molecules of the present invention can be expressed from transcription units (see for example Couture *et al.*, 1996, *TIG.*, 12, 510) inserted into DNA or RNA vectors. The recombinant vectors can be DNA plasmids or viral vectors. siNA expressing viral vectors can be constructed based on, but not limited

to, adeno-associated virus, retrovirus, adenovirus, or alphavirus. In another embodiment, pol III based constructs are used to express nucleic acid molecules of the invention (see for example Thompson, U.S. Pats. Nos. 5,902,880 and 6,146,886). The recombinant vectors capable of expressing the siNA molecules can be delivered as described above, and persist in target cells. Alternatively, viral vectors can be used that provide for transient expression of nucleic acid molecules. Such vectors can be repeatedly administered as necessary. Once expressed, the siNA molecule interacts with the target mRNA and generates an RNAi response. Delivery of siNA molecule expressing vectors can be systemic, such as by intravenous or intra-muscular administration, by administration to target cells ex-planted from a subject followed by reintroduction into the subject, or by any other means that would allow for introduction into the desired target cell (for a review see Couture et al., 1996, TIG., 12, 510).

In one aspect the invention features an expression vector comprising a nucleic acid sequence encoding at least one siNA molecule of the instant invention. The expression vector can encode one or both strands of a siNA duplex, or a single self-complementary strand that self hybridizes into a siNA duplex. The nucleic acid sequences encoding the siNA molecules of the instant invention can be operably linked in a manner that allows expression of the siNA molecule (see for example Paul et al., 2002, Nature Biotechnology, 19, 505; Miyagishi and Taira, 2002, Nature Biotechnology, 19, 497; Lee et al., 2002, Nature Biotechnology, 19, 500; and Novina et al., 2002, Nature Medicine, advance online publication doi:10.1038/nm725).

In another aspect, the invention features an expression vector comprising: a) a transcription initiation region (e.g., eukaryotic pol I, II or III initiation region); b) a transcription termination region (e.g., eukaryotic pol I, II or III termination region); and c) a nucleic acid sequence encoding at least one of the siNA molecules of the instant invention, wherein said sequence is operably linked to said initiation region and said termination region in a manner that allows expression and/or delivery of the siNA molecule. The vector can optionally include an open reading frame (ORF) for a protein operably linked on the 5' side or the 3'-side of the sequence encoding the siNA of the invention; and/or an intron (intervening sequences).

Transcription of the siNA molecule sequences can be driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters are expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type depends on the nature of the gene regulatory sequences (enhancers, silencers, etc.) present nearby. Prokaryotic 5 RNA polymerase promoters are also used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells (Elroy-Stein and Moss, 1990, Proc. Natl. Acad. Sci. U S A, 87, 6743-7; Gao and Huang 1993, Nucleic Acids Res., 21, 2867-72; Lieber et al., 1993, Methods Enzymol., 217, 47-66; Zhou et al., 1990, Mol. 10 Cell. Biol., 10, 4529-37). Several investigators have demonstrated that nucleic acid molecules expressed from such promoters can function in mammalian cells (e.g. Kashani-Sabet et al., 1992, Antisense Res. Dev., 2, 3-15; Ojwang et al., 1992, Proc. Natl. Acad. Sci. USA, 89, 10802-6; Chen et al., 1992, Nucleic Acids Res., 20, 4581-9; Yu et al., 1993, Proc. Natl. Acad. Sci. USA, 90, 6340-4; L'Huillier et al., 1992, EMBO J., 11, 4411-8; Lisziewicz et al., 1993, Proc. Natl. Acad. Sci. U. S. A, 90, 8000-4; 15 Thompson et al., 1995, Nucleic Acids Res., 23, 2259; Sullenger & Cech, 1993, Science, 262, 1566). More specifically, transcription units such as the ones derived from genes encoding U6 small nuclear (snRNA), transfer RNA (tRNA) and adenovirus VA RNA are useful in generating high concentrations of desired RNA molecules such as siNA in cells (Thompson et al., supra; Couture and Stinchcomb, 1996, supra; Noonberg et al., 1994, 20 Nucleic Acid Res., 22, 2830; Noonberg et al., U.S. Pat. No. 5,624,803; Good et al., 1997, Gene Ther., 4, 45; Beigelman et al., International PCT Publication No. WO 96/18736. The above siNA transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated virus vectors), or viral RNA 25 vectors (such as retroviral or alphavirus vectors) (for a review see Couture and Stinchcomb, 1996, supra).

In another aspect the invention features an expression vector comprising a nucleic acid sequence encoding at least one of the siNA molecules of the invention in a manner that allows expression of that siNA molecule. The expression vector comprises in one embodiment; a) a transcription initiation region; b) a transcription termination region; and c) a nucleic acid sequence encoding at least one strand of the siNA molecule,

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wherein the sequence is operably linked to the initiation region and the termination region in a manner that allows expression and/or delivery of the siNA molecule.

In another embodiment the expression vector comprises: a) a transcription initiation region; b) a transcription termination region; c) an open reading frame; and d) a nucleic acid sequence encoding at least one strand of a siNA molecule, wherein the sequence is operably linked to the 3'-end of the open reading frame and wherein the sequence is operably linked to the initiation region, the open reading frame and the termination region in a manner that allows expression and/or delivery of the siNA molecule. In yet another embodiment, the expression vector comprises: a) a transcription initiation region; b) a transcription termination region; c) an intron; and d) a nucleic acid sequence encoding at least one siNA molecule, wherein the sequence is operably linked to the initiation region, the intron and the termination region in a manner which allows expression and/or delivery of the nucleic acid molecule.

In another embodiment, the expression vector comprises: a) a transcription initiation region; b) a transcription termination region; c) an intron; d) an open reading frame; and e) a nucleic acid sequence encoding at least one strand of a siNA molecule, wherein the sequence is operably linked to the 3'-end of the open reading frame and wherein the sequence is operably linked to the initiation region, the intron, the open reading frame and the termination region in a manner which allows expression and/or delivery of the siNA molecule.

SARS virus biology and biochemistry

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The following discussion is adapted from the report, "Preliminary Clinical Description of Severe Acute Respiratory Syndrome", World Health Organization, Geneva, Switzerland, available at the Centers for Disease Control and Prevention website.

Severe acute respiratory syndrome (SARS) is a viral respiratory illness caused by a coronavirus, called SARS-associated coronavirus (SARS-CoV). SARS was first reported in Asia in February 2003. Over the next few months, the illness spread to more than two dozen countries in North America, South America, Europe, and Asia before the SARS

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global outbreak of 2003 was contained. According to the World Health Organization (WHO), a total of 8,098 people worldwide became sick with SARS during the 2003 outbreak. Of these, 774 died.

The incubation period for SARS is typically 2--7 days; however, isolated reports have suggested an incubation period as long as 10 days. The illness begins generally with a prodrome of fever (>100.4°F [>38.0°C]). Fever often is high, sometimes is associated with chills and rigors, and might be accompanied by other symptoms, including headache, malaise, and myalgia. At the onset of illness, some persons have mild respiratory symptoms. Typically, rash and neurologic or gastrointestinal findings are absent; however, some patients have reported diarrhea during the febrile prodrome.

After 3--7 days, a lower respiratory phase begins with the onset of a dry, nonproductive cough or dyspnea, which might be accompanied by or progress to hypoxemia. In 10%--20% of cases, the respiratory illness is severe enough to require intubation and mechanical ventilation. Death may result from progressive respiratory failure due to alveolar damage. The case-fatality rate among persons with illness meeting the current WHO case definition of SARS is approximately 3%.

Chest radiographs might be normal during the febrile prodrome and throughout the course of illness. However, in a substantial proportion of patients, the respiratory phase is characterized by early focal interstitial infiltrates progressing to more generalized, patchy, interstitial infiltrates. Some chest radiographs from patients in the late stages of SARS also have shown areas of consolidation.

Early in the course of disease, the absolute lymphocyte count is often decreased. Overall white blood cell counts have generally been normal or decreased. At the peak of the respiratory illness, approximately 50% of patients have leukopenia and thrombocytopenia or low-normal platelet counts (50,000--150,000/μL). Early in the respiratory phase, elevated creatine phosphokinase levels (as high as 3,000 IU/L) and hepatic transaminases (two to six times the upper limits of normal) have been noted. In the majority of patients, renal function has remained normal.

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The severity of illness might be highly variable, ranging from mild illness to death. Although a few close contacts of patients with SARS have developed a similar illness, the majority have remained well. Some close contacts have reported a mild, febrile illness without respiratory signs or symptoms, suggesting the illness might not always progress to the respiratory phase.

Treatment regimens have included several antibiotics to presumptively treat known bacterial agents of atypical pneumonia. In several locations, therapy also has included antiviral agents such as oseltamivir or ribavirin. Steroids have also been administered orally or intravenously to patients in combination with ribavirin and other antimicrobials. At present, the most efficacious treatment regimen, if any, is unknown.

The causative agent of SARS appears to be a novel coronavirus that was isolated from patients who met the case definition of SARS (see Ksiazek et al., 2003, New England Journal of Medicine, 10.1056/NEJMoa030781. Indirect fluorescent antibody tests and enzyme-linked immunosorbent assays made with the new coronavirus isolate have been used to demonstrate a virus-specific serologic response. Amplification of short regions of the polymerase gene, (the most strongly conserved part of the Coronavirus genome) by reverse transcriptase polymerase chain reaction (RT-PCR) and nucleotide sequencing revealed that the SARS virus is a novel Coronavirus which has not previously been present in human populations. This conclusion is confirmed by serological (antigenic) investigations. The sequence of the SARS associated coronavirus was recently made available through the CDC.

Viral entry into cells occurs via endocytosis and membrane fusion. Replication occurs in the cytoplasm. Initially, the 5' 20kb of the (+)sense genome is translated to produce a viral polymerase, which then produces a full-length (-)sense strand. This is used as a template to produce mRNA as a nested set of transcripts, all with an identical 5' non-translated leader sequence of 72nt and coincident 3' polyadenylated ends. Each mRNA is monocistronic, the genes at the 5' end being translated from the longest mRNA. These unusual cytoplasmic structures are produced not by splicing but by the polymerase during transcription. Between each of the genes there is a repeated intergenic sequence - UCUAAAC - which interacts with the transcriptase plus cellular factors to splice the leader sequence onto the start of each ORF. Viral assembly occurs by budding

into the golgi apparatus, and viral particles are transported to the surface of the cell and are subsequently released.

The SARS virus can be grown in Vero cells (a fibroblast cell line isolated in 1962 from a primate). This is a novel property for human cornaviruses which usually cannot be cultivated. In these cells, virus infection results in a cytopathic effect, and budding of Coronavirus-like particles from the endoplasmic reticulum within infected cells.

Detection of the SARS virus can be accomplished with serological testing and molecular diagnotic procedures. Serological testing for anti-Coronavirus antibodies consists of indirect fluorescent antibody testing and enzyme-linked immunosorbent assays (ELISA) which detect antibodies against the virus produced in response to infection. Molecular testing consists of reverse transcriptase-polymerase chain reaction (RT-PCR) tests specific for the RNA from the novel Coronavirus.

The use of small interfering nucleic acid molecules targeting SARS genes therefore provides a class of novel therapeutic agents that can be used in the treatment and diagnosis of SARS virus infection, acute respiratory failure, viral pneumonia, or any other disease or condition that responds to modulation of SARS genes.

Examples:

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The following are non-limiting examples showing the selection, isolation, synthesis and activity of nucleic acids of the instant invention.

20 Example 1: Tandem synthesis of siNA constructs

Exemplary siNA molecules of the invention are synthesized in tandem using a cleavable linker, for example, a succinyl-based linker. Tandem synthesis as described herein is followed by a one-step purification process that provides RNAi molecules in high yield. This approach is highly amenable to siNA synthesis in support of high throughput RNAi screening, and can be readily adapted to multi-column or multi-well synthesis platforms.

After completing a tandem synthesis of a siNA oligo and its complement in which the 5'-terminal dimethoxytrityl (5'-O-DMT) group remains intact (trityl on synthesis), the

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oligonucleotides are deprotected as described above. Following deprotection, the siNA sequence strands are allowed to spontaneously hybridize. This hybridization yields a duplex in which one strand has retained the 5'-O-DMT group while the complementary strand comprises a terminal 5'-hydroxyl. The newly formed duplex behaves as a single molecule during routine solid-phase extraction purification (Trityl-On purification) even though only one molecule has a dimethoxytrityl group. Because the strands form a stable duplex, this dimethoxytrityl group (or an equivalent group, such as other trityl groups or other hydrophobic moieties) is all that is required to purify the pair of oligos, for example, by using a C18 cartridge.

Standard phosphoramidite synthesis chemistry is used up to the point of introducing a tandem linker, such as an inverted deoxy abasic succinate or glyceryl succinate linker (see **Figure 1**) or an equivalent cleavable linker. A non-limiting example of linker coupling conditions that can be used includes a hindered base such as diisopropylethylamine (DIPA) and/or DMAP in the presence of an activator reagent such as Bromotripyrrolidinophosphoniumhexaflurorophosphate (PyBrOP). After the linker is coupled, standard synthesis chemistry is utilized to complete synthesis of the second sequence leaving the terminal the 5'-O-DMT intact. Following synthesis, the resulting oligonucleotide is deprotected according to the procedures described herein and quenched with a suitable buffer, for example with 50mM NaOAc or 1.5M NH4H2CO3.

Purification of the siNA duplex can be readily accomplished using solid phase extraction, for example using a Waters C18 SepPak 1g cartridge conditioned with 1 column volume (CV) of acetonitrile, 2 CV H2O, and 2 CV 50mM NaOAc. The sample is loaded and then washed with 1 CV H2O or 50mM NaOAc. Failure sequences are eluted with 1 CV 14% ACN (Aqueous with 50mM NaOAc and 50mM NaCl). The column is then washed, for example with 1 CV H2O followed by on-column detritylation, for example by passing 1 CV of 1% aqueous trifluoroacetic acid (TFA) over the column, then adding a second CV of 1% aqueous TFA to the column and allowing to stand for approximately 10 minutes. The remaining TFA solution is removed and the column washed with H2O followed by 1 CV 1M NaCl and additional H2O. The siNA duplex product is then eluted, for example, using 1 CV 20% aqueous CAN.

Figure 2 provides an example of MALDI-TOF mass spectrometry analysis of a purified siNA construct in which each peak corresponds to the calculated mass of an individual siNA strand of the siNA duplex. The same purified siNA provides three peaks when analyzed by capillary gel electrophoresis (CGE), one peak presumably corresponding to the duplex siNA, and two peaks presumably corresponding to the separate siNA sequence strands. Ion exchange HPLC analysis of the same siNA contract only shows a single peak. Testing of the purified siNA construct using a luciferase reporter assay described below demonstrated the same RNAi activity compared to siNA constructs generated from separately synthesized oligonucleotide sequence strands.

10 Example 2: Identification of potential siNA target sites in any RNA sequence

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The sequence of an RNA target of interest, such as a viral or human mRNA transcript, is screened for target sites, for example by using a computer folding algorithm. In a non-limiting example, the sequence of a gene or RNA gene transcript derived from a database, such as Genbank, is used to generate siNA targets having complementarity to the target. Such sequences can be obtained from a database, or can be determined experimentally as known in the art. Target sites that are known, for example, those target sites determined to be effective target sites based on studies with other nucleic acid molecules, for example ribozymes or antisense, or those targets known to be associated with a disease or condition such as those sites containing mutations or deletions, can be used to design siNA molecules targeting those sites. parameters can be used to determine which sites are the most suitable target sites within the target RNA sequence. These parameters include but are not limited to secondary or tertiary RNA structure, the nucleotide base composition of the target sequence, the degree of homology between various regions of the target sequence, or the relative position of the target sequence within the RNA transcript. Based on these determinations, any number of target sites within the RNA transcript can be chosen to screen siNA molecules for efficacy, for example by using in vitro RNA cleavage assays, cell culture, or animal models. In a non-limiting example, anywhere from 1 to 1000 target sites are chosen within the transcript based on the size of the siNA construct to be used. High throughput screening assays can be developed for screening siNA molecules

using methods known in the art, such as with multi-well or multi-plate assays to determine efficient reduction in target gene expression.

Example 3: Selection of siNA molecule target sites in a RNA

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The following non-limiting steps can be used to carry out the selection of siNAs targeting a given gene sequence or transcript.

- 1. The target sequence is parsed *in silico* into a list of all fragments or subsequences of a particular length, for example 23 nucleotide fragments, contained within the target sequence. This step is typically carried out using a custom Perl script, but commercial sequence analysis programs such as Oligo, MacVector, or the GCG Wisconsin Package can be employed as well.
- 2. In some instances the siNAs correspond to more than one target sequence; such would be the case for example in targeting different transcripts of the same gene, targeting different transcripts of more than one gene, or for targeting both the human gene and an animal homolog. In this case, a subsequence list of a particular length is generated for each of the targets, and then the lists are compared to find matching sequences in each list. The subsequences are then ranked according to the number of target sequences that contain the given subsequence; the goal is to find subsequences that are present in most or all of the target sequences. Alternately, the ranking can identify subsequences that are unique to a target sequence, such as a mutant target sequence. Such an approach would enable the use of siNA to target specifically the mutant sequence and not effect the expression of the normal sequence.
- 3. In some instances the siNA subsequences are absent in one or more sequences while present in the desired target sequence; such would be the case if the siNA targets a gene with a paralogous family member that is to remain untargeted. As in case 2 above, a subsequence list of a particular length is generated for each of the targets, and then the lists are compared to find sequences that are present in the target gene but are absent in the untargeted paralog.

4. The ranked siNA subsequences can be further analyzed and ranked according to GC content. A preference can be given to sites containing 30-70% GC, with a further preference to sites containing 40-60% GC.

- The ranked siNA subsequences can be further analyzed and ranked according to self-folding and internal hairpins. Weaker internal folds are preferred; strong hairpin structures are to be avoided.
 - 6. The ranked siNA subsequences can be further analyzed and ranked according to whether they have runs of GGG or CCC in the sequence. GGG (or even more Gs) in either strand can make oligonucleotide synthesis problematic and can potentially interfere with RNAi activity, so it is avoided whenever better sequences are available. CCC is searched in the target strand because that will place GGG in the antisense strand.

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- 7. The ranked siNA subsequences can be further analyzed and ranked according to whether they have the dinucleotide UU (uridine dinucleotide) on the 3'-end of the sequence, and/or AA on the 5'-end of the sequence (to yield 3' UU on the antisense sequence). These sequences allow one to design siNA molecules with terminal TT thymidine dinucleotides.
- 8. Four or five target sites are chosen from the ranked list of subsequences as described above. For example, in subsequences having 23 nucleotides, the right 21 nucleotides of each chosen 23-mer subsequence are then designed and synthesized for the upper (sense) strand of the siNA duplex, while the reverse complement of the left 21 nucleotides of each chosen 23-mer subsequence are then designed and synthesized for the lower (antisense) strand of the siNA duplex (see **Tables II and III**). If terminal TT residues are desired for the sequence (as described in paragraph 7), then the two 3' terminal nucleotides of both the sense and antisense strands are replaced by TT prior to synthesizing the oligos.
 - 9. The siNA molecules are screened in an *in vitro*, cell culture or animal model system to identify the most active siNA molecule or the most preferred target site within the target RNA sequence.

10. Other design considerations can be used when selecting target nucleic acid sequences, see for example Reynolds *et al.*, 2004, *Nature Biotechnology Advanced Online Publication*, 1 February 2004, doi:10.1038/nbt936 and Ui-Tei et al., 2004, Nucleic Acids Research, 32, doi:10.1093/nar/gkh247.

In an alternate approach, a pool of siNA constructs specific to a SARS target sequence is used to screen for target sites in cells expressing SARS RNA, such as VERO cells and/or FRhk-4 cells. The general strategy used in this approach is shown in Figure 9. A non-limiting example of such is a pool comprising sequences having SEQ ID NOs: 1-3392. Cells expressing SARS (e.g., VERO cells and/or FRhk-4 cells) are transfected with the pool of siNA constructs and cells that demonstrate a phenotype associated with SARS inhibition are sorted. The pool of siNA constructs can be expressed from transcription cassettes inserted into appropriate vectors (see for example Figure 7 and Figure 8). The siNA from cells demonstrating a positive phenotypic change (e.g., decreased proliferation, decreased SARS mRNA levels or decreased SARS protein expression), are sequenced to determine the most suitable target site(s) within the target SARS RNA sequence.

Example 4: SARS targeted siNA design

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siNA target sites were chosen by analyzing sequences of the SARS RNA target and optionally prioritizing the target sites on the basis of folding (structure of any given sequence analyzed to determine siNA accessibility to the target), by using a library of siNA molecules as described in Example 3, or alternately by using an *in vitro* siNA system as described in Example 6 herein. siNA molecules were designed that could bind each target and are optionally individually analyzed by computer folding to assess whether the siNA molecule can interact with the target sequence. Varying the length of the siNA molecules can be chosen to optimize activity. Generally, a sufficient number of complementary nucleotide bases are chosen to bind to, or otherwise interact with, the target RNA, but the degree of complementarity can be modulated to accommodate siNA duplexes or varying length or base composition. By using such methodologies, siNA molecules can be designed to target sites within any known RNA sequence, for example those RNA sequences corresponding to the any gene transcript.

Chemically modified siNA constructs are designed to provide nuclease stability for systemic administration in vivo and/or improved pharmacokinetic, localization, and delivery properties while preserving the ability to mediate RNAi activity. Chemical modifications as described herein are introduced synthetically using synthetic methods described herein and those generally known in the art. The synthetic siNA constructs are then assayed for nuclease stability in serum and/or cellular/tissue extracts (e.g. liver extracts). The synthetic siNA constructs are also tested in parallel for RNAi activity using an appropriate assay, such as a luciferase reporter assay as described herein or another suitable assay that can quantity RNAi activity. Synthetic siNA constructs that possess both nuclease stability and RNAi activity can be further modified and reevaluated in stability and activity assays. The chemical modifications of the stabilized active siNA constructs can then be applied to any siNA sequence targeting any chosen RNA and used, for example, in target screening assays to pick lead siNA compounds for therapeutic development (see for example Figure 11).

15 Example 5: Chemical Synthesis and Purification of siNA

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siNA molecules can be designed to interact with various sites in the RNA message, for example, target sequences within the RNA sequences described herein. The sequence of one strand of the siNA molecule(s) is complementary to the target site sequences described above. The siNA molecules can be chemically synthesized using methods described herein. Inactive siNA molecules that are used as control sequences can be synthesized by scrambling the sequence of the siNA molecules such that it is not complementary to the target sequence. Generally, siNA constructs can by synthesized using solid phase oligonucleotide synthesis methods as described herein (see for example Usman *et al.*, US Patent Nos. 5,804,683; 5,831,071; 5,998,203; 6,117,657; 6,353,098; 6,362,323; 6,437,117; 6,469,158; Scaringe *et al.*, US Patent Nos. 6,111,086; 6,008,400; 6,111,086 all incorporated by reference herein in their entirety).

In a non-limiting example, RNA oligonucleotides are synthesized in a stepwise fashion using the phosphoramidite chemistry as is known in the art. Standard phosphoramidite chemistry involves the use of nucleosides comprising any of 5'-O-dimethoxytrityl, 2'-O-tert-butyldimethylsilyl, 3'-O-2-Cyanoethyl N,N-diisopropylphosphoroamidite groups, and exocyclic amine protecting groups (e.g. N6-benzoyl adenosine,

N4 acetyl cytidine, and N2-isobutyryl guanosine). Alternately, 2'-O-Silyl Ethers can be used in conjunction with acid-labile 2'-O-orthoester protecting groups in the synthesis of RNA as described by Scaringe *supra*. Differing 2' chemistries can require different protecting groups, for example 2'-deoxy-2'-amino nucleosides can utilize N-phthaloyl protection as described by Usman *et al.*, US Patent 5,631,360, incorporated by reference herein in its entirety).

During solid phase synthesis, each nucleotide is added sequentially (3'- to 5'-direction) to the solid support-bound oligonucleotide. The first nucleoside at the 3'-end of the chain is covalently attached to a solid support (e.g., controlled pore glass or polystyrene) using various linkers. The nucleotide precursor, a ribonucleoside phosphoramidite, and activator are combined resulting in the coupling of the second nucleoside phosphoramidite onto the 5'-end of the first nucleoside. The support is then washed and any unreacted 5'-hydroxyl groups are capped with a capping reagent such as acetic anhydride to yield inactive 5'-acetyl moieties. The trivalent phosphorus linkage is then oxidized to a more stable phosphate linkage. At the end of the nucleotide addition cycle, the 5'-O-protecting group is cleaved under suitable conditions (e.g., acidic conditions for trityl-based groups and Fluoride for silyl-based groups). The cycle is repeated for each subsequent nucleotide.

Modification of synthesis conditions can be used to optimize coupling efficiency, for example by using differing coupling times, differing reagent/phosphoramidite concentrations, differing contact times, differing solid supports and solid support linker chemistries depending on the particular chemical composition of the siNA to be synthesized. Deprotection and purification of the siNA can be performed as is generally described in Deprotection and purification of the siNA can be performed as is generally described in Usman *et al.*, US 5,831,071, US 6,353,098, US 6,437,117, and Bellon *et al.*, US 6,054,576, US 6,162,909, US 6,303,773, or Scaringe *supra*, incorporated by reference herein in their entireties. Additionally, deprotection conditions can be modified to provide the best possible yield and purity of siNA constructs. For example, applicant has observed that oligonucleotides comprising 2'-deoxy-2'-fluoro nucleotides can degrade under inappropriate deprotection conditions. Such oligonucleotides are deprotected using aqueous methylamine at about 35°C for 30 minutes. If the 2'-deoxy-

2'-fluoro containing oligonucleotide also comprises ribonucleotides, after deprotection with aqueous methylamine at about 35°C for 30 minutes, TEA-HF is added and the reaction maintained at about 65°C for an additional 15 minutes.

Example 6: RNAi in vitro assay to assess siNA activity

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An in vitro assay that recapitulates RNAi in a cell-free system is used to evaluate siNA constructs targeting SARS RNA targets. The assay comprises the system described by Tuschl et al., 1999, Genes and Development, 13, 3191-3197 and Zamore et al., 2000, Cell, 101, 25-33 adapted for use with SARS target RNA. A Drosophila extract derived from syncytial blastoderm is used to reconstitute RNAi activity in vitro. Target RNA is generated via in vitro transcription from an appropriate SARS expressing plasmid using T7 RNA polymerase or via chemical synthesis as described herein. Sense and antisense siNA strands (for example 20 uM each) are annealed by incubation in buffer (such as 100 mM potassium acetate, 30 mM HEPES-KOH, pH 7.4, 2 mM magnesium acetate) for 1 minute at 90°C followed by 1 hour at 37°C, then diluted in lysis buffer (for example 100 mM potassium acetate, 30 mM HEPES-KOH at pH 7.4. 2mM magnesium acetate). Annealing can be monitored by gel electrophoresis on an agarose gel in TBE buffer and stained with ethidium bromide. The Drosophila lysate is prepared using zero to two-hour-old embryos from Oregon R flies collected on yeasted molasses agar that are dechorionated and lysed. The lysate is centrifuged and the supernatant isolated. The assay comprises a reaction mixture containing 50% lysate [vol/vol], RNA (10-50 pM final concentration), and 10% [vol/vol] lysis buffer containing siNA (10 nM final concentration). The reaction mixture also contains 10 mM creatine phosphate, 10 ug.ml creatine phosphokinase, 100 um GTP, 100 uM UTP, 100 uM CTP, 500 uM ATP, 5 mM DTT, 0.1 U/uL RNasin (Promega), and 100 uM of each amino acid. The final concentration of potassium acetate is adjusted to 100 mM. The reactions are pre-assembled on ice and preincubated at 25° C for 10 minutes before adding RNA, then incubated at 25° C for an additional 60 minutes. Reactions are quenched with 4 volumes of 1.25 x Passive Lysis Buffer (Promega). Target RNA cleavage is assayed by RT-PCR analysis or other methods known in the art and are compared to control reactions in which siNA is omitted from the reaction.

Alternately, internally-labeled target RNA for the assay is prepared by *in vitro* transcription in the presence of [alpha-³²p] CTP, passed over a G 50 Sephadex column by spin chromatography and used as target RNA without further purification. Optionally, target RNA is 5'-³²P-end labeled using T4 polynucleotide kinase enzyme. Assays are performed as described above and target RNA and the specific RNA cleavage products generated by RNAi are visualized on an autoradiograph of a gel. The percentage of cleavage is determined by PHOSPHOR IMAGER® (autoradiography) quantitation of bands representing intact control RNA or RNA from control reactions without siNA and the cleavage products generated by the assay.

In one embodiment, this assay is used to determine target sites the SARS RNA target for siNA mediated RNAi cleavage, wherein a plurality of siNA constructs are screened for RNAi mediated cleavage of the SARS RNA target, for example, by analyzing the assay reaction by electrophoresis of labeled target RNA, or by northern blotting, as well as by other methodology well known in the art.

15 Example 7: Nucleic acid inhibition of SARS target RNA in vitro

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siNA molecules targeted to the human SARS RNA are designed and synthesized as described above: These nucleic acid molecules can be tested for cleavage activity in vivo, for example, using the following procedure. The target sequences and the nucleotide location within the SARS RNA are given in **Table II and III**.

Two formats are used to test the efficacy of siNAs targeting SARS. First, the reagents are tested in cell culture using, for example, VERO cells and/or FRhk-4 cells, to determine the extent of RNA and protein inhibition. siNA reagents (e.g.; see Tables II and III) are selected against the SARS target as described herein. RNA inhibition is measured after delivery of these reagents by a suitable transfection agent to, for example, VERO cells and/or FRhk-4 cells. Relative amounts of target RNA are measured versus actin using real-time PCR monitoring of amplification (eg., ABI 7700 TAQMAN®). A comparison is made to a mixture of oligonucleotide sequences made to unrelated targets or to a randomized siNA control with the same overall length and chemistry, but randomly substituted at each position. Primary and secondary lead reagents are chosen for the target and optimization performed. After an optimal transfection agent

concentration is chosen, a RNA time-course of inhibition is performed with the lead siNA molecule. In addition, a cell-plating format can be used to determine RNA inhibition.

Delivery of siNA to Cells

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Cells (e.g., VERO cells and/or FRhk-4 cells infected with the SARS virus) are seeded, for example, at 1x10⁵ cells per well of a six-well dish in EGM-2 (BioWhittaker) the day before transfection. siNA (final concentration, for example 20nM) and cationic lipid (e.g., final concentration 2µg/ml) are complexed in EGM basal media (Bio Whittaker) at 37°C for 30 minutes in polystyrene tubes. Following vortexing, the complexed siNA is added to each well and incubated for the times indicated. For initial optimization experiments, cells are seeded, for example, at 1x10³ in 96 well plates and siNA complex added as described. Efficiency of delivery of siNA to cells is determined using a fluorescent siNA complexed with lipid. Cells in 6-well dishes are incubated with siNA for 24 hours, rinsed with PBS and fixed in 2% paraformaldehyde for 15 minutes at room temperature. Uptake of siNA is visualized using a fluorescent microscope.

TAQMAN® (real-time PCR monitoring of amplification) and Lightcycler quantification of mRNA

Total RNA is prepared from cells following siNA delivery, for example, using Qiagen RNA purification kits for 6-well or Rneasy extraction kits for 96-well assays. For TAQMAN® analysis (real-time PCR monitoring of amplification), dual-labeled probes are synthesized with the reporter dye, FAM or JOE, covalently linked at the 5'-end and the quencher dye TAMRA conjugated to the 3'-end. One-step RT-PCR amplifications are performed on, for example, an ABI PRISM 7700 Sequence Detector using 50 μl reactions consisting of 10 μl total RNA, 100 nM forward primer, 900 nM reverse primer, 100 nM probe, 1X TAQMAN® PCR reaction buffer (PE-Applied Biosystems), 5.5 mM MgCl₂, 300 μM each dATP, dCTP, dGTP, and dTTP, 10U RNase Inhibitor (Promega), 1.25U AMPLITAQ GOLD® (DNA polymerase) (PE-Applied Biosystems) and 10U M-MLV Reverse Transcriptase (Promega). The thermal cycling conditions can consist of 30 minutes at 48°C, 10 minutes at 95°C, followed by 40 cycles of 15 seconds at 95°C and 1 minute at 60°C. Quantitation of mRNA levels is determined relative to standards

generated from serially diluted total cellular RNA (300, 100, 33, 11 ng/rxn) and normalizing to \(\beta\)-actin or GAPDH mRNA in parallel TAQMAN® reactions (real-time PCR monitoring of amplification). For each gene of interest an upper and lower primer and a fluorescently labeled probe are designed. Real time incorporation of SYBR Green I dye into a specific PCR product can be measured in glass capillary tubes using a lightcyler. A standard curve is generated for each primer pair using control cRNA. Values are represented as relative expression to GAPDH in each sample.

Western blotting

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Nuclear extracts can be prepared using a standard micro preparation technique (see for example Andrews and Faller, 1991, *Nucleic Acids Research*, 19, 2499). Protein extracts from supernatants are prepared, for example using TCA precipitation. An equal volume of 20% TCA is added to the cell supernatant, incubated on ice for 1 hour and pelleted by centrifugation for 5 minutes. Pellets are washed in acetone, dried and resuspended in water. Cellular protein extracts are run on a 10% Bis-Tris NuPage (nuclear extracts) or 4-12% Tris-Glycine (supernatant extracts) polyacrylamide gel and transferred onto nitro-cellulose membranes. Non-specific binding can be blocked by incubation, for example, with 5% non-fat milk for 1 hour followed by primary antibody for 16 hour at 4°C. Following washes, the secondary antibody is applied, for example (1:10,000 dilution) for 1 hour at room temperature and the signal detected with SuperSignal reagent (Pierce).

Example 8: RNAi mediated inhibition of SARS RNA expression

siNA constructs (e.g., siNA constructs shown in **Table III**) are tested for efficacy in reducing SARS RNA expression in, for example, VERO cells and/or FRhk-4 cells. Cells are plated approximately 24h before transfection in 96-well plates at 5,000-7,500 cells/well, 100 μl/well, such that at the time of transfection cells are 70-90% confluent. For transfection, annealed siNAs are mixed with the transfection reagent (Lipofectamine 2000, Invitrogen) in a volume of 50 μl/well and incubated for 20 minutes at room temperature. The siNA transfection mixtures are added to cells to give a final siNA concentration of 25 nM in a volume of 150 μl. Each siNA transfection mixture is added to 3 wells for triplicate siNA treatments. Cells are incubated at 37° for 24h in the

continued presence of the siNA transfection mixture. At 24h, RNA is prepared from each well of treated cells. The supernatants with the transfection mixtures are first removed and discarded, then the cells are lysed and RNA prepared from each well. Target gene expression following treatment is evaluated by RT-PCR for the target gene and for a control gene (36B4, an RNA polymerase subunit) for normalization. The triplicate data is averaged and the standard deviations determined for each treatment. Normalized data are graphed and the percent reduction of target mRNA by active siNAs in comparison to their respective inverted control siNAs is determined.

In a non-limiting example, a siNA construct comprising ribonucleotides and 3'-terminal dithymidine caps is assayed along with a chemically modified siNA construct comprising 2'-deoxy-2'-fluoro pyrimidine nucleotides and purine ribonucleotides in which the sense strand of the siNA is further modified with 5' and 3'-terminal inverted deoxyabasic caps and the antisense strand comprises a 3'-terminal phosphorothioate internucleotide linkage. Additional stabilization chemistries as described in Table IV are similarly assayed for activity. These siNA constructs are compared to appropriate matched chemistry inverted controls. In addition, the siNA constructs are also compared to untreated cells, cells transfected with lipid and scrambled siNA constructs, and cells transfected with lipid alone (transfection control).

Example 9: Animal Models

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Evaluating the efficacy of anti-SARS agents in animal models is an important prerequisite to human clinical trials. Byron et al., 2003, Nature, 425, 915, describe ferret and feline animal models of SARS virus infection. Haagmans et al., 2004, Nature Medicine, 10, 290-293, describe the use of pegylated interferon-alpha in protecting type 1 pneumocytes against SARS coronavirus infection in macaques. Gao et al., 2003, Lancet, 362, 1895-6, describe the use of a SARS virus vaccine in monkeys. All of these models can be adapted for use for pre-clinical evaluation of the efficacy of nucleic acid compositions of the invetention in modulating SARS virus gene expression toward therapeutic use.

Example 10: Indications

The present body of knowledge in SARS research indicates the need for methods to assay SARS activity and for compounds that can regulate SARS expression for research, diagnostic, and therapeutic use. As described herein, the nucleic acid molecules of the present invention can be used in assays to diagnose disease state related of SARS levels. In addition, the nucleic acid molecules can be used to treat disease state related to SARS levels.

Particular degenerative and disease states that can be associated with SARS expression modulation include, but are not limited to, SARS virus infection, liver failure, hepatocellular carcinoma, cirrhosis, and/or other disease states associated with SARS virus infection.

Immunomodulators, steroids, and anti-vrial compounds are non-limiting examples of pharmaceutical agents that can be combined with or used in conjunction with the nucleic acid molecules (e.g. siNA molecules) of the instant invention. The use of ribavirin and oseltamivir are non-limiting examples of chemotherapeutic agents that can be combined with or used in conjunction with the nucleic acid molecules (e.g. siNA molecules) of the instant invention. Those skilled in the art will recognize that other anti-cancer compounds and therapies can similarly be readily combined with the nucleic acid molecules of the instant invention (e.g. siNA molecules) and are hence within the scope of the instant invention.

20 Example 11: Interferons

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Interferons represent a non-limiting example of a class of compounds that can be used in conjuction with the siNA molecules of the invention for treating the diseases and/or conditions described herein. Type I interferons (IFN) are a class of natural cytokines that includes a family of greater than 25 IFN- α (Pesta, 1986, *Methods Enzymol.* 119, 3-14) as well as IFN- β , and IFN- ω . Although evolutionarily derived from the same gene (Diaz *et al.*, 1994, *Genomics* 22, 540-552), there are many differences in the primary sequence of these molecules, implying an evolutionary divergence in biologic activity. All type I IFN share a common pattern of biologic effects that begin with binding of the IFN to the cell surface receptor (Pfeffer & Strulovici, 1992, Transmembrane secondary messengers for IFN- α/β . In: *Interferon. Principles and*

Medical Applications., S. Baron, D.H. Coopenhaver, F. Dianzani, W.R. Fleischmann Jr., T.K. Hughes Jr., G.R. Kimpel, D.W. Niesel, G.J. Stanton, and S.K. Tyring, eds. 151-160). Binding is followed by activation of tyrosine kinases, including the Janus tyrosine kinases and the STAT proteins, which leads to the production of several IFN-stimulated gene products (Johnson et al., 1994, Sci. Am. 270, 68-75). The IFN-stimulated gene 5 products are responsible for the pleotropic biologic effects of type I IFN, including antiviral, antiproliferative, and immunomodulatory effects, cytokine induction, and HLA class I and class II regulation (Pestka et al., 1987, Annu. Rev. Biochem 56, 727). Examples of IFN-stimulated gene products include 2-5-oligoadenylate synthetase (2-5 10 OAS), \(\beta_2\)-microglobulin, neopterin, p68 kinases, and the Mx protein (Chebath & Revel, 1992, The 2-5 A system: 2-5 A synthetase, isospecies and functions. In: Interferon. Principles and Medical Applications, S. Baron, D.H. Coopenhaver, F. Dianzani, W.R. Jr. Fleischmann, T.K. Jr Hughes, G.R. Kimpel, D.W. Niesel, G.J. Stanton, and S.K. Tyring, eds., pp. 225-236; Samuel, 1992, The RNA-dependent P1/eIF-2α protein kinase. In: 15 Interferon. Principles and Medical Applications. S. Baron, D.H. Coopenhayer, F. Dianzani, W.R. Fleischmann Jr., T.K. Hughes Jr., G.R. Kimpel, D.W. Niesel, G.H. Stanton, and S.K. Tyring, eds. 237-250; Horisberger, 1992, MX protein: function and Mechanism of Action. In: Interferon. Principles and Medical Applications. S. Baron, D.H. Coopenhaver, F. Dianzani, W.R. Fleischmann Jr., T.K. Hughes Jr., G.R. Kimpel, 20 D.W. Niesel, G.H. Stanton, and S.K. Tyring, eds. 215-224). Although all type I IFN have similar biologic effects, not all the activities are shared by each type I IFN, and in many cases, the extent of activity varies quite substantially for each IFN subtype (Fish et al, 1989, J. Interferon Res. 9, 97-114; Ozes et al., 1992, J. Interferon Res. 12, 55-59). More specifically, investigations into the properties of different subtypes of IFN- α and molecular hybrids of IFN-α have shown differences in pharmacologic properties 25 (Rubinstein, 1987, J. Interferon Res. 7, 545-551). These pharmacologic differences can arise from as few as three amino acid residue changes (Lee et al., 1982, Cancer Res. 42, 1312-1316).

Eighty-five to 166 amino acids are conserved in the known IFN- α subtypes. Excluding the IFN- α pseudogenes, there are approximately 25 known distinct IFN- α subtypes. Pairwise comparisons of these nonallelic subtypes show primary sequence

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differences ranging from 2% to 23%. In addition to the naturally occurring IFNs, a non-natural recombinant type I interferon known as consensus interferon (CIFN) has been synthesized as a therapeutic compound (Tong et al., 1997, Hepatology 26, 747-754).

Interferon is currently in use for at least 12 different indications, including infectious and autoimmune diseases and cancer (Borden, 1992, *N. Engl. J. Med.* 326, 1491-1492). For autoimmune diseases, IFN has been utilized for treatment of rheumatoid arthritis, multiple sclerosis, and Crohn's disease. For treatment of cancer, IFN has been used alone or in combination with a number of different compounds. Specific types of cancers for which IFN has been used include squamous cell carcinomas, melanomas, hypernephromas, hemangiomas, hairy cell leukemia, and Kaposi's sarcoma. In the treatment of infectious diseases, IFNs increase the phagocytic activity of macrophages and cytotoxicity of lymphocytes and inhibits the propagation of cellular pathogens. Specific indications for which IFN has been used as treatment include hepatitis B, human papillomavirus types 6 and 11 (i.e. genital warts) (Leventhal et al., 1991, N Engl J Med 325, 613-617), chronic granulomatous disease, and SARS virus.

Pegylated interferons, i.e., interferons conjugated with polyethylene glycol (PEG), have demonstrated improved characteristics over interferon. Advantages incurred by PEG conjugation can include an improved pharmacokinetic profile compared to interferons lacking PEG, thus imparting more convenient dosing regimes, improved tolerance, and improved antiviral efficacy. Such improvements have been demonstrated in clinical studies of both polyethylene glycol interferon alfa-2a (PEGASYS, Roche) and polyethylene glycol interferon alfa-2b (VIRAFERON PEG, PEG-INTRON, Enzon/Schering Plough).

siNA molecules in combination with interferons and polyethylene glycol interferons have the potential to improve the effectiveness of treatment of SARS or any of the other indications discussed above. siNA molecules targeting RNAs associated with SARS virus infection can be used individually or in combination with other therapies such as interferons and polyethylene glycol interferons and to achieve enhanced efficacy.

Example 12: Diagnostic uses

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The siNA molecules of the invention can be used in a variety of diagnostic applications, such as in the identification of molecular targets (e.g., RNA) in a variety of applications, for example, in clinical, industrial, environmental, agricultural and/or research settings. Such diagnostic use of siNA molecules involves utilizing reconstituted RNAi systems, for example, using cellular lysates or partially purified cellular lysates. siNA molecules of this invention can be used as diagnostic tools to examine genetic drift and mutations within diseased cells or to detect the presence of endogenous or exogenous, for example viral, RNA in a cell. The close relationship between siNA activity and the structure of the target RNA allows the detection of mutations in any region of the molecule, which alters the base-pairing and three-dimensional structure of the target RNA. By using multiple siNA molecules described in this invention, one can map nucleotide changes, which are important to RNA structure and function in vitro, as well as in cells and tissues. Cleavage of target RNAs with siNA molecules can be used to inhibit gene expression and define the role of specified gene products in the progression of disease or infection. In this manner, other genetic targets can be defined as important mediators of the disease. These experiments will lead to better treatment of the disease progression by affording the possibility of combination therapies (e.g., multiple siNA molecules targeted to different genes, siNA molecules coupled with known small molecule inhibitors, or intermittent treatment with combinations siNA molecules and/or other chemical or biological molecules). Other in vitro uses of siNA molecules of this invention are well known in the art, and include detection of the presence of mRNAs associated with a disease, infection, or related condition. Such RNA is detected by determining the presence of a cleavage product after treatment with a siNA using standard methodologies, for example, fluorescence resonance emission transfer (FRET).

In a specific example, siNA molecules that cleave only wild-type or mutant forms of the target RNA are used for the assay. The first siNA molecules (*i.e.*, those that cleave only wild-type forms of target RNA) are used to identify wild-type RNA present in the sample and the second siNA molecules (*i.e.*, those that cleave only mutant forms of target RNA) are used to identify mutant RNA in the sample. As reaction controls,

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synthetic substrates of both wild-type and mutant RNA are cleaved by both siNA molecules to demonstrate the relative siNA efficiencies in the reactions and the absence of cleavage of the "non-targeted" RNA species. The cleavage products from the synthetic substrates also serve to generate size markers for the analysis of wild-type and mutant RNAs in the sample population. Thus, each analysis requires two siNA molecules, two substrates and one unknown sample, which is combined into six reactions. The presence of cleavage products is determined using an RNase protection assay so that full-length and cleavage fragments of each RNA can be analyzed in one lane of a polyacrylamide gel. It is not absolutely required to quantify the results to gain insight into the expression of mutant RNAs and putative risk of the desired phenotypic changes in target cells. The expression of mRNA whose protein product is implicated in the development of the phenotype (i.e., disease related or infection related) is adequate to establish risk. If probes of comparable specific activity are used for both transcripts, then a qualitative comparison of RNA levels is adequate and decreases the cost of the initial diagnosis. Higher mutant form to wild-type ratios are correlated with higher risk whether RNA levels are compared qualitatively or quantitatively.

All patents and publications mentioned in the specification are indicative of the levels of skill of those skilled in the art to which the invention pertains. All references cited in this disclosure are incorporated by reference to the same extent as if each reference had been incorporated by reference in its entirety individually.

One skilled in the art would readily appreciate that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as well as those inherent therein. The methods and compositions described herein as presently representative of preferred embodiments are exemplary and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art, which are encompassed within the spirit of the invention, are defined by the scope of the claims.

It will be readily apparent to one skilled in the art that varying substitutions and modifications can be made to the invention disclosed herein without departing from the scope and spirit of the invention. Thus, such additional embodiments are within the scope of the present invention and the following claims. The present invention teaches

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one skilled in the art to test various combinations and/or substitutions of chemical modifications described herein toward generating nucleic acid constructs with improved activity for mediating RNAi activity. Such improved activity can comprise improved stability, improved bioavailability, and/or improved activation of cellular responses mediating RNAi. Therefore, the specific embodiments described herein are not limiting and one skilled in the art can readily appreciate that specific combinations of the modifications described herein can be tested without undue experimentation toward identifying siNA molecules with improved RNAi activity.

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The invention illustratively described herein suitably can be practiced in the absence of any element or elements, limitation or limitations that are not specifically disclosed herein. Thus, for example, in each instance herein any of the terms "comprising", "consisting essentially of", and "consisting of" may be replaced with either of the other two terms. The terms and expressions which have been employed are used as terms of description and not of limitation, and there is no intention that in the use of such terms and expressions of excluding any equivalents of the features shown and described or portions thereof, but it is recognized that various modifications are possible within the scope of the invention claimed. Thus, it should be understood that although the present invention has been specifically disclosed by preferred embodiments, optional features, modification and variation of the concepts herein disclosed may be resorted to by those skilled in the art, and that such modifications and variations are considered to be within the scope of this invention as defined by the description and the appended claims.

In addition, where features or aspects of the invention are described in terms of Markush groups or other grouping of alternatives, those skilled in the art will recognize that the invention is also thereby described in terms of any individual member or subgroup of members of the Markush group or other group.

WO 2004/092383 PCT/US2004/011320

Table I: SARS virus Accession Numbers

5 LOCUS NC_004718 29736 bp ss-RNA linear VRL 15-APR-2003 DEFINITION SARS coronavirus, complete genome. ACCESSION NC_004718

Table II: SARS siNA and Target Sequences

SARS CoV NC_004718

Pos	Seq	Seq ID	UPos	Upper sea	Sea ID	LPos	Lower sea	Sea ID
3	ACCCAGGAAAAGCCAACCA	1	3	ACCCAGGAAAAGCCAACCA	1	21	neenneecnnnccneeen	1652
21	AACCUCGAUCUCUUGUAGA	2	21	AACCUCGAUCUCUUGUAGA	2	39	UCUACAAGAGAUCGAGGUU	1653
39	AUCUGUUCUCUAAACGAAC	3	39	AUCUGUUCUCUAAACGAAC	3	22	GUUCGUUUAGAGAACAGAU	1654
22	CUUUAAAAUCUGUGUAGCU	4	22	CUUUAAAAUCUGUGUAGCU	4	75	AGCUACACAGAUUUUAAAG	1655
75	NGUCGCUCGCCUGCAUGCC	5	75	nencecnceecnecynece	5	93	GGCAUGCAGCCGAGCACA	1656
93	CUAGUGCACCUACGCAGUA	9	93	CUAGUGCACCUACGCAGUA	9	111	UACUGCGUAGGUGCACUAG	1657
111	AUAAACAAUAAUAAAUUUU	7	111	AUAAACAAUAAUAAAUUUU	7	129	AAAAUUUAUUAUUGUUUAU	1658
129	UACUGUCGUUGACAAGAAA	8	129	UACUGUCGUUGACAAGAAA	8	147	UUUCUUGUCAACGACAGUA	1659
147	ACGAGUAACUCGUCCCUCU	6	147	ACGAGUAACUCGUCCCUCU	6	165	AGAGGACGAGUUACUCGU	1660
165	UNCUGCAGACUGCUUACGG	10	165	UNCUGCAGACUGCUUACGG	10	183	CCGUAAGCAGUCUGCAGAA	1661
183		11	183	GUUCGUCCGUGUUGCAGU	11	201	ACUGCAACACGGACGAAAC	1662
201	UCGAUCAUCAGCAUACCUA	12	201	UCGAUCAUCAGCAUACCUA	12	219	UAGGUAUGCUGAUGAUCGA	1663
219	AGGUUUCGUCCGGGUGUGA	13	219	AGGUUUCGUCCGGGUGUGA	13	237	UCACACCCGGACGAAACCU	1664
237	ACCGAAAGGUAAGAUGGAG	14	237	ACCGAAAGGUAAGAUGGAG	14	255	CUCCAUCUUACCUUUCGGU	1665
255	GAGCCUUGUUCUUGGUGUC	15	255	GAGCCUUGUUCGUGUC	15	273	GACACCAAGAACAAGGCUC	1666
273	CAACGAGAAAACACGUC	16	273	CAACGAGAAAACACACGUC	16	291	GACGUGUGUUUCUCGUUG	1667
291	CCAACUCAGUUUGCCUGUC	17	291	ccaacucaguuugccuguc	17	309	GACAGGCAAACUGAGUUGG	1668
309	CCUUCAGGUUAGAGACGUG	18	309	ccuucageunagagaceug	18	327	CACGUCUCUAACCUGAAGG	1669
327	GCUAGUGCGUGGCUUCGGG	19	327	ecuaeueceueecuuceee	19	345	CCCGAAGCCACGCACUAGC	1670
345	GGACUCUGUGGAAGAGGCC	20	345	GGACUCUGUGGAAGAGGCC	20	363	GGCCUCUUCCACAGAGUCC	1671
363	CCUAUCGGAGGCACGUGAA	21	363	CCUAUCGGAGGCACGUGAA	21	381	UUCACGUGCCUCCGAUAGG	1672
381	ACACCUCAAAAAUGGCACU	22	381	ACACCUCAAAAAUGGCACU	22	399	AGUGCCAUUUUUGAGGUGU	1673
333	UUGUGGUCUAGUAGAGCUG	23	399	UNGUGGUCUAGUAGAGCUG	23	417	CAGCUCUACUAGACCACAA	1674
417	GGAAAAGGCGUACUGCCC	24	417	GGAAAAGGCGUACUGCCC	24	435	GGGCAGUACGCCUUUUUCC	1675
435	CCAGCUUGAACAGCCCUAU	25	435	CCAGCUUGAACAGCCCUAU	25	453	AUAGGGCUGUUCAAGCUGG	1676
453	UGUGUUCAUUAAACGUUCU	56	453	UGUGUUCAUUAAACGUUCU	76	471	AGAACGUUUAAUGAACACA	1677
471	UGAUGCCUUAAGCACCAAU	27	471	UGAUGCCUUAAGCACCAAU	27	489	AUUGGUGCUUAAGGCAUCA	1678
489	UCACGGCCACAAGGUCGUU	28	489	UCACGGCCACAAGGUCGUU	28	202	AACGACCUUGUGGCCGUGA	1679
207	UGAGCUGGUUGCAGAAAUG	29	207	UGAGCUGGUUGCAGAAAUG	59	525	CAUUUCUGCAACCAGCUCA	1680
525	GGACGGCAUUCAGUACGGU	30	525	GGACGGCAUUCAGUACGGU	30	543	ACCGUACUGAAUGCCGUCC	1681
543	UCGUAGCGGUAUAACACUG	31	543	UCGUAGCGGUAUAACACUG	31	561	CAGUGUUAUACCGCUACGA	1682
261	GGGAGUACUCGUGCCACAU	32	561	GGGAGUACUCGUGCCACAU	32	579	AUGUGGCACGAGUACUCCC	1683
579	UGUGGGCGAAACCCCAAUU	33	579	UGUGGGCGAAACCCCAAUU	33	597	AAUUGGGGUUUCGCCCACA	1684
297	UGCAUACCGCAAUGUUCUU	34	597	UGCAUACCGCAAUGUUCUU	8	615	AAGAACAUUGCGGUAUGCA	1685
615	UCUUCGUAAGAACGGUAAU	35	615	UCUUCGUAAGAACGGUAAU	35	633	AUUACCGÜUCUUACGAAGA	1686
633	UAAGGGAGCCGGUGGUCAU	36	633	UAAGGGAGCCGGUGGUCAU	36	651	AUGACCACCGGCUCCCUUA	1687

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1688	1689	1690	1691	1692	1693	1694	1695	1696	1697	1698	1699	1700	1701	1702	1703	1704	1705	1706	1707	1708	1709	1710	1711	1712	1713	1714	1715	1716	1717	1718	1719	1720	1721	1722	1723	1724	1725	1726	1727	1728	1729
UAGAUCGAUGCCAUAGCUA	ACCUAAGUCAUAAGACUUU	AUCAGUGCCAAGCUCGUCA	UUCAUAAUCUUCAAUGGGA	CUUAGUUCCAGUUUGU	GAGUGCACCACUGCCAUGC	CUCACGAGUGAGUUCACGG	GACUGCACCUCCAUUGAGC	GUUGUCGACAUAGCGAGUG	AUCUGGGCCACAGAAAUUG	GCAAUCAAGAGGGUACCCA	UGCGAGAAAUCUUUGAUG	CAUUGACUUGCCCGCGCGU	UUGUUCGGAAAGAGUGCAC	CGACUCGAUGUAAUCAAGU	GCAGUAGACACCUCUCUUC	AUGCUCAUGGUCACGGCAG	AGUGAACCAGGCAAUUUCA	GCUCUUAUCAGAGCGCUCA	GGGUGUCUGGUGCUCGUAG	GGCACUCUUAAUUUCGAAG	GAAAGUGUCAAAUUUCUUG	CUUUGGGCAUUCCCCUUUG	GUUAAGAGGAAACACAAAC	AAUGACUUUGACUUUUGAG	CUUUUCAACACGUGGUUGA	GAAACCCUCAGUCUUUUC	AGAGCGUAUACGCCCCAUG	AGAUGCAACAGGGUACACA	AUUGUUACACUCCUGUGGA	CAAGGUAGACAAGUGCAUA	GCAAUGAUUACAUC	CUGCCAUGAAACUUCAUCG	UUUCAGAAAGUCGCACGUC	ACAAUGUUCACAAGUGGCU	AACUAAAUUUUCAGUGCCA	UGUAGUAGGUCCUUCAAUA	AGUAGGUAGGUACCCACAU	CAUUUUCACUACAGCAUUA	UUGACAGGCAGGACAUGGC	AGGUCCAAUCUCUGGGUCU	AUCUGCAACACUAUGCUCA
699	687	705	723	741	759	777	795	813	831	849	867	885	903	921	939	957	975	993	1011	1029	1047	1065	1083	1101	1119	1137	1155	1173	1191	5071	122/	1245	1263	1281	1299	1317	1335	1353	1371	1389	1407
37	88	39	4	41	42	43	44	45	46	47	48	49	22	2	52	23	72	22	26	22	28	29	99	61	23	8	2	65	3 8	۵	8 8	60	2	71	72	73	74	75	9/	77	78
UAGCUAUGGCAUCGAUCUA	AAAGUCUUAUGACUUAGGU	UGACGAGCUUGGCACUGAU	UCCCAUUGAAGAUUAUGAA	ACAAAACUGGAACACUAAG	GCAUGGCAGUGGUGCACUC	CCGUGAACUCACUCGUGAG	GCUCAAUGGAGGUGCAGUC	CACUCGCUAUGUCGACAAC	CAAUUUCUGUGGCCCAGAU	UGGGUACCCUCUUGAUUGC	CAUCAAAGAUUUUCUCGCA	ACGCGCGGCCAAGUCAAUG	GUGCACUCUUUCCGAACAA	ACUUGAUUACAUCGAGUCG	GAAGAGGUGUCUACUGC	CUGCCGUGACCAUGAGCAU	UGAAAUUGCCUGGUUCACU	UGAGCGCUCUGAUAAGAGC	CUACGAGCACCAGACACCC	CUUCGAAAUUAAGAGUGCC	CAAGAAAUUUGACACUUUC	CAAAGGGGAAUGCCCAAAG	GUUUGUGUUUCCUCUUAAC	CUCAAAAGUCAAAGUCAUU	UCAACCACGUGUUGAAAG	GAAAAGACUGAGGGUUUC	CAUGGGGCGUAUACGCUCU	UGUGUACCCUGUUGCAUCU	UCCACAGGAGUGUAACAAU	CALICA A LICITIA LICALITICS	GAUGAAAUGUAAUCAUUGC	CGAUGAAGUUUCAUGGCAG	GACGUGCGACUUUCUGAAA	AGCCACUUGUGAACAUUGU	UGGCACUGAAAAUUUAGUU	UAUUGAAGGACCUACUACA	AUGUGGGUACCUACO	UAAUGCUGUAGUGAAAAUG	GCCAUGUCCUGCCUGUCAA	AGACCCAGAGAUUGGACCU	UGAGCAUAGUGUUGCAGAU
651	699	/89	705	723	741	759	777	795	813	831	849	867	882	903	921	939	957	975	993	101	1029	1047	1065	1083	1101	1119	113/	133	11/3	1200	1227	1276	1243	2021	1281	1299	1317	1335	1353	1371	1389
37	88	£ 5	40	41	42	43	4	45	46	_														وا	20 25	3 3	4 2	8 8	00	a a	3 8	60 6	2 6	- F	7)	5 5	74	75	9/		8
UAGCUAUGGCAUCGAUCUA	116ACGACCIIICCCACIICALI		UCCCAUGGAGAUDAUGAA	ACAAACUGGAACACUAAG	GCAUGECAGUGGACUC	CCGUGAACUCACUCGUGAG	GCUCAAUGGAGGUGCAGUC	CACUCGCUAUGUCGACAAC	CAAUUUCUGUGGCCCAGAU	UGGGUACCCUCUUGAUUGC	CAUCAAAGAUUUUCUCGCA	ACCCGGGCAAGUCAAUG	GUGCACCAACAA	Acudeaudacadceaeuce	GIOCOLIO	COGCCGOGACCAOGAGCAO	UGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	GUNGAGER	CUACGAGCACCAGACACCC	CUUCGAAAUUAAGAGUGCC	CAAGAAUUUGACACUUUC	CAAAGGGGAAUGCCCAAAG	GUGAAAGUGAAGUGAAC	COCAGAGOCAGO	GAAAAAAAAAAG	CALICOCOCITALIACOCITALIA	LIGHT INCIDENTIAL STREET	UCCACAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	UALIGCACIII IGI ICI IACCIII IG	GALIGAAALIGITAALICALIIIGC	CGALIGAAGIIIIICALIGGGAG	GACGUGCGACIIIIICIGAAA	AGCCACILICIAACAIIIIOII	HOCCACIOGAGAACAOGGO	US III O TO TO THE OTHER OF THE OTHER OTHE	VAUGEACCUACUACA	AUGUGGGUACCUACU	UAAUGCUGUAGUGAAAUG	GCCAUGUCCUGCCUGUCAA	AGACCCAGAGAUUGGACCU	ucaccada e ucaccada u
651	682	3 5	202	57,	4 6	60,		GF 5	813	. S	843	200	600	303	35	-	22		- 1		- 1		200	1100	5 2	1137	115	1173	1191	1209	1227	1245	1963	1262	1200	1217	1337	1333	1333	1380	1503

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1730	1731	1732	1733	1734	1735	1736	1737	1738	1739	1740	1741	1742	1743	1744	1745	1746	1747	1748	1749	1750	1751	1752	1753	1754	1755	1756	1757	1758	1759	1760	1761	1/62	1763	1764	1765	1766	1767	1768	1769	1770	1771
GUUUGAGUGGUUGUGAUAA	GCGGAGUCGAGUUCAAUG	UCUAGUCCUACCUCCCUUG	CACACAGCCUCCAAAACAU	GCAGCCAACAUAGGCAAAC	GUAGGCACGCUUAUUAUAG	ACUAGCACGAGGAACCCAG	GCCUGAGCCAAUAUCAGCA	ACCAGUAAUGCCAGUAUGG	CAAGGUCUCCACAUUGUCA	CUCAAGGAGAUCCUCAUUC	ACGUUCACGACUCAGUAUC	AACAAUGUUAAUGUUAACA	AUUCAAAUGAAAAUCGCCA	AAUGAUGGCAACCUCUUCA	AGCAGAGAAGAUGCCAAA	AAUAAAGGCACUUGUAGAA	AAGACUCUUUAUAGUGUCA	UUUGAAAGACUUGUAAUCA	GCAGGACUCAACAAUGGUU	GGUAACUUUAUAGUUACCG	UNUNACGGGCUUNCCCUUG	UCCAAUGUUCCAAGCACCU	UAAAACUGAUCUCUGUUGU	AAAACCACACAGUGGUGUU	ACCAGCAGCCUGUGAGGGA	AAAAAUUGAUCUGAUAACA	UGCAUCAAGUGUGCGCGCA	AGGAAUUGAGUGGUUUGCU	AGCUGCUCUUUGCAAAUCA	ACCAUCAAGUAUGGUGACA	UAAUGACUGUUCAGAAUA	CAUGGCGUCGACAGGCGU	CAGGUCUGAAGUANAAACC	AAUGACACUGUUGGUGAGC	AGUUACAUAUGCCAUAAUA	CUGUUGUACAAGACCACCA	AGACAACCACUGAGAAGUC	AGUAGUGCCCAAAAGAUUA	AGGCCUGAGUUUUUCAACA	CUCAAUCCAUUCAAAGAUA	UCCUGCACUAAGUUUCGCC
1425	1443	1461	1479	1497	1515	1533	1551	1569	1587	1605	1623	1641	1659	1677	1695	1713	1731	1749	1767	1785	1803	1821	1839	1857	1875	1893	1911	1929	1947	1965	1983	500	2019	2037	2025	2073	2091	2109	2127	2145	2163
79	8	84	82	83	8	82	98	87	88	68	6	91	95	93	8	95	96	97	86	66	100	101	102	103	104	105	106	107	89	60	2 ;	- 6	211	113	114	115	116	117	118	119	120
UNAUCACAACCACUCAAAC	CAUUGAAACUCGACUCCGC	CAAGGGAGGUAGGACUAGA	AUGUUUGGAGGCUGUGUG	GUUUGCCUAUGUUGGCUGC	CUAUAAUAAGCGUGCCUAC	CUGGGUUCCUCGUGCUAGU	UGCUGAUAUUGGCUCAGGC	CCAUACUGGCAUUACUGGU	UGACAAUGUGGAGACCUUG	GAAUGAGGAUCUCCUUGAG	GAUACUGAGUCGUGAACGU	UGUUAACAUUAACAUUGUU	UGGCGAUUUCAUUUGAAU	UGAAGAGGUUGCCAUCAUU	UNGGCANCUUCUCUCCO	UUCUACAAGUGCCUUUAUU	UGACACUAUAAAGAGUCUU	UGAUUACAAGUCUUUCAAA	AACCAUUGUUGAGUCCUGC	CGGUAACUAUAAAGUUACC	CAAGGGAAAGCCCGUAAAA	AGGUGCUUGGAACAUUGGA	ACAACAGAGAUCAGUUUUA	AACACCACUGUGGUUUU	UCCCUCACAGGCUGCUGGU	UGUUAUCAGAUCAAUUUUU	UGCGCGCACACUUGAUGCA	AGCACACCACUCAAUUCCU	UGAUUUGCAAAGAGCAGCU	IIAIIII ICIICAAAAAAAAAAA	ACCIONICACCOCATION	SOLO O O O O O O O O O O O O O O O O O O	COLICACIONACIONI ICALIENTE	GCUCACCAACAGUGUCAUU	UAUUAUGGCAUAUGUAACU	OGGUGGUCUUGUACAGG	GACUUCUCAGUGGUUGUCU	UAAUCUUUUGGGCACUACU	UGUUGAAAACUCAGGCCU	UAUCUUUGAAUGGAUUGAG	GGCGAAACUUAGUGCAGGA
1407	1425	1443	1461	1479	1497	1515	1533	1551	1569	1587	1605	1623	1641	1659	/91	1695	1/13	1731	1749	1767	1785	1803	1821	1839	1857	18/5	1893	1 2 2	1929	1065	1083	2004	2040	2000	2000	2002	5073	2091	2109	2127	2145
62	2 2	-	+	83	20 2	82	98	87	88	89	8	9	92	23	25 2	3	<u>8</u>	9/	86	66	100	101	102	20 5	404	3	9 2	101	9 2	13	111	113	113	5 4	140	2 6	2	-1	118	119	120
UNAUCACAACCACUCAAAC	CANGGGAAACUCGACUCCGC	ALICITIONACCORCIONO	AugundegAggCugugu	GUNGECCUANGUNGGCUGC	CURORAUAGCEUECCUAC	COGGGUUCCUCGUGCUAGU	UGCUGAUAUUGGCUCAGGC	CCAUACUGGCAUUACUGGU	UGACAAUGUGGAGACCUUG	GAAUGAGGAUCUCCUUGAG	GAUACUGAGUCGUGAACGU	UGUUAACAUUGUU	UCAACAGUUCAUUGAAU	Verace GOOGCCAUCAU	COORDINATION OF THE PROPERTY O	UCACAGO CCOUDADO	UGALACUAUAAAGAGUCUU	VSAUUACAAGUCUUUCAAA	AACCAUGEUUGAGUCCUGC	CGGUAACUAUAAAGUUACC	CAAGGGAAAGCCCGUAAAA	AGGUGCUUGGAACAUUGGA	ACAACAGAGACAGAAAAAAAAAAAAAAAAAAAAAAAAAA	MCACCACOGOGOGOGOOOO	I I I I I I I I I I I I I I I I I I I	16CGCGCACACACACCOOO	AGGAAACCACIICAAIIIICCII	I IGALIII IGCAAAGAGCAGCI I	UGUCACCAIJACIIIGALIGGII	UAUUUCUGAACAGUCATITA	ACGUCUUGUCGACGCCALIG	GGUUUAUACIIIICAGACCIIG	GCICACCAACAGIGIICAIIII	UALITATION	HGGHGGHCIIIIGHACACAC	GACHICIDAGIDGITIGITI	110411040000000000000000000000000000000	UNAUCUUUUGGGCACUACU	UGUUGAAAACUCAGGCCU	GGCGAACIIIACIICAGGA	GGCGAACOOAGOGCAGGA
1407	1443	146.	1470	14/3	1637	010	255	1351	1569	1587	200	1644	1850	1677	1808	1712	1734	1240	1767	1707	1,03	1003	1830	1857	1875	1803	1911	1929	1947	1965	1983	2001	2019	2037	2055	2073	2004	2400	2103	2145	01.7

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1814	1815	1816	1817	1818	1819	1820	1821	1822	1823	1824	1825	1826	1827	1828	1829	1830	1831	1832	1833	1834	1835	1836	1837	1838	1839	1840	1841	1842	1843	1844	1845	1846	1847	1848	1849	1850	1851	1852	1853	1854	1855
AAUACCCAUGUUGGUAAGG	ACUCCACUCAUCAAGAUCA	UAAGUAGAAUGUAGCUACA	UUCACCAGCAUCAUCAAAU	ACGUGAUGAAAAGUUUUCU	GUAAAAGGAACAAUACAUA	UUCUUCCUCAUCUGGAGGG	ACACUCUGCAUCGUCCUCU	AUCAAUUUCUUCCUCA	CUCAUGUUCACAGGUUUCA	AUCAUCCUCUGUACCGUAC	CAGAGGGAGCCUUGAUAA	AGCUGAGGCACCAAAUUCC	CUCAACUCGAACUGUUUCA	GUCUNCCUCUNCUNCC	AGUAGUAUCAUCCAGCCAG	CUCAAUCUCUGAUUGCUCA	UGUAGGUUCUGGU	AUUAACUGGUUCUUCAGGU	UAAAUAACCAGUAAACUGA	AACAUUGUCAGUAAGUUUU	GUCAACACAUUNAAUGGCA	UUGUGCCUCCUUAACGAUG	CACCAUAGGAUUAGCACUU	GUUAGCAGCAUUUACAAUC	ACCAUGUUUCAGGUGUAUG	UGCACCUGCUACACCACCA	AUUGGUUGCCUUGUUGAGU	CUCCUUUUGCAUGGCACCA	CUUAAUGUAAUCAUCACUC	UGUAAGAGGGCCAUUUAGC	CAAACAAGACCCUCCUACU	AAGAUUAUGUCCAGAAAGC	AUGCAGACACUUCUUAGCA	UAGGUUAGGUCCAACAACA	GAUGUCCUCACCUGCAUUU	UGCUGCCUUAAGAAGCUGG	UGAAUUGAAAUUUUCAUAU	UGCAAGUAAGAUGUCCUGU	GCCUGCUGACAACAAUGGU	UGGUUUAGCACCAAAUAUG	CACUUGUAAAGACUGAAGU
2937	2955	2973	2991	3009	3027	3045	3063	3081	3099	3117	3135	3153	3171	3189	3207	3225	3243	3261	3279	3297	3315	3333	3351	3369	3387	3405	3423	3441	3459	3477	3495	3513	3531	3549	3567	3585	3603	3621	3639	3657	3675
163	164	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	181	182	183	184	185	186	187	188	189	190	191	192	193	194	195	196	197	198	199	200	201	202	203	204
CCUUACCAACAUGGGUAUU	UGAUCUUGAUGAGUGGAGU	UGUAGCUACAUUCUACUUA	AUUUGAUGAUGCUGGUGAA	AGAAAACUUUUCAUCACGU	UAUGUAUUGUUCCUUUUAC	CCCUCCAGAUGAGGAAGAA	AGAGGACGAUGCAGAGUGU	UGAGGAAGAAGAAUUGAU	UGAAACCUGUGAACAUGAG	GUACGGUACAGAGGAUGAU	UNAUCAAGGUCUCCCUCUG	GGAAUUUGGUGCCUCAGCU	UGAAACAGUUCGAGUUGAG	GGAAGAAGAGGGAAGAC	CUGGCUGGAUGAUACUACU	UGAGCAAUCAGAGAUUGAG	GCCAGAACCAGAACCUACA	ACCUGAAGAACCAGUUAAU	UCAGUUUACUGGUUAUUUA	AAAACUUACUGACAAUGUU	UGCCAUUAAAUGUGUUGAC	CAUCGUUAAGGAGGCACAA	AAGUGCUAAUCCUAUGGUG	GAUUGUAAAUGCUGCUAAC	CAUACACCUGAAACAUGGU	UGGUGGUGUAGCAGGUGCA	ACUCAACAAGCAACCAAU	UGGUGCCAUGCAAAAGGAG	GAGUGAUGAUUACAUUAAG	GCUAAAUGGCCCUCUUACA	AGUAGGAGGGUCUUGUUUG	GCUUUCUGGACAUAAUCUU	UGCUAAGAAGUGUCUGCAU	UGUUGUUGGACCUAACCUA	AAAUGCAGGUGAGGACAUC	CCAGCUUCUUAAGGCAGCA	AUAUGAAAUUUCAAUUCA	ACAGGACAUCUUACUUGCA	ACCAUUGUUGUCAGCAGGC	CAUAUUUGGUGCUAAACCA	ACUUCAGUCUUUACAAGUG
2919	2937	2925	2973	2991	3009	3027	3045	3063	3081	3099	3117	3135	3153	3171	3189	3207	3225	3243	3261	3279	3297	3315	3333	3351	3369	3387	3405	3423	3441	3459	3477	3495	3513	3531	3549	3567	3585	3603	3621	3639	3657
163	<u>4</u>	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	181	_		184		186		188	189	190	191	192	193	194	195	196	197	198	199	200	201	202	203	204
CCUUACCAACAUGGGUAUU	UGAUCTUGAUGAGUGGAGU	UGUAGCUACAUUCUACUUA	AUUUGAUGAUGCUGGUGAA	AGAAAACUUUUCAUCACGU	UAUGUAUUGUUCCUUUUAC	CCCUCCAGAUGAGGAAGAA	AGAGGACGAUGCAGAGUGU	UGAGGAAGAAGAAAUUGAU	UGAAACCUGUGAACAUGAG	GUACGGUACAGAGGAUGAU	UNAUCAAGGUCUCCCUCUG	GGAAUUUGGUGCCUCAGCU	UGAAACAGUUCGAGUUGAG	GGAAGAAGAAGAGACA	CUGGCUGGAUGAUACUACU	UGAGCAAUCAGAGAUUGAG	GCCAGAACCAGAACCUACA	ACCUGAAGAACCAGUUAAU	UCAGUUUACUGGUUAUUUA	AAAACUUACUGACAAUGUU	UGCCAUUAAAUGUGUUGAC	CAUCGUUAAGGAGGCACAA	AAGUGCUAAUCCUAUGGUG	GAUUGUAAAUGCUGCUAAC	CAUACACCUGAAACAUGGU	UGGUGGUAGCAGGUGCA	ACUCAACAAGGCAACCAAU	UGGUGCCAUGCAAAAGGAG	GAGUGAUGAUACAUUAAG	SCUAAAUGGCCCUCUUACA	AGUAGGAGGGUCUUGUUG	SCOUNCUGGACAUAAUCUU	UGCUAAGAGGGGCGU	UGUUGGACCUAACCUA	AAAUGCAGGUGAGGACAUC	CCAGCUUCUUAAGGCAGCA	AUAUGAAAUUUCAAUUCA	ACAGGACAUCUUACUUGCA	ACCAUGUGUCAGCAGGC	CAUAUUGGUGCUAAACCA	ACUUCAGUCUUUACAAGUG
2919	2055	2200	5/67	2991	3003	3027	3045	3063	3081	3099	3117	3135	3153	27/12	3189	3207	3225	3243		3279			- 1		3369	3387	24.53	2473	2450	2472	3405	25430	2013	200	3249	/000	3282	3003	1700	3657	1000

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1856	1858	1859	1860	1861	1862	1863	1864	1865	1866	1867	1868	1869	1870	1871	1872	1873	1874	1875	1876	1877	1878	1879	1880	1881	1882	1883	1884	1885	1886	1887	1888	1889	1890	1891	1892	1893	1894	1895	1896	1897
ACGAACCGUCUGCACCAC	AAGAGCIIIIIGACII	CAUGACAACCUGCUCAUAA	CAGGUUAUCAAGAUAAUCC	UGCUUCCACUCUAGGCUUC	UGGCUCCUCUUGUUAGGU	GGAAUCUUCUGUGUUUGGU	AGAUUUCUCCUCAGUUUUG	GACAGGCUUCUGUACGACA	AAUUUUGGCUUCACAUCG	CUCAUCAAUGCAGGCCUUA	UUCCAGUGUUGUGGUAACC	GGUAAGAAACUUAGUUUCU	AAACAAGAGUAACUUAUUG	CUUACCAUUGAUAUCAGCA	CUGAGAAUCAUGGUAAAGC	UUCACCUCUAAGCAUGUUC	CUCAAGGAAAGACAUAUCU	CAUGUAAGGUGCAUCCUUC	AGUGAUAACAUCACCUACC	ACAAGUGAUAUCACCACUA	UUUGGAGGGUAUUACAACA	AGUAGUGCCACCAGCCUUU	AGCUCUUGAGAGCAUCUCA	AACUGGCACUUUCUUCAAA	CGUGGUUAUAUACUCAUCA	ACAUCCUUGUCCAGGGUAC	CUCAAGUGUAUAACCAGCA	AAGAGCAGUCUUAGCUUCC	UGCAGAUUUGCAUUUCUUA	UGAAGGUAGUACAUAAAAU	CUUAGCAUUAGGUGCUUCU	AGUUCCUAGAAUCUCUUCC	UCUCAAAUUCCAGGAUACA	AGCAUGAGCAAGCAUUUCU	UAAUUUUCUUGUCUCUUCA	AUCCAUGCAUAUAGGCAUU	UGCCAUUAUGGCUCUAACA	AUACUUACGUUGGAUGGUU	UUGAAUUUUAAUUCCUUUA	AUAGUCAACGAUGCCCUCU
3693	3729	3747	3765	3783	3801	3819	3837	3855	3873	3891	3909	3927	3945	3963	3981	3999	4017	4035	4053	4071	4089	4107	4125	4143	4161	4179	4197	4215	4233	4251	4269	4287	4305	4323	4341	4359	4377	4395	4413	4431
205	202	208	509	210	211	212	213	214	215	216	217	218	219	220	221	222	223	224	225	226	227	228	529	230	231	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246
GUGCGUGCAGACGGUUCGU	AGLICAALIGACAAAGCLICIIII	UNAUGAGCAGGUUGUCAUG	GGAUUAUCUUGAUAACCUG	GAAGCCUAGAGUGGAAGCA	ACCUAAACAAGAGGAGCCA	ACCAAACACAGAAGAUUCC	CAAAACUGAGGAGAAAUCU	UGUCGUACAGAAGCCUGUC	CGAUGUGAAGCCAAAAAUU	UAAGGCCUGCAUUGAUGAG	GGUUACCACACACUGGAA	AGAAACUAAGUUUCUUACC	CAAUAAGUUACUCUUGUUU	UGCUGAUAUCAAUGGUAAG	GCUUUACCAUGAUUCUCAG	GAACAUGCUUAGAGGUGAA	AGAUAUGUCUUUCCUUGAG	GAAGGAUGCACCUUACAUG	GGUAGGUGAUGUUAUCACU	UAGUGGUGAUAUCACUUGU	UGUUGUAAUACCCUCCAAA	AAAGGCUGGUGGCACUACU	UGAGAUGCUCUCAAGAGCU	UUUGAAGAAAGUGCCAGUU	UGAUGAGUAUAUAACCACG	GUACCCUGGACAAGGAUGU	UGCUGGUUAUACACUUGAG	GGAAGCUAAGACUGCUCUU	UAAGAAAUGCAAAUCUGCA	AUUUNAUGUACUACCUUCA	AGAAGCACCUAAUGCUAAG	GGAAGAUUCUAGGAACU	UGUAUCCUGGAAUUUGAGA	AGAAAUGCUUGCUCAUGCU	UGAAGAGACAAGAAAUUA	AAUGCCUAUAUGCAUGGAU	UGUUAGAGCCAUAAUGGCA	AACCAUCCAACGUAAGUAU	UAAAGGAAUUAAAAUUCAA	AGAGGGCAUCGUUGACUAU
3675	3711	3729	3747	3765	3783	3801	3819	3837	3855	3873	3891	3909	3927	3945	3963	3981	3999	4017	4035	4053	4071	4089	4107	4125	4143	4161	4179	4197	4215	4233	4251	4269	4287	4305	4323	4341	4359	4377	4395	4413
205	202	208	509	210	211	212	213	214	215	216	217	218	219	220	221	222	223	224	225	226	227	228	229	230	231	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246
GUGCGUGCAGACGGUUCGU	AGUCAAUGACAAAGCIICIIII	UNAUGAGCAGGUUGUCAUG	GGAUUAUCUUGAUAACCUG	GAAGCCUAGAGUGGAAGCA	ACCUAAACAAGAGGAGCCA	ACCAAACACAGAAGAUUCC	CAAAACUGAGGAGAAAUCU	UGUCGUACAGAAGCCUGUC	CGAUGUGAAGCCAAAAAUU	UAAGGCCUGCAUUGAUGAG	GGUUACCACACACUGGAA	AGAAACUAAGUUUCUUACC	CAAUAAGUUACUCUUGUUU	UGCUGAUAUCAAUGGUAAG	GCUUUACCAUGAUUCUCAG	GAACAUGCUUAGAGGUGAA	AGAUAUGUCUUUCCUUGAG	GAAGGAUGCACCUUACAUG	GGUAGGUGAUGUUAUCACU	UAGUGGUGAUAUCACUUGU	UGUUGUAAUACCCUCCAAA	AAAGGCUGGUGGCACUACU	UGAGAUGCUCUCAAGAGCU	UUUGAAGAAGUGCCAGUU	UGAUGAGUAUAUAACCACG	GUACCCUGGACAAGGAUGU	UGCUGGUUAUACACUUGAG	GGAAGCUAAGACUGCUCUU	UAAGAAAUGCAAAUCUGCA	AUUUUAUGUACUACCUUCA	AGAAGCACCUAAUGCUAAG	GGAAGAGUUCUAGGAACU	UGUAUCCUGGAAUUUGAGA		UGAAGAGACAAGAAAUUA	AAUGCCUAUAUGCAUGGAU	UGUUAGAGCCAUAAUGGCA	AACCAUCCAACGUAAGUAU	UAAAGGAAUUAAAAUUCAA	AGAGGCAUCGUUGACUAU
3693	3711	3729	3747	3765	3783	3801	3819	3837	3855	3873	3891	3909	3927	3945	3963	3981	3999	4017	4035	4053	4071	4089	4107	4125	4143	4161	4179	4197	4215	4233	4251	4269	4287	4305	4323	4341	4359	4377	4395	4413

4431
UDADACUAGUAAAGAGCCO
GAAGCUGAACUCUCUAAAU
UGAGCCGCUUGUCACAAUG
GCCAAUUGGUUAUGUGACA
ACAUGGUUUUAAUCUUGAA
AGAGGCUGCGCGCUGUAUG
GCGUCCUCAAAGCUCCU
VGCCGUAGUGUCAGUAUCA
UACAUAUAAUGGAUACCUC
CACUUCGUCAUCAAAGACA
AUCUGAGGAGCACUUUGUA
AGAAACAGUUUCUUUGGCL
UGGCUCUUACAGAGAUUGG
GUCCUAUUCAGGACAGCGL
UACAGAGUUAGGUGUUGAA
AUUUCUUAAGCGUGGUGAC
CAAAAUUGUGUACCACACL
UCUGGAGAGCCCCGUCGAG
GUUUCAUCUUGACGGUGAG
GGUUCUUUCACUUGACAAA
ACUAAAGAGUCUCUUAUCC
CCUGCGGGAGGUUAAGACL
UAUAAAAGUGUUCACAACU
UGUGGACAACACUAAUCUC
CCACACAGCUUGUGGAL
<u>UAUGUCUAUGACAUAUGGA</u>
ACAGCAGUUUGGUCCAACA
AUACUUGGAUGGUGCUGAI
UGUUACAAAAAUUAAACCU
UCAUGUAAAUCAUGAGGGU
UAAGACUUUCUUUGUACUA
ACCUAGUGAUGACACACUA
ACGUAGUGAAGCUUUCGAG
GUACUACCAUACUCUUGAU
UGAGAGUUUUCUUGGUAGG
GUACAUGUCUGCUUUAAAC
CCACACAAAGAAAUGGAAA
AUUUCCUCAAGUUGGUGGL
UUUAACUUCAAUUAAAUGG

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1940	1941	1942	1943	1944	1945	1946	1947	1948	1949	1950	1921	1952	1953	1954	1955	1956	1957	1958	1959	1960	1961	1962	1963	1964	1965	1966	1967	1968	1969	1970	1971	1972	1973	1974	1975	1976	1977	1978	1979	1980
AUAACAAUUGUIAUCAGCC	UAAUAAACACUAGACAAA	UUCAAGCUGUUGAAGUGCU	UGGUGCAUUGAAUUUGACU	AUAAGCCUCUUGAAGUGCU	ACCAGCACGGCCUCUAUAA	ACAAAAGUUAGCAGCAUCA	GUAAGCGAGUAUGAGUGCA	GCCAACAGUUUUAUUACUG	UCUGACAUCACCAAGCUCG	AAGAUGGGUCAUAGUUUCU	CAAAUUAGCAUGCUGUAGA	AACUCGCUUUGCAGAUUCC	UUUACACACCACAUUAAGA	AGUUUUCUGACCACAAUGU	UACACCCGUUAAGGUAGUA	CAUAUACAUCACAGCUUCU	AUCAUAAGAUAGAGUACCC	AACACCUGUCUUAAGAUUA	ACACACAUGGAAUGGAA	UUGUGUAGCAUCACGACCA	CUCUUGUUGUACUAGAUAU	CAUCAUAACAAAGAAGAC	CUCAGCAGGUGCAGAC	ACCUUGCUGUAAUUUAUAC	AUUCGCACAUAAGAAUGUA	AUAGUUACCAGUGUACUCA	AGUGUAAUGACCACACUGA	CUCCUUAGCAGUUAUAUGA	GUCAAUACGAUAGAGGGUC	CUUUGUAAGGUGAGCUCCG	UCCUUUGUACUCUGACAUC	GAAAACAUCAGUCACUGGU	GUAAGAUGUUCCUUGUAG	AGGCUUGAUGGUUGUAGUG	AUCGAGUUUAUACGACACA	CUCUGUGUAAGUAACUCCA	AUCCAAUUUGGUUCAAUC	AUCCUUUUUAUAAUACCCA	CUCUGUAUAGUAAGCAUUA	UACAAGGUCUAUAGGCUGC
5205	5223	5241	5259	5277	5295	5313	5331	5349	5367	5385	5403	5421	5439	5457	5475	5493	5511	5529	5547	5565	5583	5601	5619	5637	5655	5673	5691	5709	5727	5745	5763	5781	5799	5817	5835	5853	5871	2889	2907	5925
289	290	291	292	293	294	295	296	297	298	299	300	301	302	303	304	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329
GGCUGAUAACAAUUGUUAU	UUUGUCUAGUGUUUUAUUA	AGCACUUCAACAGCUUGAA	AGUCAAAUUCAAUGCACCA	AGCACUUCAAGAGGCUUAU	UNAUAGAGCCCGUGCUGGU	UGAUGCUGCUAACUUUGU	UGCACUCAUACUCGCUUAC	CAGUAAUAAAACUGUUGGC	CGAGCUUGGUGAUGUCAGA	AGAAACUAUGACCCAUCUU	UCUACAGCAUGCUAAUUUG	GGAAUCUGCAAAGCGAGUU	UCUUAAUGUGGUGUGUAAA	ACAUUGUGGUCAGAAACU	UACUACCUUAACGGGUGUA	AGAAGCUGUGAUGUAUAUG	GGGUACUCUAUCUUAUGAU	UAAUCUUAAGACAGGUGUU	unccannccanenenenen	UGGUCGUGAUGCUACACAA	AUAUCUAGUACAACAAGAG	GUCUUCUUUGUUAUGAUG	GUCUGCACCACCUGCUGAG	GUAUAAAUUACAGCAAGGU	UACAUUCUUAUGUGCGAAU	UGAGUACACUGGUAACUAU	UCAGUGUGGUCAUUACACU	UCAUAUAACUGCUAAGGAG	GACCCUCUAUCGUAUUGAC	CGGAGCUCACCUUACAAAG	GAUGUCAGAGUACAAAGGA	ACCAGUGACUGAUGUUUC	CUACAAGGAAACAUCUUAC	CACUACAACCAUCAAGCCU	UGUGUCGUANAAACUCGAU	UGGAGUUACUUACACAGAG	GAUUGAACCAAAAUUGGAU	UGGGUAUUAUAAAAAGGAU	UAAUGCUUACUAUACAGAG	GCAGCCUAUAGACCUUGUA
5187	5205	5223	5241	5259	5277	5295	5313	5331	5349	5367	5385	5403	5421	5439	5457	5475	5493	5511	5529	5547	2265	5583	5601	5619	5637	5655	5673	5691	5709	5727	5745	5763	5781	5799	5817	5835	5853	5871	5889	2907
289	290	291	292	293	294	295	596	297	298	299	300	301	302	303	304	305	306	307	808	608	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329
GGCUGAUAACAAUUGUUAU	UUUGUCUAGUGUUUAUUA	AGCACUUCAACAGCUUGAA	AGUCAAAUUCAAUGCACCA	AGCACUUCAAGAGGCUUAU	UNAUAGAGCCCGUGCUGGU	UGAUGCUGCUAACUUUGU	UGCACUCAUACUCGCUUAC	CAGUAAUAAAACUGUUGGC	CGAGCUUGGUGAUGUCAGA	AGAAACUAUGACCCAUCUU	UCUACAGCAUGCUAAUUUG	GGAAUCUGCAAAGCGAGUU	UCUUAAUGUGGUGUGUAAA	ACAUUGUGGUCAGAAAACU	UACUACCUUAACGGGUGUA	AGAAGCUGUGAUGUAUAUG	GGGUACUCUAUCUUAUGAU	UAAUCUUAAGACAGGUGUU	UUCCAUUCCAUGUGUGUGU	UGGUCGUGAUGCUACACAA	AUAUCUAGUACAACAAGAG	GUCUUCUUUGUUAUGAUG	GUCUGCACCACCUGCUGAG	GUAUAAAUUACAGCAAGGU	UACAUUCUUAUGUGCGAAU	UGAGUACACUGGUAACUAU	UCAGUGUGGUCAUUACACU	UCAUAUAACUGCUAAGGAG	GACCCUCUAUCGUAUUGAC	CGGAGCUCACCUUACAAAG	GAUGUCAGAGUACAAAGGA	ACCAGUGACUGAUGUUUC	CUACAAGGAAACAUCUUAC	CACUACAACCAUCAAGCCU	UGUGUCGUAUAAACUCGAU	UGGAGUUACUUACACAGAG	GAUUGAACCAAAAUUGGAU	UGGGUAUUAUAAAAAGGAU		GCAGCCUAUAGACCUUGUA
5187	5205	5223	5241	5259	5277	5295	5313	5331	5349	5367	5385	5403	5421	5439	5457	5475	5493	5511	5529	5547	5565	5583	5601	5619	5637	5655	5673	5691	5709	5727	5745	5763	5781	5799	5817	5835	5853	5871	5889	2907

UAACACAAAAUUUGCUGAU		2		332	5979	AGAACAUGUGAGUUUGAAA	1983
	333	5979	UAACACAAAAUUUGCUGAU	333	2997	AUCAGCAAAUUUUGUGUUA	1984
1	334	2997	UGAUUUAAAUCAAAUGACA	334	6015	UGUCAUUUGAUUUAAAUCA	1985
\forall	335	6015	AGGCUUCACAAAGCCAGCU	335	6033	AGCUGGCUUUGUGAAGCCU	1986
_	336	6033	UUCACGAGAGCUAUCUGUC	336	6051	GACAGAUAGCUCUCGUGAA	1987
-	337	6051	CACAUUCUUCCCAGACUUG	337	6909	CAAGUCUGGGAAGAAUGUG	1988
nossi	338	6909	GAAUGGCGAUGUAGUGGCU	338	6087	AGCCACUACAUCGCCAUUC	1989
CUAU	339	6087	UAUUGACUAUAGACACUAU	339	6105	AUAGUGUCUAUAGUCAAUA	1990
GAAA	340	6105	UUCAGCGAGUUUCAAGAAA	340	6123	UNUCUUGAAACUCGCUGAA	1991
1	341	6123	AGGUGCUAAAUUACUGCAU	341	6141	AUGCAGUAAUUUAGCACCU	1992
	342	6141	UAAGCCAAUUGUUUGGCAC	342	6159	GUGCCAAACAAUUGGCUUA	1993
CAUUAACCAGGCUACAACC 3	343	6159	CAUUAACCAGGCUACAACC	343	6177	GGUUGUAGCCUGGUUAAUG	1994
	344	6177	CAAGACAACGUUCAAACCA	344	6195	UGGUUUGAACGUUGUCUUG	1995
\dashv	345	6195	AAACACUUGGUGUUUACGU	345	6213	ACGUAAACACCAAGUGUUU	1996
	346	6213	UUGUCUUUGGAGUACAAAG	346	6231	CUUUGUACUCCAAAGACAA	1997
AAAU	347	6231	GCCAGUAGAUACUUCAAAU	347	6249	AUUUGAAGUAUCUACUGGC	1998
_	348	6249	UUCAUUUGAAGUUCUGGCA	348	6267	UGCCAGAACUUCAAAUGAA	1999
\dashv	349	6267	AGUAGAAGACACACAAGGA	349	6285	nccnnenenencnncnvcn	2000
	350	6285	AAUGGACAAUCUUGCUUGU	320	6303	ACAAGCAAGAUUGUCCAUU	2001
CACC	351	6303	UGAAAGUCAACAACCCACC	351	6321	GGUGGGUUGUGACUUCA	2002
	352	6321	CUCUGAAGAAGUAGUGGAA	352	6339	UUCCACUACUUCUUCAGAG	2003
SAAG	353	6339	AAAUCCUACCAUACAGAAG	353	6357	CUUCUGUAUGGUAGGAUUU	2004
	354	6357	GGAAGUCAUAGAGUGUGAC	354	6375	GUCACACUCUAUGACUUCC	2005
	355	6375	CGUGAAAACUACCGAAGUU	355	6393	AACUUCGGUAGUUUUCACG	2006
UGUAGGCAAUGUCAUACUU 3	356	6393	UGUAGGCAAUGUCAUACUU	356	6411	AAGUAUGACAUUGCCUACA	2007
-	357	6411	UAAACCAUCAGAUGAAGGU	357	6459	ACCUUCAUCUGAUGGUUUA	2008
	358	6429	UGUUAAAGUAACACAAGAG	358	6447	cucungugunacunnaaca	2009
ncnn	359	6447	GUUAGGUCAUGAGGAUCUU	329	6465	AAGAUCCUCAUGACCUAAC	2010
GGAA	360	6465	UAUGGCUGCUUAUGUGGAA	360	6483	UUCCACAUAAGCAGCCAUA	2011
	361	6483	AAACACAAGCAUUACCAUU	361	6501	AAUGGUAAUGCUUGUGUUU	2012
1	362	6501	UAAGAAACCUAAUGAGCUU	362	6519	AAGCUCAUUAGGUUUCUUA	2013
UNNA	363	6519	UUCACUAGCCUUAGGUUUA	363	6537	UAAACCUAAGCUAGUGAA	2014
ICAU	364	6537	AAAAACAAUUGCCACUCAU	364	6555	$\overline{}$	2015
\dashv	365	6555	UGGUAUUGCUGCAAUUAAU	365	6573		2016
UAGUGUUCCUUGGAGUAAA 3	366	6573	UAGUGUUCCUUGGAGUAAA	366	6591	UNUACUCCAAGGAACACUA	2017
\neg	367	6591	AAUUUUGGCUUAUGUCAAA	367	6099	UUUGACAUAAGCCAAAAUU	2018
7	368	6099	ACCAUUCUUAGGACAAGCA	368	6627	uecuuenccuvveavueen	2019
AAU	369	6627	AGCAAUUACAACAUCAAAU	369	6645	_	2020
AGCA	370	6645	UUGCGCUAAGAGAUUAGCA	370	6663		2021
CAAU	371	6663	ACAACGUGUGUUAACAAU	371	6681		2022
UNAUAUGCCUUAUGUGUUU 3	372	6681	UNAUAUGCCUUAUGUGUUU	372	6699	AAACACAUAAGGCAUAUAA	2023

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2024	2020	9707	707/	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	202	2053	2054	2055	2056	202	2058	2059	2060	2061	2062	2063	2064	2065
CAAUUGGAACAAUAAUGUA	ACCOCONAGOAAAGOACAC	OCUAAUUCUAGAAUUGGUA	AGUUGUAGGUAGUGAAGCU	AACACUAUUUUUAGCAAUA	UAAUUUAGCAACACUCUUA	AAUGCCGGCAUCCAAACAU	GGGUGACUUCACAUAAUUA	GAACAAUUUAGAAAAUUUG	UAGCCACAUAGCGAUUGUG	GCAAAUACUUAACAACAAU	ACAGAUUAGAGAACCUAAG	ACCAAAAGCAGCAGUUACA	AAAAUUAGAUAAGAGUACA	ACAAUAAGAAGGAGCACCA	CAAUUCUCUAACGCCAUUA	GUUAGACGAAUUAAGAUAC	GAAAUCCAUAGUAGUAACG	AGGAAAGAACCUUCACAG	ACUUAAACAAAUGCUGCAA	AUCAAGGGAGUCUAAUCCA	UUCAAGAGCUGGAUAAGAA	AAUCGUCACCUGAAUGGUU	GUCUAGCUUGUACGAUGAA	CAGACCUAAAAUUGUCAAG	CAAAACCCACUCAGCGGCC	UGUGAACAACAUAUAUGCC	UAAUAAAUAAAAGAAUUUU	CAUUAUAGCUGAAAGACCU	AUAGCCAAAGAACACCUGC	GAUGAAAUGACUAGCAAAA	CAUGAGCCAAGAAUUGCUG	AAUACUAAUGAUAAACCAC	AACGGGUGCCAUUUGUACA	CAUCCUAACCAUUGCAGAA	AGAAGCAAAGAAGAUGUAC	CUUCCAUAUGUAGUAGAAA	CAUGAUAUGAACAUAGCUC	CGAAGAGGUGCAACCAUCC	AUAGCACAUCAUGCAAGUC	UGUGGCACGAUUGCGCUUA	AGUUGUACACUCAACGCGU
6717	6753	56/0	1//9	6489	6807	6825	6843	6861	688	6897	6915	6933	6951	6969	2869	7005	7023	7041	7059	7077	2002	7113	7131	7149	7167	7185	7203	7221	7239	7257	7275	7293	7311	7329	7347	7365	7383	7401	7419	7437	7455
373	375	373	3/6	377	378	379	380	381	382	383	384	385	386	387	388	389	330	391	392	393	394	395	396	397	398	399	400	401	402	403	404	405	406	407	408	409	410	411	412	413	414
UACAUUAUUGUUCCAAUUG	GUGUACUUUACUAAAAGU	UACCAAUUCUAGAAUUAGA	AGCUUCACUACCUACA	UAUUGCUAAAAAUAGUGUU	UAAGAGUGUUGCUAAAUUA	AUGUUUGGAUGCCGGCAUU	UAAUUAUGUGAAGUCACCC	CAAAUUUUCUAAAUUGUUC	CACAAUCGCUAUGUGGCUA	AUUGUUGUUAAGUAUUUGC	CUUAGGUUCUCUAAUCUGU	UGUAACUGCUGCUUUUGGU	UGUACUCUUAUCUAAUUUU	UGGUGCUCCUUCUUAUUGU	UAAUGGCGUUAGAGAAUUG	GUAUCUUAAUUCGUCUAAC	CGUUACUACUAUGGAUUUC	CUGUGAAGGUUCUUUUCCU	UUGCAGCAUUUGUUUAAGU	UGGAUUAGACUCCCUUGAU	UUCUUAUCCAGCUCUUGAA	AACCAUUCAGGUGACGAUU	UUCAUCGUACAAGCUAGAC	CUUGACAAUUUUAGGUCUG	GCCCCUCAGUGGGUUUUG	GGCAUAUAUGUUGUUCACA	AAAAUUCUUUUAUUAUUA	AGGUCUUUCAGCUAUAAUG	GCAGGUGUUCUUUGGCUAU	UUUUGCUAGUCAUUCAUC	CAGCAAUUCUUGGCUCAUG	GUGGUUUAUCAUUAGUAUU	UGUACAAAUGGCACCCGUU	UNCUGCAAUGGUUAGGAUG	GUACAUCUUCUUUGCUUCU	UUUCUACUACAUAUGGAAG	GAGCUAUGUUCAUAUCAUG	GGAUGGUUGCACCUCUUCG	GACUUGCAUGAUGUGCUAU	UAAGCGCAAUCGUGCCACA	ACGCGUUGAGUGUACAACU
6699	6735	0/32	6/33	6771	6289	6807	6825	6843	6861	6879	6897	6915	6933	6951	6969	6987	7005	7023	7041	7059	7077	7095	7113	7131	7149	7167	7185	7203	7221	7239	7257	7275	7293	7311	7329	7347	7365	7383	7401	7419	7437
373	375	373	3/0	377	378	379	380	381	382	383	384	385	386	387	388	389	390	391	392	393	394	395	396			366	400	401	402	403	404	405	406	407	408	409	410	411	412	413	414
UACAUUAUUGUUCCAAUUG	GUGUACUUNACUAAAAGU	ACCI III CACI IA CALACIA CALACA	AGCUUCACUACCUACAACU	UAUUGCUAAAAUAGUGUU	UAAGAGUGUUGCUAAAUUA	AUGUNUGGAUGCCGGCAUU	UAAUUAUGUGAAGUCACCC	CAAAUUUUCUAAAUUGUUC	CACAAUCGCUAUGUGGCUA	AUUGUUGUUAAGUAUUUGC	CUUAGGUUCUCUAAUCUGU	UGUAACUGCUGCUUUUGGU	UGUACUCUUAUCUAAUUUU	UGGUGCUCCUUCUUAUUGU	UAAUGGCGUUAGAGAAUUG	GUAUCUUAAUUCGUCUAAC	CGUUACUACUAUGGAUUUC	CUGUGAAGGUUCUUUUCCU	UUGCAGCAUUUGUUUAAGU	UGGAUUAGACUCCCUUGAU	UUCUUAUCCAGCUCUUGAA	AACCAUUCAGGUGACGAUU	UUCAUCGUACAAGCUAGAC	CUUGACAAUUUUAGGUCUG	GGCCGCUGAGUGGGUUUUG	GGCAUAUGUUGUUCACA	AAAAUUCUUUUAUUAUUA	AGGUCUUUCAGCUAUAAUG	GCAGGUGUUCUUUGGCUAU	UNUNGCUAGUCAUUCAUC	CAGCAAUUCUUGGCUCAUG	GUGGUUUAUCAUUAGUAUU	UGUACAAAUGGCACCCGUU	UNCUGCAAUGGUUAGGAUG	GUACAUCUUCUUUGCUUCU	UUUCUACUACAUAUGGAAG	GAGCUAUGUUCAUAUCAUG	GGAUGGUUGCACCUCUUCG	GACUUGCAUGAUGUGCUAU	UAAGCGCAAUCGUGCCACA	ACGCGUUGAGUGUACAACU
6699	6735	07.53	0/33	6//1	6289	6807	6825	6843	6861	6889	6897	6915	6933	6951	6969	2869	7005	7023	7041	7059	7077	7095	7113	7131	7149	7167	7185	7203	7221	7239	7257	7275	7293	7311	7329	7347	7365	7383	7401	7419	7437

7455	UAUUGUUAAUGGCALIGAAG	415	7155		14.5	77.10		
7473	GAGALICIIIICHALIGUCHALI	$^{+}$	7/73	CACALICITICITATIONS	413	74/3	CUUCAUGCCAUUAACAAUA	2066
7491		+	7473	I GCA A I I GCA GGCCC I GCC	410	7500	AUAGACAUAGAAGAUCUC	2067
7509	CUUCIIGCAAGACIICACAAII	╁	7500	CHICACONOCIONACIONACIONACIONACIONACIONACIO	41/	7509	GCCACGCCUCCAUUGCA	2068
7527	UUGGAAUUGUCUCAAIIIGII	419	7527	III GGAAII IGII CHOAAIII ICII	4 4	1221	AUUGUGAGUCUUGCAGAAG	2069
7545	UGACACALILILIGEACIIGEII	420	7575	TOACACALLILLICATORIO	2 5	77.00	ACAAUUGAGACAAUUCCAA	2070
7563	UAGUACALIUCALIIAGUGAI	424	7563	14611464111641116411	420	7503	ACCAGUGCAAAAUGUGUCA	2071
7581	HGAAGIIIIGCIICALIII	╁	7504	UASUACAUUCAUUAGUGAU	421	/381	AUCACUAAUGAAUGUACUA	2072
7599	GUCACIICCAGIIIIIAAAAGA	+-	7500	GICACITICAGIIIIAAAACA	422	7599	CAAAUCACGAGCAACUUCA	2073
7617	ACCAMICAACCITACITAC	424	7617	ACCACUCCAGUULAAAAGA	423	/61/	UCUUUUAAACUGGAGUGAC	2074
7635	CCAGUCAUCGUAUAUUGUU	425	7635	CCAGINALICATION	424	7652	GUCAGUAGGGUUGAUUGGU	2075
7653	UGAUAGUGUUGCUGUGAAA	426	7653	LIGATIAGLIGITI IGCI IGAAA	420	7674	AACAAUAUACGAUGACUGG	2076
7671	AAAUGGCGCGCUUCACCUC	427	7671	AAAUGGGGGGCIIICACCIIC	420	7680	GACCITAACCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	2077
7689	CUACUUUGACAAGGCUGGU	428	7689	CUACUUGACAAGGCIIGGII	428	7707	ACCAGOO III IGIICAAAGI IAG	2070
7077	UCAAAAGACCUAUGAGAGA	429	7077	UCAAAAGACCUAUGAGAGA	429	7725	UCUCUCAUAGGIICIIIIIIIIIIIIIIIIIIIIIIIIII	2080
7725	ACAUCCGCUCUCCCAUUUU	430	7725	ACAUCCGCUCUCCCAUUUU	430	7743	AAAAUGGGAGAGCGGAUGU	2081
//43	UGUCAAUUUAGACAAUUUG	431	7743	UGUCAAUUUAGACAAUUUG	431	7761	CAAAUUGUCUAAAUUGACA	2082
1/61	GAGAGCUAACAACACUAAA	432	7761	GAGAGCUAACACACUAAA	432	7779	UNUAGUGUUGUUAGCUCUC	2083
6111	AGGUUCACUGCCUAUUAAU	433	7779	AGGUUCACUGCCUAUUAAU	433	7677	AUUAAUAGGCAGUGAACCU	2084
7015	UGUCAUAGUUUUGAUGGC	434	7797	UGUCAUAGUUUUUGAUGGC	434	7815	GCCAUCAAAAACUAUGACA	2085
(010)	CAAGUCCAAAUGCGACGAG	435	7815	CAAGUCCAAAUGCGACGAG	435	7833	CUCGUCGCAUUUGGACUUG	2086
7053	GUCUGCUUCUAAGUCUGCU	436	7833	GUCUGCUUCUAAGUCUGCU	436	7851	AGCAGACUUAGAAGCAGAC	2087
1007	GOLOGOGOACOACAGOCAG	437	7851	UUCUGUGUACUACAGUCAG	437	7869	CUGACUGUAGUACACAGAA	2088
7887	SCUGAUGUGCCAACCUAUU	438	7869	GCUGAUGUGCCAACCUAUU	438	7887	AAUAGGUUGGCACAUCAGC	2089
7005	UCUGUUGCUUGACCAAGCU	439	/88/	Ucueuuecuueaccaaecu	439	7905	AGCUUGGUCAAGCAGA	2090
7023	ACAUGA DE LA COLOGICA DEL COLOGICA DEL COLOGICA DE LA COLOGICA DEL C	440	7905	UCUUGUAUCAGACGUUGGA	40	7923		2091
70/1	COLLIANCALICITICALICOL	441	7573	AGAUAGUACUGAAGUUUCC	4	7941	GGAAACUUCAGUACUAUCU	2002
7050	LILIVITION ACADED IN 194	442	7941	CGUUAAGAUGUUUGAUGCU	442	7959		2093
7077	ACCAACT III IACI ICIII IOOLI	443	7959	UNAUGUCGACACCUUUUCA	443	7977		2094
7005	1 A LOCA A A A CHILLA A COCA	444	7,67	AGCAACUUUAGUGUUCCU	444	7995		2095
8013	ACIII GIII GOLIACACO I CAC	C#4.	2867	UAUGGAAAAACUUAAGGCA	445	8013	_	2096
8031	CAGCGAGIIIAGCAACOCAC	440	8013	ACUUGUUGCUACAGCUCAC	446	8031		2097
8040	SACCOMPANAGED IN THE PROPERTY OF THE PROPERTY	441	003	CAGCGAGUDAGCAAAGGGU	447	8049	<u></u>	2098
2500	SOLUTION ACCUONAGE USOLUTION ACCUONDED	440	8049	UGUAGCUUUAGAUGGUGUC	448	8067	GACACCAUCUAAAGCUACA	2099
7000	CCUUUCUACAUUCGUGUCA	449	8067	CCUUUCUACAUUCGUGUCA	449	8085	UGACACGAAUGUAGAAAGG	2100
0000	AGCUGCCCGACAAGGUGUU	450	8085	AGCUGCCCGACAAGGUGUU	450	8103		2101
0103	OSOUGAUACCGAUGUUGAC	451	8103	UGUUGAUACCGAUGUUGAC	451	8121	<u> </u>	2102
0121	CACAAAGGAUGUAUUGAA	452	8121	CACAAAGGAUGUUAUUGAA	452	8139	UUCAAUAACAUCCUUUGUG	2103
0153	AUGUCUCAAACUUUCACAU	453	8139	AUGUCUCAAACUUUCACAU	453	8157		2104
8175	GACAGGIGACAGIIIICIIAAC	424	8157	UCACUCUGACUUAGAAGUG	454	8175		2105
8103	CAALIIIICALICALICALIALI	453 77	81/5	GACAGGUGACAGUUGUAAC	455	8193	-	2106
,	CANOCAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	420	8193	CAAUUUCAUGCUCACCUAU	456	8211	AUAGGUGAGCAUGAAAUUG	2107

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2150	2151	2152	2153	2154	2155	2156	2157	2158	2159	2160	2161	2162	2163	2164	2165	2166	2167	2168	2169	2170	2171	2172	2173	2174	2175	2176	2177	2178	2179	2180	2181	2182	2183	2184	2185	2186	2187	2188	2189	2190	2191
GUCAUAACAAUAUGGCACA	ACCCUCUAGCAAAUUAGUG	CUCACUAUAAGAAAUAGAA	ACGAGUGUCUGGACGAAGC	ACCAUCCAUAAGCACAUAA	AGGAAACUGUAUGAUGGAA	ACCCUCCAGGUAAGUGUUA	UGUUACUACUCUAACAGAA	GUACUCAGCAUCAAAAGUU	GCAUGUACCAUGUCUACAG	ACCUACUCUGACCUUUCG	ACUGGUAGAUAGGCAAAUA	AUUAAGAACCCAUCUACCA	AGCUCUGUAAUGCUCAUUA	ACAGAAACUCCUGAUAGA	AUUCAUCGCAUCAACACCA	AAAGAUGUUAGCUAUGAGA	AGGUUGCACAAGAGGAGUA	CACAUCUAAAGCACCCACA	AGCCACUACUGAAGCAGAC	UAUGGCAAUAAUACCACCA	GGCAGCACAAGUCACCAAU	GAAUUUCAUAAAGUAGUAG	CUCACCAAAAACACGUCUG	AGCAACAUGGUUGUAC	AAACAAAGUGCAUUAGCA	UAUAGUGAAAGACAUCAAA	AGCUGGUACCAGACAGAGU	UCCCGGCAGAAGCUGUAA	GUAAAAGACUGAGUAGACU	AUAGAAUGUCAAGUACAAG	UGAAACAUCAUUGGUGAAA	UUGAAGGUGAGCCAAGAAU	AGAAAACAUGGCAAACCAU	CCAAAAAGGCACAAUAGGA	UACAUAGAUUGCUGUUAUC	CUUCAGAGAAAUACAGAAU	AAAGAACCAAUGGCAGUGC	UUUCCUAAGAUAGUUGUUA	UCCAUUAAACAUGACUCUU	GAAGGUACUAAAUGUAACU	ACACAAAGCAGCCUCCUCG
8985	9003	9021	9039	9057	9075	9093	9111	9129	9147	9165	9183	9201	9219	9237	9255	9273	9291	9309	9327	9345	9363	9381	9399	9417	9435	9453	9471	9489	9507	9525	9543	9261	9579	9597	9615	9633	9651	6996	9687	9705	9723
499	200	501	205	203	504	202	206	207	208	509	510	511	512	513	514	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	537	538	539	540
UGUGCCAUAUUGUUAUGAC	CACUAAUUUGCUAGAGGGU	UUCUAUUCUUAUAGUGAG	GCUUCGUCCAGACACUCGU	UNAUGUGCUUAUGGAUGGU	UUCCAUCAUACAGUUUCCU	UAACACUUACCUGGAGGGU	UUCUGUUAGAGUAGUAACA	AACUUUUGAUGCUGAGUAC	CUGUAGACAUGGUACAUGC	CGAAAGGUCAGAAGUAGGU	UAUUUGCCUAUCUACCAGU	UGGUAGAUGGGUUCUUAAU	UAAUGAGCAUUACAGAGCU	UCUAUCAGGAGUUUUCUGU	UGGUGUUGAUGCGAUGAAU	UCUCAUAGCUAACAUCUUU	UACUCCUCUUGUGCAACCU	UGUGGGUGCUUUAGAUGUG	GUCUGCUUCAGUAGUGGCU	UGGUGGUAUUAUUGCCAUA	AUUGGUGACUUGUGCUGCC	CUACUACUUNAUGAAAUUC	CAGACGUGUUUUUGGUGAG	GUACAACCAUGUUGUUGCU	UGCUAAUGCACUUUUGUUU	UNUGAUGUCUUUCACUAUA	ACUCUGUCUGGUACCAGCU	UNACAGCUUUCUGCCGGGA	AGUCUACUCAGUCUUUAC	CUUGUCACAUUCUAU	UUUCACCAAUGAUGUUUCA	AUUCUUGGCUCACCUUCAA	AUGGUUUGCCAUGUUUUCU	Uccuauugugccuuuuugg	GAUAACAGCAAUCUAUGUA	AUUCUGUAUUUCUCUGAAG	GCACUGCCAUUGGUUCUUU	UAACAACUAUCUUAGGAAA	AAGAGUCAUGUUUAAUGGA	AGUUACAUUUAGUACCUUC	CGAGGAGGCUGCUUUGUGU
8967	8985	9003	9021	9039	9057	9075	9093	9111	9129	9147	9165	9183	9201	9219	9237	9255	9273	9291	9309	9327	9345	9363	9381	9399	9417	9435	9453	9471	9489	/006	9525	9543	19261	9579	9597	9615	9633	9651	9669	9687	9705
499	200	5	202	203	204	505	206	207	208	209	510	511	512	513	514	515	516	517	518	519	220	521	522	523	524	525	526	527	228	670	230	25	232	533	534	535	536	537	538	539	540
UGUGCCAUAUUGUUAUGAC	CACUAAUUUGCUAGAGGGU	GCIIICCIICCACACACACI	GCOCCAGACACOCGO	UNAUGUGCUUAUGGAUGGU	UNCCAUCAUACAGOUUCCU	UAACACUUACCUGGAGGGU	UUCUGUUAGAGUAGUAACA	AACUUUUGAUGCUGAGUAC	CUGUAGACAUGGUACAUGC	CGAAAGGUCAGAAGUAGGU	UAUUUGCCUAUCUACCAGU	UGGUAGAUGGGUUCUUAAU	UAAUGAGCAUUACAGAGCU	UCUAUCAGGAGUUUUCUGU	UGGUGUUGAUGCGAUGAAU	UCUCAUAGCUAACAUCUUU	UACUCCUCUUGUGCAACCU	Ocucecuorunagaugus	<u>eucuecuucaguaguagen</u>	UGGUGGUAUUAUUGCCAUA	AUUGGUGACUUGUGCUGCC	CUACUACUUAUGAAAUUC	CAGACGUGUUUUGGUGAG	GUACAACCAUGUUGUUGCU	UGCUAAUGCACUUUGUUU	UNUGAUGUCUUCACUAUA	ACUCUGUCUGGUACCAGCU	VOACAGCOUNCUGCCGGA	CHICHACHERACHIDIAN	III II ICACCAALICALICIII II ICA	ALI ICI II COLI ICA COLI ICA A	AUCCOUGGCOCACCOOCAA	HOCHAI I OHOOCH III III OO	Occudade describing de la company de la comp	GAUAACAGCAAUCUAUGUA	AUCUGUAUUCUCUGAAG	GCACUGCCAUUGGUUCUUU	UAACAACUAUCUUAGGAAA	AAGAGUCAUGUUAAUGGA	AGUACAUUNAGUACCUUC	CeAeGAGCOGCOOOGOGO
8967	282	9003	202	9039	9037	200	3033	2 6	8718	9147	9165	200	370	87.8	323/	6760	9273	328	9309	932/	9345	9363	9381	9299	1000	3433	+	+	0507	t	0543	╁	+	+	+	+	3033	+	9009	+	\dashv

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2192	2193	2194	2195	2196	2197	2198	2199	2200	2201	2202	2203	2204	2205	2206	2207	2208	2209	2210	2211	2212	2213	2214	2215	2216	2217	2218	2219	2220	2221	2222	2223	2224	2225	2226	2227	2228	5229	2230	2231	2232	2233
CUUGUUGAGCAAAAAGGUA	CAAUUUAGGUACAUUUCC	CAACAGUGUCUCGCUACGC	GUUAUACUGUGUAAGUGGC	AUAUAGAGCAAGAUACCUG	GAAAUACUUGUACUUGUUA	AGUAUCUAAGGCUCCACUG	UGCUUCACGAUAGCUGGUA	UGCUAAGUGGCAGCAAGCU	AAAGUCAUUUAGAGCCUUU	AUCAGCACCUGAGUUGCUA	UGGUGGUUGGUAGAGACA	AGAAGUGAUUGAUGUCUGU	ACCACUCUGCAGAACAGCA	GAAUGCCAUUUUCCUAAAA	_	UACUUGUACCAUGCACCCU	AGUUGUAGUUCCACAGGUU	CAACCACAGUCCAUUAAGA	ш	GCAAAUGACAUGUCUUGGA		\vdash	GGAUUUGCGAAUGAGCAGA	AACAAGAAAGCUAUGGUUG		_	CAGACAAUUUUGCAUAGAA	AUCAACUUUAAGCCUAAGC	UGUCUUAGGGUUAGAAGUA	_	_	UGCUAGAACUGAAAAUGUU	UGGUGAACCAUUGUAGCAU	ACACUGAUAAACACCAGAU	AUGAUUAGGUCUCAUGGCA	GAAAGAACCUUUAAUGGUA	ACCACAUGAUCCAUUAAGG	AAUGUUAAAACCAACACUA	AGACACGCAAUCAUAAUCA	AUGAUGCAUAUAGCAGAAA	UCCUGUUGGAAGCUCCAUA
9741	9759	9777	9795	9813	9831	9849	29867	9885	9903	9921	9939	9957	9975	9993	10011	10029	10047	10065	10083	10101	10119	10137	10155	10173	10191	10209	10227	10245	10263	10281	10299	10317	10335	10353	10371	10389	10407	10425	10443	10461	10479
24	242	543	544	545	546	547	548	549	550	551	552	553	554	555	256	222	258	559	260	561	562	563	564	565	999	267	568	569	570	571	572	573	574	575	929	27.7	218	629	280	581	582
UACCUUUUUGCUCAACAAG	GGAAAUGUACCUAAAAUUG	GCGUAGCGAGACACUGUUG	GCCACUUACACAGUAUAAC	CAGGUAUCUUGCUCUAUAU	UAACAAGUACAAGUAUUUC	CAGUGGAGCCUUAGAUACU	UACCAGCUAUCGUGAAGCA	AGCUUGCUGCCACUUAGCA	AAAGGCUCUAAAUGACUUU	UAGCAACUCAGGUGCUGAU	UGUUCUCUACCACCACCA	ACAGACAUCAGUUCU	UGCUGUUCUGCAGAGUGGU	UUUUAGGAAAAUGGCAUUC	CCCGUCAGGCAAAGUUGAA	AGGGUGCAUGGUACAAGUA	AACCUGUGGAACUACAACU	UCUUAAUGGAUUGUGGUUG			CACAGCAGAAGACAUGCUU		UCUGCUCAUUCGCAAAUCC	_		\dashv	UUCUAUGCAAAAUUGUCUG	\dashv		-1	\dashv		-		UGCCAUGAGACCUAAUCAU	UACCAUUAAAGGUUCUUUC		UAGUGUUGGUUUUAACAUU	\vdash	-	UAUGGAGCUUCCAACAGGA
9723	9741	9759	9777	616	9813	9831	9849	2986	9885	9903	9921	9939	9957	9975	9993	10011	10029	10047	10065	10083	10101	10119	10137	10155	10173	10191	10209	10227	10245	10263	10281	10299	10317	10335	10353	10371	10389	10407	10425	10443	10461
541	542	543	544	545	546	547	548	549	220	551	552	553	554	555						561	562	563	564	565	999	267	268	269	220	571	572	573	574	575	929	27.2	278	219	280	281	582
UACCUUUUUGCUCAACAAG	GGAAAUGUACCUAAAAUUG	GCGUAGCGAGACACUGUUG	GCCACUUACACAGUAUAAC	CAGGUAUCUUGCUCUAUAU	UAACAAGUACAAGUAUUUC	CAGUGGAGCCUUAGAUACU	UACCAGCUAUCGUGAAGCA	AGCUUGCUGCCACUUAGCA	AAAGGCUCUAAAUGACUUU	UAGCAACUCAGGUGCUGAU	UGUUCUCUACCACCACCA	ACAGACAUCAAUCACUUCU	UGCUGULCUGCAGAGUGGU	UUUUAGGAAAAUGGCAUUC	CCCGUCAGGCAAAGUUGAA	AGGGUGCAUGGUACAAGUA	AACCUGUGGAACUACAACU	UCUUAAUGGAUUGUGGUUG	GGAUGACACAGUAUACUGU	UCCAAGACAUGUCAUUUGC	CACAGCAGAAGACAUGCUU	UAAUCCUAACUAUGAAGAU	UCUGCUCAUUCGCAAAUCC	CAACCAUAGCUUUCUUGUU	UCAGGCUGGCAAUGUUCAA	ACUUCGUGUUAUUGGCCAU	UUCUAUGCAAAAUUGUCUG	GCUUAGGCUUAAAGUUGAU	UACUUCUAACCCUAAGACA	ACCCAAGUAUAAAUUUGUC	CCGUAUCCAACCUGGUCAA	AACAUUUUCAGUUCUAGCA	AUGCUACAAUGGUUCACCA	AUCUGGUGUUNAUCAGUGU	UGCCAUGAGACCUAAUCAU	UACCAUUAAAGGUUCUUUC	CCUUAAUGGAUCAUGUGGU	UAGUGUUGGUUUUAACAUU	UGAUUAUGAUUGCGUGUCU	UUUCUGCUANAUGCAUCAU	UAUGGAGCUUCCAACAGGA
9723	9741	9759	9777	9795	9813	9831	9849	2986	9885	9903	9921	9939	9957	9975		10011		10047	10065	10083	10101	10119	10137	10155	10173	10191	10209	10227	10245	10263	10281	10299	10317	10335	10353	10371	10389	10407	10425	10443	10461

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2234	2235	2236	2237	2238	2239	2240	2241	2242	2243	2244	2245	2246	2247	2248	2249	2250	2251	2252	2253	2254	2255	2256	2257	2258	2259	2260	2261	2262	2263	2264	2265	2266	2267	2268	2269	2270	2271	2272	2273	2274	2275
	-	UCUGUCAACAAAUGGACCA	UGCAGCCUGUGCAGUUUGU	UAUGGUUGUGUCUGUACCU	UGCCAAAACAUUUAAUGUU		CCACCUAUCACCAUUGAUA	GGUGAAUCUAUUAAGAAAC	AAAGUCAUUCAAAGUAGUG	CUUCAUUGCCACAAGGUUA	CAAAGGUUCAUAGUUGUAC	GUCAACAUGAUCUUGUGUC	AGAAAGAGGUCCCAAUAUG	GGCAAUUCCUGUUUGAGCA	AGCACACAUAUCUAAGACG		ACCAUUCAUACCAUUCUGC	GCUACCAAGGAUAGUACGA	CUCAUCUUCUAAAAUAGUG	_	ACCAGAGCAUUGUCUAACA	CUUACCUUGGAAGGUAACA	CUUAACAAUUUUCUUGAAC	CAUCCAAUGAUGAGUGCCC			UGACCACUGUGUACUUUGA			CAUAAUACCAAGAGUAAAU	AGCACAUGCAGCAAUUGCC	AUGCUUAACAAGCAGCAUA	GCACAAGAAUGCGUGCUUA	_	GUAAGCAACUGUUGCAAGA	CAUGUAGACCAUAUUAAAG	CAUCACCCAGCUAGCAGGC	AAGCCAUGUCAUGAUACGC	GCUAGUGUCAGCCAAUUCA	AAGCCUAUAACCAGACAAG	AUACAUAACACAAUCCUUA
10497	10515	10533	10551	10569	10587	10605	10623	10641	10659	10677	10695	10713	10731	10749	10767	10785	10803	10821	10839	10857	10875	10893	10911	10929	10947	10965	10983	11001	11019	11037	11055	11073	11091	11109	11127	11145	11163	11181	11199	11217	11235
583	584	585	286	282	288	289	290	591	265	593	594	595	296	297	298	299	009	601	602	603	604	902	909	209	809	609	610	611	612	613	614	615	616	617	618	619	620	621	622	623	624
\vdash	\dashv		ACAAACUGCACAGGCUGCA	_	AACAUUAAAUGUUUGGCA	_	UAUCAAUGGUGAUAGGUGG	_	CACUACUUUGAAUGACUUU	-	GUACAACUAUGAACCUUUG		CAUAUUGGGACCUCUUUCU	\Box				_	CACUAUUUAGAAGAUGAG		UGUUAGACAAUGCUCUGGU			-		\dashv		-	\dashv	-1	-	_	_		-	-	_	Ť	_	CUUGUCUGGUUAUAGGCUU	UAAGGAUUGUGUUAUGUAU
10479	10497	10515	10533	10551	10569	10587	10605	10623	10641	10659	10677	10695	10713	10731	10749	10767	10785	10803	10821	10839	10857	10875	10893	10911	10929	10947	10965	10983	11001	11019	11037	11055	11073	11091	11109	11127	11145	11163	11181	11199	11217
583	584	585	586	587	588	589	290	591	592	593	594	595	296	297	298	299	009	601	602	603	604	902	909	209	809	609	610	611	612	613	614	615	616	617	618	619	620	621	622	623	624
\vdash	-	UGGUCCAUUUGUUGAG		\dashv	\dashv	\rightarrow	-			UAACCUUGUGGCAAUG	GUACAACUAUGAACCU	GACACAAGAUCAUGUU	CAUAUUGGGACCUCUL	UGCUCAAACAGGAAUU	-	UGCUUUGAAAGAGCUG		UCGUACUAUCCUUGGU	CACUAUUUUAGAAGAU	GUUUACACCAUUUGAU		10875 UGUUACCUUCCAAGGUAAG	-+	_	10929 GCUUUUAACUUUCUUGACA	_	-	\dashv	\rightarrow	11019 AUUUACUCUUGGUAUUAUG	\rightarrow	-	\dashv	CUUGUUUCUGUUACCL	-	11127 CUUUAAUAUGGUCUACAUG			UGAAUUGGCUGACACU		11217 UAAGGAUUGUGUUAUGUAU

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2276	2278	2279	2280	2281	2282	2283	2284	2285	2286	2287	2288	2289	2290	2291	2292	2293	2294	2295	2296	2297	2298	2299	2300	2301	2302	2303	2304	2305	2306	2307	2308	2309	2310	2311	2312	2313	2314	2315	2316	2317
CAAAACUAAAGCUGAAGCA	+	\vdash	AAUGACAUUCAUCAGUGUC	GACUUUGUAAACAAGUGUA	UAAAGCAUUACCAUAGUAG	\vdash	AGAAAUAACUAAGGCCCAC	AGAAUAGUUAGAGGUUACA	GAUAGUCGUAACGACACCA	AGCUCUAGCUAAAAACAUG	AACACACACACACUAUA	-			-	4	_	\dashv	\dashv	\dashv	-	CAAAAGCCCCUGGGAGUUC	_	-	\vdash	-	-	-	UACCACAGAUGUGCACUUU	UUGAAGAACCGAGAGCAGU	UGACUCUACUCUAAGUUGU	_		UGCAAGAAGAAUAUCAUUG	AGCUUCAGUUGUGUCUUUU	-1	UAGCAAAACAGACAAAAGA	\sqcup	GCACAACCUAUUAAUGUCU	GUUAUCGAGCAUUUCCUCG
11253	11289	11307	11325	11343	11361	11379	11397	11415	11433	11451	11469	11487	11505	11523	11541	11559	11577	11595	11613	11631	11649	11667	11685	11703	11721	11739	11757	11775	11793	11811	11829	11847	11865	11883	11901	11919	11937	11955	11973	11991
625	627	628	629	630	631	632	633	634	635	636	637	638	639	640	<u>8</u>	642	643	644	645	646	647	648	649	650	651	652	653	654	655	656	657	658	629	099	991	662	663	994	999	999
11235 UGCUUCAGCUUUAGUUUUG 11253 GCUUAUUCUCAUGACAGCU	-	11289 UGCUGCUAGACGUGUUUGG	11307 GACACUGAUGAAUGUCAUU	11325 UACACUUGUUUACAAAGUC	11343 CUACUAUGGUAAUGCUUUA		11379 GUGGGCCUUAGUUAUUCU		11415 UGGUGUCGUUACGACUAUC	11433 CAUGUUUUAGCUAGAGCU	11451 UAUAGUGUUUGUGUGUU	\dashv	-1			\dashv	-	_	-	-		11649 GAACUCCCAGGGGCUUUUG			긤			-1	\dashv	+	\dashv	-		-	\dashv	_	11919 UCUUUUGUCUGUUUUGCUA		-1	11973 CGAGGAAAUGCUCGAUAAC
625 626	627	628	629	630	631	632	633	634	635	636	637	638	639	640	25	642	643	644	645	646	647	648	649	650	651	652	653	654	655	929	/29	828	629	099	661	995	663	664	992	999
11235 UGCUUCAGCUUUAGUUUUG 11253 GCUUAUUCUCAUGACAGCU	+	UGCUGCUAGACGUGUL	-		-	-	GUGGGCCUUAGUUAUI	UGUAACCUCUAACUAU		CAUGUUUUAGCUAGA	-	-	-	-	UGUUUAUUGUUUCUUA	CUAUUGUUGCUGCUGC	\dashv	-	GCUUACUCUUGGUGUI	-+	-+	GAACUCCCAGGGGCUL	GCCUCCUAAGAGUAGU	\dashv	\rightarrow	11721 UGGAGGUAAACCAUGUAUC	-		AAAGUGCACAUCUGUG	Acuecucuceguucu	-	_	11847 ACAAUGUGUACAACUCCAC	-	AAAAGACACAACUGAAG	UUUCGAGAAGAUGGUU	11919 UCUUUUGUCUGUUUUGCUA	AUCCAUGCAGGGUGCL	AGACAUUAAUAGGUUG	11973 CGAGGAAAUGCUCGAUAAC

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693 12459 UGCACUCUGGGAAAUCCAG
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2360	2362	2363	2364	2365	2366	2367	2368	2369	2370	2371	2372	2373	2374	2375	2376	2377	2378	2379	2380	2381	2382	2383	2384	2385	2386	2387	2388	2389	2390	2391	2392	2393	2394	2395	2396	2397	2398	2399	2400	2401
UCUAGCCCAUUUGAGAUCU		-	UGUGUCUGUAACAAACCUA	CACUUUAGGCCCUUUUGGU	GAUGAAGUACAAGUAUUUC	┢	CAGCACCAUACCUCUAUUU	UGUAGCAGCUAAACUGCCC	UCCAGCCUGAAGACGUACU		AAGCACAGUUGAAUUGGCA	UGCAAAAGCACAGAAGGAA	ш	UGCUAGGUAAUCCUUAUAU	GAUUGGUUGUCCUCCACUU	CAUCUUCACACAGUUGGUG	ACCAGUGUGUGUACACAC	AGUAAUUGCCUGUCCUGUA	GUUAGCUUCUGGUGUUACA	AAAGGACUCUUGGUCCAUG	Ь	GUGGCAUCUACAAUACAGA	AGGAUUGGAUGGUCAAUG	\vdash	ш		_					AGACUGCAUCAAGGGUUCG	AAACGUUGAUGCAUCCGCA	CACCGCAAACCCGUUUAAA	UAAGACGGGCUGCACUUAC	UGCCUGUGCCGCACGGUGU	AGACGACAUCAGUACUAGU	AAAUAUCAAAAGCCCUGUA	CAGCAACUUUUCGUUGUA	UNAGGAACUUUGCAAAACC
12765	12801	12819	12837	12855	12873	12891	12909	12927	12945	12963	12981	12999	13017	13035	13053	13071	13089	13107	13125	13143	13161	13179	13197	13215	13233	13251	13269	13287	13305	13323	13341	13359	13377	13395	13413	13431	13449	13467	13485	13503
709	711	712	713	714	715	716	717	718	719	720	721	722	723	724	725	726	727	728	729	730	731	732	733	734	735	736	737	738	739	740	741	742	743	744	745	746	747	748	749	750
AGAUCUCAAAUGGGCUAGA	-	AGAACUGGAACCACCUUGU	UAGGUUUGUUACAGACACA	ACCAAAAGGGCCUAAAGUG	GAAAUACUUGUACUUCAUC	_	AAAUAGAGGUAUGGUGCUG	GGGCAGUUUAGCUGCUACA	AGUACGUCUUCAGGCUGGA	AAAUGCUACAGAAGUACCU	UGCCAAUUCAACUGUGCUU	UNCCUNCUGUGCUUUUGCA	AGUAGACCCUGCUAAAGCA	AUAUAAGGAUUACCUAGCA	AAGUGGAGGACAACCAAUC	CACCAACUGUGUGAAGAUG	GUUGUGUACACACACUGGU	UACAGGACAGCAAUUACU	UGUAACACCAGAAGCUAAC	CAUGGACCAAGAGUCCUUU	UGGUGGUGCUUCAUGUUGU	UCUGUAUUGUAGAUGCCAC	CAUUGACCAUCCAAAUCCU	UAAAGGAUUCUGUGACUUG	GAAAGGUAAGUACGUCCAA	AAUACCUACCACUUGUGCU	UAAUGACCCAGUGGGUUUU	UACACUUAGAAACACAGUC	CUGUACCGUCUGCGGAAUG	GUGGAAAGGUUAUGGCUGU	UAGUUGUGACCAACUCCGC	CGAACCCUUGAUGCAGUCU	UGCGGAUGCAUCAACGUUU	UNUAAACGGGUUUGCGGUG	GUAAGUGCAGCCCGUCUUA	ACACCGUGCGGCACAGGCA	ACUAGUACUGAUGUCGUCU	UACAGGGCUUUUGAUAUUU	UACAACGAAAAAGUUGCUG	GGUUUUGCAAAGUUCCUAA
12747	12783	12801	12819	12837	12855	12873	12891	12909	12927	12945	12963	12981	12999	13017	13035	13053	13071	13089	13107	13125	13143	13161	13179	13197	13215	13233	13251	13269	13287	13305	13323	13341	13359	13377	13395	13413	13431	13449	13467	13485
709	711	712	713	714	715	716	717	718	719	720	721	722	723	724	725	726	727	728	729	730	731	732	733	734	735	736	737	738	739	740	741	742	743	744	745	746	747	748	749	750
47 AGAUCUCAAAUGGGCUAGA	UACAGGUACAAUUUAG	\vdash	19 UAGGUUUGUUACAGACACA	-1		-	AAAUAGAGGUAUGGU			AAAUGCUACAGAAGU/	UGCCAAUUCAACUGU	-1	AGUAGACCCUGCUAA/	AUAUAAGGAUUACCUX	AAGUGGAGGACAACC,	CACCAACUGUGUGAA	GUUGUGUACACACAC	UACAGGACAGGCAAUI	UGUAACACCAGAAGCL		-	UCUGUAUUGUAGAUG	CAUUGACCAUCCAAAL	\rightarrow	GAAAGGUAAGUACGU	AAUACCUACCACUUGL	UAAUGACCCAGUGGG	-	-+		\rightarrow		-+		GUAAGUGCAGCCCGU	{		-	-+	S GGUUUUGCAAAGUUCCUAA
12747	12783	12801	12819	12837	12855	12873	12891	12909	12927	12945	12963	12981	12999	13017	13035	13053	13071	13089	13107	13125	13143	13161	13179	13197	13215	13233	13251	13269	13287	13305	13323	13341	13359	13377	13395	13413	13431	13449	13467	13485

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AGCGACAGCAAUUAGUUUU	AGUCUAAUAAAUUGCCUUC	UCUUAACUACAAAGUAAGA	AGUUAGACAUAGUAUGCCU	UAGUCUCCUCAUGUUGGUA	cuunaaccaaguuanaaan	CAGCAACCGCUGGACAAUC	ACUUGAAAAAGUCAUGGAC	UGUCACCAUCUACUCUAAA	GUGAUAUGUGGUACCAU	AUUUAGUUAGACGCUGACG	CUAAAUCAGCCAUUGUGUA	AAUGACGUAGAGCAUAGAC	CACAAUUACCCUCAUCAAA	GUAUUUCUUUUAAUGUAUC	AGCAAUUGUAUGUGACGAG	UGAAAUAAUCAUCACA	CAUACCAAUCCUUCUUAUU	CAGGAUUCUCUACGAAGUC	CAUAUACGCGUAAGAUGUC	CACGCUCACCUAAGUUAGC	UNAAUAAUGAUUGGCGUAC	CGCAGAAUUGUACAGUCUU	CUGCAUCACGCAUAGCAUC	UCAGUACGCCUACAAUGCC	GAUCCUGAUUAUCUAAUGU	cenaccaeuucccauuaae	GUACGAAAUCACCGAAAUC	CGCAGCCUGGUGCUACUUG	AAUCCACAAUAGGAACUCC	UCAGCAAUGAGUAAUAUGA	UCAAAGUGAGGAUGGGCAU	CAGCAGCCAAUGCCCUAGU	CAGCAUCCAUAUGGGACUC	UAAGUGGUUUUGCGAGAUC	ucagcaaaucccacuuaau	CUUCCGUAAAAUCAUAUUU	CGAAGAGACAAAGUCUCUC	AAUAUUUAAAAUAACGGUC		AACAGUUAAUACAAUUGGG
13521	13557	13575	13593	13611	13629	13647	13665	13683	13701	13719	13737	13755	13773	13791	13809	13827	13845	13863	13881	13899	13917	13935	13953	13971	13989	14007	14025	14043	14061	14079	14097	14115	14133	14151	14169	14187	14205	14223	14241	14259
751	753	754	755	756	157	758	759	760	761	762	763	764	765	992	191	292	692	022	771	772	773	774	775	922	777	778	779	780	781	782	783	784	785	286	787	788	789	790	791	792
13503 AAAACUAAUUGCUGUCGCU	╁	⊢	13575 AGGCAUACUAUGUCUAACU	13593 UACCAACAUGAAGAGACUA	13611 AUUUAUAACUUGGUUAAAG	-	13647 GUCCAUGACUUUUUCAAGU	13665 UUUAGAGUAGAUGGUGACA				3737 GUCUAUGCUCUACGUCAUU	-	13773 GAUACAUUAAAAGAAAUAC	3791 CUCGUCACAUACAAUUGCU	3809 UGUGAUGAUGAUUAUUUCA	13827 AAUAAGAAGGAUUGGUAUG		$\overline{-}$	-	3899 GUACGCCAAUCAUUAUUAA	_	\dashv		-						14079 AUGCCCAUCCUCACUUUGA		14115 GAGUCCCAUAUGGAUGCUG	14133 GAUCUCGCAAAACCACUUA	14151 AUUAAGUGGGAUUUGCUGA	-1	14187 GAGAGACUUUGUCUCUUCG			14241 CCCAAUUGUAUUAACUGUU
751 1	ļ <u>.</u>	-	755 1	756 1	_			760 1			763		`		767	768	769		771		_													786 1	787		789			792
NACAGGAGAGGAUGAGG UUCCAGGAGGAAGGAUGAGG	+	1	-		AUUUAUAACUUGGUUAAAG	-	-	UUUAGAGUAGAUGGU	_	CGUCAGCGUCUAACUAAAU	UACACAAUGGCUGAUUUAG	GUCUAUGCUCUACGU	UUUGAUGAGGGUAAUI	GAUACAUUAAAAGAAA	CUCGUCACAUACAAUL	UGUGAUGAUGAUUAUI		GACUUCGUAGAGAAUCCUG	GACAUCUNACGCGUANANG	GCUAACUUAGGUGAGCGUG	GUACGCCAAUCAUUAUUAA	AAGACUGUACAAUUCUGCG	GAUGCUAUGCGUGAUGCAG	GGCAUUGUAGGCGUACUGA	ACAUUAGAUAAUCAGGAUC	CUUAAUGGGAACUGGUACG	GAUUUCGGUGAUUUCGUAC	-+	-	UCAUAUUACUCAUUGC			-	_	-			GACCGUUAUUUUAAAU	UGGGACCAGACAUACCAUC	CCCAAUUGUAUUAACUGUU
13503	13539	13557	13575	13593	13611	13629	13647	13665	13683	13701	13719	13737	13755	13773	13791	13809	13827	13845	13863	13881	13899	13917	13935	13953	13971	13989	14007	14025	14043	14061	14079	14097	14115	14133	14151	14169	14187	14205	14223	14241

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15015	GCUCGCACCGUAGCUGGUG	835	15015	GCUCGCACCGUAGCUGGUG	835	15033	CACCAGCHACGGHGCGAGC	2486
15033		836	15033	GUCUCUAUCUGUAGUACUA	836	15051	UAGUACUACAGAUAGAGAC	2487
15051	_	837	15051	AUGACAAAUAGACAGUUUC	837	15069	GAAACUGUCUAUUUGUCAU	2488
15069	CAUCAGAAAUUAUUGAAGU	838	15069	CAUCAGAAAUUAUUGAAGU	838	15087	ACUUCAAUAAUUUCUGAUG	2489
15087	UCAAUAGCCGCCACUAGAG	839	15087	UCAAUAGCCGCCACUAGAG	839	15105	CUCUAGUGGCGGCUAUUGA	2490
15105	GGAGCUACUGUGGUAAUUG	840	15105	GGAGCUACUGUGGUAAUUG	840	15123	CAAUUACCACAGUAGCUCC	2491
15123	GGAACAAGCAAGUUUUACG	841	15123	GGAACAAGCAAGUUUUACG	841	15141	CGUAAAACUUGCUUGUUCC	2492
15141	-	842	15141	GGUGGCUGGCAUAAUAUGU	842	15159	ACAUAUUAUGCCAGCCACC	2493
15159	1	843	15159	UUAAAAACUGUUUACAGUG	843	15177	CACUGUAAACAGUUUUUAA	2494
15177	GAUGUAGAAACUCCA	844	15177	GAUGUAGAAACUCCACACC	844	15195	GEUGUGGAGUUUCUACAUC	2495
15195		845	15195	CUUAUGGGUUGGGAUUAUC	845	15213	GAUAAUCCCAACCCAUAAG	2496
15213	CCAAAAUGUGACAGA	846	15213	CCAAAAUGUGACAGAGCCA	846	15231	UGGCUCUGUCACAUUUUGG	2497
15231		847	15231	AUGCCUAACAUGCUUAGGA	847	15249	UCCUAAGCAUGUUAGGCAU	2498
15249	AUAAUGGCCUCUCUUGUUC	848	15249	AUAAUGGCCUCUCUUGUUC	848	15267	GAACAAGAGGGCCAUUAU	2499
15267		849	15267	CUUGCUCGCAAACAUAACA	849	15285	UGUUAUGUUUGCGAGCAAG	2500
15285		820	15285	ACUUGCUGUAACUUAUCAC	850	15303	GUGAUAAGUUACAGCAAGU	2501
15303		821	15303	CACCGUUUCUACAGGUUAG	851	15321	CUAACCUGUAGAAACGGUG	2502
15321	GCUAACGAGUGUGCGCAAG	852	15321	GCUAACGAGUGUGCGCAAG	852	15339	cnnececacacncennaec	2503
15339	GUAUUAAGUGAGAUGGUCA	853	15339	GUAUUAAGUGAGAUGGUCA	853	15357	UGACCAUCUCACUUAAUAC	2504
15357	AUGUGGCGCCUCACUAU	854	15357	AUGUGUGGCGCCUCACUAU	854	15375	AUAGUGAGCCGCCACACAU	2505
15375	UAUGUUAAACCAGGUGGAA	855	15375	UAUGUUAAACCAGGUGGAA	855	15393	UUCCACCUGGUUUAACAUA	2506
15393	ACAUCAUCCGGUGAUGCUA	826	15393	ACAUCAUCCGGUGAUGCUA	826	15411	UAGCAUCACCGGAUGAUGU	2507
15411	ACAACUGCUUAUGCUAAUA	857	15411	ACAACUGCUUAUGCUAAUA	857	15429	UAUUAGCAUAAGCAGUUGU	2508
15429	AGUGUCUUNAACAUUUGUC	828	15429	AGUGUCUUNAACAUUUGUC	828	15447	-	2509
15447	CAAGCUGUUACAGCCAAUG	859	15447	CAAGCUGUUACAGCCAAUG	628	15465	CAUUGGCUGUAACAGCUUG	2510
15465	GUAAAUGCACUUCUUUCAA	860	15465	GUAAAUGCACUUCUUUCAA	098	15483	UUGAAAGAAGUGCAUUUAC	2511
15483	ACUGAUGGUAAUAAGAUAG	861	15483	ACUGAUGGUAAUAAGAUAG	861	15501		2512
15501	GCUGACAAGUAUGUCCGCA	862	15501	GCUGACAAGUAUGUCCGCA	862	15519	UGCGGACAUACUUGUCAGC	2513
15519	AAUCUACAACACAGGCUCU	863	15519	AAUCUACAACACAGGCUCU	863	15537	AGAGCCUGUGUUGUAGAUU	2514
15537		864	15537	UAUGAGUGUCUCUAUAGAA	864	15555	UUCUAUAGAGACACUCAUA	2515
15555	AAUAGGGAUGUUGAUCAUG	865	15555	AAUAGGGAUGUUGAUCAUG	865	15573		2516
15573	GAAUUCGUGGAUGAGUUUU	998	15573	GAAUUCGUGGAUGAGUUUU	998	15591	AAAACUCAUCCACGAAUUC	2517
15591	UACGCUUACCUGCGUAAAC	867	15591	UACGCUUACCUGCGUAAAC	867	15609	_	2518
15609	CAUUCUCCAUGAUGAUUC	898	15609	CAUUUCUCCAUGAUGAUUC	868	15627	_	2519
15627	CUUUCUGAUGAUGCCGUUG	869	15627	CUUUCUGAUGAUGCCGUUG	698	15645		2520
15645	GUGUGCUAUAACAGUAACU	870	15645	GUGUGCUAUAACAGUAACU	870	15663		2521
15663	UAUGCGCCUCAAGGUUUAG	871	15663	UAUGCGGCUCAAGGUUUAG	871	15681	CUAAACCUUGAGCCGCAUA	2522
15681	GUAGCUAGCAUUAAGAACU	872	15681	GUAGCUAGCAUUAAGAACU	872	15699	AGUUCUUAAUGCUAGCUAC	2523
15699	UUUAAGGCAGUUCUUUAUU	873	15699	UUUAAGGCAGUUCUUUAUU	873	15717	AAUAAAGAACUGCCUUAAA	2524
15717	UAUCAAAAUAAUGUGUUCA	874	15717	UAUCAAAAUAAUGUGUUCA	874	15735		2525
15735	AUGUCUGAGGCAAAAUGUU	875	15735	AUGUCUGAGGCAAAAUGUU	875	15753		2526
15/53	UGGACUGAGACCUUA	876	15753	UGGACUGAGACUGACCUUA	876	15771	UAAGGUCAGUCCA	2527

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2528	2529	2530	2531	2532	2533	2534	2535	2536	2537	2538	2539	2540	2541	2542	2543	2544	2545	2546	2547	2548	2549	2550	2551	2552	2553	2554	2555	2556	2557	2558	2559	2560	2561	2562	2563	2564	2565	2566	2567	2568
AUUCGUGAGGUCCUUUAGU	UUGUAUGCUGUGAGCAAAA	CUCCUUGUUUAACUAGCAU	GCAGGUACACGUAAUCAUC	├	AGCCUGCGCCUAAUAUUCU	CAAUAUCAUCGACAAACA	GUGUACCAUCUGUUUUGAC	-	CAUCAAUAGCCAGUGACAC	GUUUUGUAAGUGGGUAAGC	CAUACUCCUGAUUAGGAUG	ACAAGUGAAAGACAUCAGC	UUCUAAUGUAUUGUAAAUA		UGUCCAACAUGUGGCCAGU		GUGAGGUGUUAUCAUUAGU	ACUCAGGUUCCCAGUACCG	UGUACAUAGCCUCAUAAAA		CACAAGCACCUACAGCCUG	_	-	AUGGUCUCCUAAUACAGGC	\rightarrow	_			\rightarrow	\dashv	-	-4	_	-	AAACCUGACCAUUAGCACA	UGUUUUGUAUAAACCAAA	UGUCACUGCCUACACAUGU		AAUCACAUGUUGCUAUCGC	AAHCGCCAGCAHHAGHCCA
15789	15807	15825	15843	15861	15879	15897	15915	15933	15951	15969	15987	16005	16023	16041	16059	16077	16095	16113	16131	16149	16167	16185	16203	16221	16239	16257	16275	16293	16311	16329	16347	16365	16383	16401	16419	16437	16455	16473	16491	16509
877	878	879	880	881	882	883	884	882	988	887	888	889	068	891	892	893	894	895	968	897	868	899	900	901	902	903	904	905	906	206	808	606	910	911	912	913	914	915	916	917
ACUAAAGGACCUCACGAAU	UUUUGCUCACAGCAUACAA	AUGCUAGUUAAACAAGGAG	GAUGAUUACGUGUACCUGC	CCUUACCCAGAUCCAUCAA	AGAAUAUUAGGCGCAGGCU	UGUUUGUCGAUGAUAUUG	GUCAAAACAGAUGGUACAC	CUUAUGAUUGAAAGGUUCG	GUGUCACUGGCUAUUGAUG	GCUUACCCACUUACAAAAC	CAUCCUAAUCAGGAGUAUG	GCUGAUGUCUUUCACUUGU	UAUUUACAAUACAUUAGAA	AAGUUACAUGAUGAGCUUA	ACUGGCCACAUGUUGGACA	AUGUAUUCCGUAAUGCUAA	ACUAAUGAUAACACCUCAC	CGGUACUGGGAACCUGAGU	UNUNAUGAGGCUAUGUACA	ACACCACAUACAGUCUUGC	CAGGCUGUAGGUGCUUGUG	GUAUUGUGCAAUUCACAGA	ACUUCACUUCGUUGCGGUG	GCCUGUAUUAGGAGACCAU	UUCCUAUGUUGCAAGUGCU	UGCUAUGACCAUGUCAUUU	UCAACAUCACACAAAUUAG	GUGUUGUCUGUUAAUCCCU	UAUGUUUGCAAUGCCCCAG	GGUUGUGAUGUCACUGAUG	GUGACACAGUGUAUCUAG	GGAGGUAUGAGCUAUUAUU	UGCAAGUCACAUAAGCCUC	CCCAUUAGUUUUCCAUUAU	UGUGCUAAUGGUCAGGUUU	UUUGGUUUAUACAAAAACA	ACAUGUGUAGGCAGUGACA	AAUGUCACUGACUUCAAUG	GCGAUAGCAACAUGUGAUU	LIGGACI JAALIGCI IGGCGALILL
15771	15789	15807	15825	15843	15861	15879	15897	15915	15933	15951	15969	15987	16005	16023	16041	16059	16077	16095	16113	16131	16149	16167	16185	16203	16221	16239	16257	16275	16293	16311	16329	16347	16365	16383	16401	16419	16437	16455	16473	16491
877	878	879	880	881	882	883	884	882	886	887	888	688	890	891	892	893	894	895	968	897	868	833	900	901	305	903	904	905	906	200	806	606	910	911	912	913	914	915	916	917
ACUAAAGGACCUCACGAAU	UUUUGCUCACAGCAUACAA		_			\dashv	\dashv	CUUAUGAUUGAAAGGUUCG	-	GCUUACCCACUUACA		\rightarrow	-	AAGUUACAUGAUGAGCUUA	ACUGGCCACAUGUUGGACA	AUGUAUUCCGUAAUGCUAA	ACUAAUGAUAACACCUCAC	CGGUACUGGGAACCUGAGU	UUUUAUGAGGCUAUGUACA	ACACCACAUACAGUCUUGC	CAGGCUGUAGGUGCUUGUG	GUAUUGUGCAAUUCACAGA	ACUUCACUUCGUUGCGGUG	GCCUGUAUUAGGAGACCAU	UUCCUAUGUUGCAAGUGCU	UGCUAUGACCAUGUCAUUU	UCAACAUCACACAAAUUAG	GUGUUGUCUGUUAAUCCCU	UAUGUUUGCAAUGCCCCAG	GGUUGUGAUGUCACUGAUG	GUGACACACUGUAUCUAG	GGAGGUAUGAGCUAUUAUU	UGCAAGUCACAUAAGCCUC	CCCAUUAGUUUUCCAUUAU	UGUGCUAAUGGUCAGGUUU	UUUGGUUUAUACAAAACA	ACAUGUGUAGGCAGUGACA	AAUGUCACUGACUUCAAUG	GCGAUAGCAACAUGUGAUU	UGGACUAAUGCUGGCGAUU
15771	15789	15807	15825	15843	15861	15879	15897	15915	15933	15951	15969	15987	16005	16023	16041	16059	16077	16095	16113	16131	16149	16167	16185	16203	16221	16239	16257	16275	16293	16311	16329	16347	16365	16383	16401	16419	16437	16455	16473	16491

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2570	2571	2572	2573	2574	2575	2576	2577	2578	2579	2580	2581	2582	2583	2584	2585	2586	2587	2588	2589	2590	2591	2592	2593	2594	2595	2596	2597	2598	2599	2600	2601	2602	2603	2604	2605	2606	2607	2608	2609	2610	2611
\vdash	GCGUUUCUGCUGCGAAAAG	-	\dashv	CGCGUACAGUGGCAAUACC	CUCUGUCAGAGAGUACUUC	CCCAUGAAAGAUGCAAUUC	GUCUAGGUUUUCCAACCUC	AGUUUCUGUUCAAUGGUGG	GGUAACCAGUAAAGACAUA	UACUAUUUUAGUUACACG	ACUCUCCAAUCUGUACUUU	CACCUUUUUCAAAGGUGUA	CAACAGCAUCACCAUAGUC	UCGUAGUACCUCUGUACAC	CAACAUUCAACUUGUAUGU	UCAACACAAAGUAAUCACC	GCAUUACAGUGUGAGAUGU	GAGUAGGUGCACUAAGUGG	AGUGCUCUUGUGGCACUAG	AGCCAGUAAUUCUCACAUA	UGUUGAGUGUUGGGUACAA	UAGAAACUCAUCUGAGAU	GAUAAUUUGCAACAUUGCU	UNUGCAUGCCGACCUUUUG	CUUGGAGUGUAGAGUACUU	UACCAGUACCAGGUGGUCC	CGAUGGCAAAAUGACUCUU	GGUAAUAGAGAGCAAGUCC	ACACUAUGCGAGCAGAUGG	CAUGAGAGCAUGCCGUAUA	AUAGGGCAUCAACAGCUGC	AUUUUAAUGCCUUUUCACA	AUUNAUCUAUGGGCAAAUA	CAGGUAUGAUUCUACUACA	ACUCUACGCGCGCACGCGC	CUUUGAAUUUAUCAAAACA	GUUCUAGUGUUGAAUUCAC	CAGUGCAGAAAACAUACUG	UNUCUGGCAAUGCAUUUAC	CUACAAUGUCAGCAGUUGU	UAGAGAUUUCAUCAAAGAC
16545	16563	16581	16599	16617	16635	16653	16671	16689	16707	16725	16743	16761	16779	16797	16815	16833	16851	16869	16887	16905	16923	16941	16959	16977	16995	17013	17031	17049	17067	17085	17103	17121	17139	17157	17175	17193	17211	17229	17247	17265	17283
919	920	921	922	923	924	925	926	927	928	929	930	931	932	933	934	935	936	937	938	939	940	941	942	943	944	945	946	947	948	949	950	951	952	953	954	955	926	957	958	959	096
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16527	16545	16563	16581	16599	16617	16635	16653	16671	16689	16707	16725	16743	16/61	16779	16797	16815	16833	16851	16869	16887	16905	16923	16941	16959	16977	16995	17013	17031	17067	17005	17083	1 103	1/1/21	17139	17157	17175	17193	17211	17229	17247	17265
919	920	126	375	923	924	925	926	927	928	929	930	931	932	933	934	_ .	_	_	_						944	945	946	947	948	949	200	200	706	202	장 [955	926	957	328	959	960
16527 UGUACUGAGAGACUCAAGC	16563 CHOAAACCAGAAACGC	+	+	+-	16625 CAALILIOON IN 19711000	18653 GAAUUGCAUCUUUCAUGGG	18653 GAGGUUGGAAAACCUAGAC	CCACCAUDGAACAGAA	+	CGUGUAACUAAAAUA	+	16743 UACACCUUUGAAAAGGUG	16/770 CHOUNGEOGRADECUGUUG	GUGUACAGAGGUACUA	ACAUACAAGUUGAAUG	16823 ACALICITOTOTO	18854 PCAUCUCACACUGUAAUGC	16831 CCACUDAGUGCACCUACUC	16869 CUAGUGCCACAGAGCACU	1688/ UAUGUGAGAAUUACUGGCU	16905 UUGUACCCAACACACA	16923 AUCUCAGAUGAGUUUUCUA	16941 AGCAAUGUUGCAAAUUAUC		1697 AAGUACUCUACACUCCAAG	-	-	17049 CCALICLICCALIACION	_	+	—	$\overline{}$	17130 HOLIVOLIACIANTIANOLIA		+	-			4CAAACHCCAUUGCCAGA	1728F CHOUNDAY	1/203 GUCUUUGAUGAAAUCUCUA

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\vdash	UAGCAUGACACACUCAA	+	-	UAGUCAGCAAUGUGCGGGG	CUGGUUCUAGUGUGCCUUU	\vdash	UUGUUUCANAAGUCUGCA	GGAACAUGUCUGGACCUAU		CAACAAUUUCAGCAGGACA	CUAAAGCACUCACAGUGUC	UUAGCUUAUUGUCAUAAAC	ACUUAUCCUUGUGUGCUUU	_	UAACACCUUUGUAGAACAU	AUGAAACAUCAUGUGUAAU		CUCUUACAACGCCUAUUUG	GAUUGCGUGUAAGAAAUUC	CAGCUUUUCUCCAAGCAGG	UAUAAGGUGAGAUAAAAAC	CUACAGCGUUCUGUGAAUU	AUCCUAAGAUUUUUGAAGC	_	-		_	-	_	-	-	_		GUGGUAUUUCUAGACUUGU	AUGUAGCCACAUUGCGACG	UNACAUUUUCUGCUUGUAA	AGUCCUUAAAAAGUCCAGU			100 100 VI 101 101 100 V 101 1000
17301	17319	17355	17373	17391	17409	17427	17445	17463	17481	17499	17517	17535	17553	17571	17589	17607	17625	17643	17661	17679	17697	17715	17733	17751	17769	17787	17805	17823	17841	17859	17877	17895	17913	17931	17949	17967	17985	18003	18021	12020
961	395 296 3	964	965	996	296	896	696	970	971	972	973	974	975	926	977	828	979	086	981	982	983	984	985	986	987	988	686	066	991	992	993	994	995	966	997	968	666	1000	1001	1002
AUGGCUACUAAUUAUGACU	UUGAGUGUUGUCAAUGCUA	UACGUCUAUAUUGGCGAUC	CCUGCUCAAUUACCAGCCC	CCCCGCACAUUGCUGACUA	AAAGGCACACUAGAACCAG	GAAUAUUUUAAUUCAGUGU	UGCAGACUUAUGAAAACAA	AUAGGUCCAGACAUGUUCC	CUUGGAACUUGUCGCCGUU	nencchecheavannenne	GACACUGUGAGUGCUUUAG	GUUUAUGACAAUAAGCUAA	AAAGCACACAAGGAUAAGU	UCAGCUCAAUGCUUCAAAA	AUGUUCUACAAAGGUGUUA	AUUACACAUGAUGUUCAU	UCUGCAAUCAACAGACCUC	CAAAUAGGCGUUGUAAGAG	GAAUUCUUACACGCAAUC	CCUGCUUGGAGAAAGCUG	GUUUUAUCUCACCUUAUA	AAUUCACAGAACGCUGUAG	GCUUCAAAAAUCUUAGGAU	UUGCCUACGCAGACUGUUG	GAUUCAUCACAGGGUUCUG	GAAUAUGACUAUGUCAUAU	UUCACACAAACUACUGAAA	ACAGCACACUCUUGUAAUG	GUCAACCGCUUCAAUGUGG	GCUAUCACAAGGGCAAAAA	AUUGGCAUUUUGUGCAUAA	AUGUCUGAUAGAGAUCUUU	UAUGACAAACUGCAAUUUA	ACAAGUCUAGAAAUACCAC	CGUCGCAAUGUGGCUACAU	UUACAAGCAGAAAAUGUAA	ACUGGACUUUUNAAGGACU	UGUAGUAAGAUCAUUACUG	GGUCUUCAUCCUACACAGG	SOUND ACADA LOCATION ACTOR ACT
17283	17301	17337	17355	17373	17391	17409	17427	17445	17463	17481	17499	17517	17535	17553	17571	17589	17607	17625	17643	17661	17679	17697	17715	17733	17751	17769	17787	17805	17823	17841	17859	17877	17895	17913	17931	17949	17967	17985	18003	18031
961	962	964	965	996	296	896	696	920	971	972	973	974	975	926	226	826	979	980	981	982	983	984	985	986	987	988	686	066	991	392	993	994	995	966	266	866	666	1000	1001	100
AUGGCUACUAAUUAU	AGACIIII CGI IGCAAAACACII	UACGUCUAUAUUGGC	-	H	_	GAAUAUUUUAAUUCA	-	-	\dashv	-	GACACUGUGAGUGCUUUAG		-	\dashv	I AUGUUCUACAAAGGUGUUA	-		CAAAUAGGCGUUGUA	-	CCUGCUUGGAGAAAA	GUUUUUAUCUCACCU	\rightarrow	-	UNGCCUACGCAGACUGUUG	-	GAAUAUGACUAUGUCAUAU	UUCACACAAACUACUC		-	-	-	\dashv		ACAAGUCUAGAAAUAC	-	_	_	UGUAGUAAGAUCAUU/		GCACCHACACACCHCAGCG
17283	17319	17337	17355	17373	17391	17409	17427	17445	17463	17481	17499	17517	17535	17553	17571	17589	17607	17625	17643	17661	17679	17697	17715	17733	17751	17769	17787	17805	17823	17841	17859	17877	17895	17913	17931	17949	17967	17985	18003	18021

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\vdash	\dashv			UUCUGCAAGCAGAAUUAAC	CAACCAUGUGUUGUACUUU	CAAGCAAUGCAGACUUCAC	GAACUGGAAACUUAUCAGC	GAUUUCCAAUGUCAUGAAG	CACACUUGAUAGCCUUUGG	CUACUUCAGCCUGAGGCAC		UGUCACUACAUGGCUGAGC	CCUCUAUUUGUAAGCUUU	_	AUUUAUCGUGAUGUGUAGC	AACAAACACCAUCAGUGAA	CGUUACAAUUCCAAAACAA	UGGCUGGGUAACGAUCAAC	ACCUACACACAUUGCAUU	ACAAGACUCUUGUGUCAAA	CUGGUAAGUUCAAGUUUGA	AACUACCACCAUCACAGCC	Ш	AAGCUGGAGUGUGGAAUGC				-		GAACAUAAUCAAUAUCCGA	ACGUAGCAGAUUUGAGUGG	AAUUGCAUCGUGUAAUACA	UGCAAACAGCACCACCUAA	ACUCAUUUGCAUGGUGUCU	CAUCCAAGUACUGUCGGUA	AAAUCAUCAUAUAUAUGC		CAAAUUGUUUGUAAAUCCA	UCCACAGGUUAUAAGUAUC	GUAACCUGGUAAAUGUAUU	CCACAUUUUCUAAACUCUG
18813	18831	18849	18867	18885	18903	18921	18939	18957	18975	18993	19011	19029	19047	19065	19083	19101	19119	19137	19155	19173	19191	19209	19227	19245	19263	19281	19299	19317	19335	19353	19371	19389	19407	19425	19443	19461	19479	19497	19515	19533	19551
1045	1046	1047	1048	1049	1050	1021	1052	1053	1054	1055	1056	1057	1058	1059	1060	1061	1062	1063	1064	1065	1066	1067	1068	1069	1070	1071	1072	1073	1074	1075	1076	1077	1078	1079	1080	1081	1082	1083	1084	1085	1086
GCAGUCCAUGAGUGCUUUG	GUUAAGCGCGUUGAUUGGU	UCUGUUGAAUACCCUAUUA	AUAGGAGAUGAACUGAGGG	GUUAAUUCUGCUUGCAGAA	AAAGUACAACACAUGGUUG	GUGAAGUCUGCAUUGCUUG	GCUGAUAAGUUUCCAGUUC	CUUCAUGACAUUGGAAAUC	CCAAAGGCUAUCAAGUGUG	GUGCCUCAGGCUGAAGUAG	GAAUGGAAGUUCUACGAUG	GCUCAGCCAUGUAGUGACA	AAAGCUUACAAAAUAGAGG	GAACUCUUCUAUUCUUAUG	GCUACACAUCACGAUAAAU	UUCACUGAUGGUGUUGUU	UUGUUUUGGAAUUGUAACG	GUUGAUCGUUACCCAGCCA	AAUGCAAUUGUGUGUAGGU	UUUGACACAAGAGUCUUGU	UCAAACUUGAACUUACCAG	GGCUGUGAUGGUGGUAGUU	UUGUAUGUGAAUAAGCAUG	GCAUUCCACACUCCAGCUU	UUCGAUAAAAGUGCAUUUA	ACUAAUUUAAAGCAAUUGC	CCUUUCUUUACUAUUCUG	GAUAGUCCUUGUGAGUCUC	CAUGGCAAACAAGUAGUGU	UCGGAUAUUGAUUAUGUUC	CCACUCAAAUCUGCUACGU	UGUAUUACACGAUGCAAUU	UUAGGUGGUGCUGUUUGCA	AGACACCAUGCAAAUGAGU	UACCGACAGUACUUGGAUG	GCAUAUAAUAUGAUGAUUU	UCUGCUGGAUUUAGCCUAU	UGGAUUUACAAACAAUUUG	GAUACUUAUAACCUGUGGA	AAUACAUUUACCAGGUUAC	CAGAGUUUAGAAAAUGUGG
18795	18813	18831	18849	18867	18885	18903	18921	18939	18957	18975	18993	19011	19029	19047	19065	19083	19101	19119	19137	19155	19173	19191	19209	19227	19245	19263	19281	19299	19317	19335	19353	19371	19389	19407	19425	19443	19461	19479	19497	19515	19533
1045	1046	1047	1048	1049	1050	1051	1052	1053	1054	1055	1056	1057	1058	1059	1060	1061	1062	1063	1064	1065	1066	1067	1068	1069	1070	1071	1072	1073	1074	1075	1076	1077	1078	1079	1080	1081	1082	1083	1084	1085	1086
GCAGUCCAUGAGUGC	GUUAAGCGCGUUGAU	-+		\dashv	-		-	-	7 CCAAAGGCUAUCAAGUGUG	-		ccucaeccauguagu	AAAGCUUACAAAAUAC	\dashv	-	UNCACUGAUGGUGUU	UUGUUUUGGAAUUGU		7 AAUGCAAUUGUGUGUAGGU	\vdash	UCAAACUUGAACUUAC	\dashv	UNGUAUGUGAAUAAG	GCAUUCCACACUCCA	UUCGAUAAAAGUGCAI				\rightarrow	5 UCGGAUAUUGAUUAUGUUC	-+	UGUAUUACACGAUGC,	\dashv	-		GCAUAUAAUAUGAUGA	UCUGCUGGAUUUAGC	UGGAUUUACAAACAAI	\blacksquare	5 AAUACAUUUACCAGGUUAC	3 CAGAGUUUAGAAAAUGUGG
18795	18813	18831	18849	18867	18885	18903	18921	18939	18957	18975	18993	19011	19029	19047	19065	19083	19101	19119	19137	19155	19173	19191	19209	19227	19245	19263	19281	19299	19317	19335	19353	19371	19389	19407	19425	19443	19461	19479	19497	19515	19533

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UCACGAGAUUCACCACUUA 1129 20307 UCACAAGAUUCACCACUUA 1139 20315 DAAUUUGAGCACUUAUCCUCCAUNGAGCAUUUAUCC 1130 20315 AAAUUUAGCAGCAUUCAUACCA 1131 20315 AAAUUUAGCAGCAUUCAUACCA 1131 20315 AAAUUUAGCAGCAUUCAUACCA 1131 20315 AAAUUUCAUACACAGCACACACACACACACACACACACAC	0276	2781	2782	2783	2784	2785	2786	2787	2788	2789	2790	2791	2792	2793	2794	2795	2796	2797	2798	2799	2800	2801	2802	2803	2804	2805	2806	2807	2808	2809	2810	2811	2812	2813	2814	2815	2816	2817	2818	2819	2820
UCACAGAGAUUCACCACUUA 1129 20397 UCACAGAGAUUCACCACUUA 1129 20355 AAAUUAGAGGAUUUAUCCC 1130 20325 AAAUUAGAGGAUUUUAUCCC 1130 20343 AAAAAUUAGUCAAAGGAUUCAUCAGAGAGAGAGAGAGAGA	\vdash	╁	H	_	 	_	-			\vdash				_								-							Н				7					-			
UCACCAGAUUCACCACUUA 1129 20307 UCACCAGAUUCACCACUUA AAAUUAGAGGAUUUUAUCC 1130 20325 AAAUUAGAGGAUUUUAUCC CCUAUGGACAGCAGUGA 1131 20335 CCUAUGGACGCACAGUGA AAAAUUACUUCAULAACAG 1132 20336 CCUAUGGACACACAGUUCA GAUGCGCAAACAGGUUCAU 1133 20339 CCUAUGGACACAGGUUCAU GAUGCGCAAACAGGUUCAU 1134 20337 LOCAAAUUUCAUUCAUCAGGUUCAU GAUGCGCAAACAGGUUCAU 1134 20337 LOCAAAGUUCAUCAAGGUUCAU GAUGCGCAAACUUCA 1138 20435 GAUGCGCAAACAGGUUCAU GAUGCGCAAACAGUUCA 1138 20437 GAUGCGCAAAGGUUCAU JUACCUUUGAUCAUUUACAUUCAU 1138 20437 GAUGCGCAAAGUUCU JUACCAAACUUCAAAGUUCA 1142 20536 JUACAGCUUCAAAGUUCA JUACCCAAAACUUCAAACUUCA 1142 20537 JUACAGCAUCAAACUUCA JUACCCAAAACUUCAAACUUCA 1144 20537 JUACAGCAUCAAACUUCA JUACCUUCAAAUUCACUUCAAAGUUCA 1148 20540 JUCACAAACUUCAAACUUCA JUCGUUUCAGAAUUACCAAACUUCA 1148 20540	\vdash	+	\vdash	-	\vdash	<u> </u>	├		-	├─	<u> </u>					-				Н	\vdash	_				ш	-		-			-	\dashv	\dashv	_		_		⊢	<u> </u>	_
UCACAGGUUCACCACUUA 1129 20307 AAAUUAGAGGAUUUAAUCC 1130 20325 CCUAUUGAGGAUUUUAACAG 1131 20343 AAAAAAUUACUCAUAACAG 1132 20379 GAUGCGCAACAGGUUCAU 1135 20415 GAUGCGCAACAGGUUCAU 1135 20415 UCAAAAUUACUUUUUACUUG 1134 20487 GUCAUUGAUCAAAGUUG 1135 20487 GUCAUUUGACAAUUGACU 1139 20487 UCAGUGAUUUCAUUCA 1140 20505 AUAAAGUCACAAGAUUCA 1141 20523 GCACUUGAAAUUUCAUUCA 1142 20541 UCAGUGUUGAAACUUCA 1143 20559 AUUCCAAACUUCAAAGCAA 1144 20577 AUCCAAACUUCAAAGCAA 1143 20550 AUCCAAACUUCAAAGAAGAAGAAGAAGAAGAAGAAGAAGAAGAAGAAGAA	11.20	1130	1131	1132	1133	1134	1135	1136	1137	1138	1139	1140	1141	1142	1143	1144	1145	1146	1147	1148	1149	1150	1151	1152	1153	1154	1155	1156	1157	1158	1159	1160	1161	1162	1163	1164	1165	1166	1167	1168	4400
UCACAAGAUUCACCACUUA AAAAUUAGAGAUUCACCACUUA AAAAUUAGAGAUUCACAGAUAACAG AAAAAUUGAUGAUCAUAAAG GAGAGUUUCAAAAGUUCA ITTT AUGAGUUUCAAAAGUUCA ITTT AUGCUUGAAAUUUCAAAAGUUCA ITTT AUGCUUGAAAUUUCAAAAGUUCA ITTT AUGCUUGAAACUUUCAAAAGUUCA ITTT AUGCUUCGAAAUUUCAAAAGUUCA ITTT AUGCUUCGAAAUUUCAAAAGUUCA ITTT AUGCUUCGAAAUUUCAAAAGUUCA ITTT AUGCUUCGAAAUUUCAAAAGUUCA ITTT AUGCUUUGAAAACUUCA ITTT AUGCUUCAAAACUUCAAAGAA ITTT AUGCUUCAAAACUUCAAAGAA ITTT AACCAAAACCAAACUUCAAACAA ITTT AACCAAAACCAAACAACAAACAAACAAACAAAAAAAA	VIII.000000111	AAAUUAGAGGAUUUUAUCC	CCUAUGGACAGCACAGUGA	AAAAAUUACUUCAUAACAG	GAUGCGCAAACAGGUUCAU	ucaaaauguguguucug	GUGAUUGAUCUUUNACUUG	GAUGACUUUGUCGAGAUAA	AUAAAGUCACAAGAUUUGU	UCAGUGAUUUCAAAAGUGG	GUCAAGGUUACAAUUGACU	UAUGCUGAAAUUUCAUUCA	AUGCUUUGGUGUAAGGAUG	GGACAUGUUGAAACCUUCU	UACCCAAAACUACAAGCAA	AGUCGAGCGUGGCAACCAG	GGUGUUGCGAUGCCUAACU	UUGUACAAGAUGCAAAGAA	AUGCUUCUUGAAAAGUGUG	GACCUUCAGAAUUAUGGUG	GAAAAUGCUGUUAUACCAA	AAAGGAAUAAUGAUGAAUG	GUCGCAAAGUAUACUCAAC	cugugucaanacunaaana	ACACUUACUUUAGCUGUAC	CCCUACAACAUGAGAGUUA	AUUCACUUUGGUGCUGGCU	UCUGAUAAAGGAGUUGCAC	CCAGGUACAGCUGUGCUCA	AGACAAUGGUUGCCAACUG	GGCACACUACUUGUCGAUU	UCAGAUCUUAAUGACUUCG	GUCUCCGACGCAUAUUCUA	ACUUNAAUUGGAGACUGUG	GCAACAGUACAUACGGCUA	AAUAAAUGGGACCUUAUUA	AUUAGCGAUAUGUAUGACC	ccuaggaccaaacauguga	ACAAAAGAGAAUGACUCUA	AAAGAAGGGUUUUUCACUU	V V I V I I I I V C C I C I C I C I V I V
UCACAAGAUUCACCACUUA AAAUUUGACCACACACACACACACACACACACACACACAC	20207	20325	20343	20361	20379	20397	20415	20433	20451	20469	20487	20505	20523	20541	20559	20577	20595	20613	20631	20649	20667	20685	20703	20721	20739	20757	20775	20793	20811	20829	20847	20865	20883	20901	20919	20937	20955	20973	20991	21009	04007
UCACAGGAUUCACCA AAAAUUAGGAUUUUU CCUAUGGACAGCACA AAAAAUUGGACAGGGU UCAAAAUUGGUGAAAA GAAAAACAAAAAAAAAA	1120	1130	1131	1132	1133	1134	1135	1136	1137	1138	1139	1140	1141	1142	1143	1144	1145	1146	1147	1148	1149	1150	1151	1152	1153	1154	1155	1156	1157	1158	1159	1160	1161	1162	1163	1164	1165	1166	1167	1168	4460
ᄀᄋᅝᅁᅘᄬᄯᅜᇎᇎᅜᅜᅜᅜᅜᅜᅜᅜᅜᅜᅼᇏᅜᅼᇏᇎᅜᅼᇎᇎᅜᅜᆄᇏᅜᅼᇎᅜᅼᇄᇋᇏᇬᇎᄺᆄᄧᅹᄍᇎᆔᆔᇋᆑᆔᆔᇎᇎᆄᆄ	20302 1110400400111104000	AAAUUAGAGGAUUUL	-	AAAAAUUACUUCAUA	Ь—	20397 UCAAAAUGUGUGUGUUCUG	20415 GUGAUUGAUCUUUNACUUG	\vdash	_	UCAGUGAUUUCAAAA	GUCAAGGUUACAAUL						\vdash	_	AUGCUUCUUGAAAAG	GACCUUCAGAAUUAU	-	-	GUCGCAAAGUAUACI	CUGUGUCAAUACUUA	ACACUUACUUAGCU	CCCUACAACAUGAGA	AUUCACUUUGGUGCI	UCUGAUAAAGGAGUL	-	\dashv	GGCACACUACUUGUC	UCAGAUCUUAAUGAC	\dashv		GCAACAGUACAUACG	AAUAAAUGGGACCUU	AUUAGCGAUAUGUAU	ccuaggaccaaacau	20991 ACAAAAGAGAAUGACUCUA	-	21027 114/10/16/16/16/16/11/11/11/14

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2822	2823	4202	2822	2826	2827	2828	2829	2830	2831	2832	2833	2834	2835	2836	2837	2838	2839	2840	2841	2842	2843	2844	2845	2846	2847	2848	2849	2850	2851	2852	2853	2854	2855	2856	2857	2858	2859	2860	2861	2862	2863
\vdash	-		4	\dashv	AUGCAUUUACAUUUGUAAC	AAAAUGCUUCCGAUGAUGA	\vdash	GUUCCUUCGCUUGCCAAG	UGGUAUAGCCAUCAAUUUG	AAAUGUAGUUAGCAUGCAU	GAUUUGUGUUCCUCCAGAA	AGGAAGACAACUGGAUAGG	UCAUGUCAAAGAGÜGAAUA	AUUUAAGAGGAAAUUUGCU	UNACAGCAGUUCCUCUUAA	-	AAAUCAUAUCAUUGAUUUG	CUUUUUCCAGAAGAGAAUA	CUCUAAUGAUAAGCCUACC	CCACAACUCUGUUGUUUUC	CAAGAAUAUCACUUGAAAC	nenncennnvennennvvc	_	CACUAGUGAGAGUAAGAAA	ACCGGUCAGGUCACUACC	CAUCAUCAAAAGUGGUGCA			\dashv	\dashv	-		UAACAUUAGAAUAAAAUGG	UAAUAGUAUGAAACCCUGU	GGUUGCCAAACGUAUGAUU	CCUUAAAAGGUAUGACAGG	CAGCAAAUAAAUACCAUC	cauuugauuucucuguggc	AAACCCAACCACGGACAAC		UCACCGACUGUGACUUGUU
21081	21099	71117	21135	21153	21171	21189	21207	21225	21243	21261	21279	21297	21315	21333	21351	21369	21387	21405	21423	21441	21459	21477	21495	21513	21531	21549	21567	21585	21603	21621	21639	21657	21675	21693	21711	21729	21747	21765	21783	21801	21819
1171	11/2	5/17	11/4	1175	1176	1177	1178	1179	1180	1181	1182	1183	1184	1185	1186	1187	1188	1189	1190	1191	1192	1193	1194	1195	1196	1197	1198	1199	1200	1201	1202	1203	1204	1205	1206	1207	1208	1209	1210	1211	1212
-			-	-	GUUACAAAUGUAAAUGCAU	UCAUCAUCGGAAGCAUUUU	_	CUUGGCAAGCCGAAGGAAC	CAAAUUGAUGGCUAUACCA	AUGCAUGCUAACUACAUUU	UUCUGGAGGAACACAAAUC	ccuauccaeuueucuuccu	UAUUCACUCUUUGACAUGA	AGCAAAUUUCCUCUUAAAU	UUAAGAGGAACUGCUGUAA		CAAAUCAAUGAUAUGAUUU	UAUUCUCUUGGAAAAAG	GGUAGGCUUAUCAUUAGAG	GAAAACAACAGAGUUGUGG	GUUUCAAGUGAUAUUCUUG	GUUAACAACUAAACGAACA	AUGUUUAUUUUCUUAUUAU	UUUCUUACUCUCACUAGUG	GGUAGUGACCUUGACCGGU	UGCACCACUUUUGAUGAUG	GUUCAAGCUCCUAAUUACA		_	CCUGAUGAAAUUUUAGAU	UCAGACACUCUUUAUUUAA	ACUCAGGAUUUAUUUCUUC	CCAUUUUAUUCUAAUGUUA	_	AAUCAUACGUUUGGCAACC		GAUGGUAUUNAUUUGCUG	GCCACAGAGAAAUCAAAUG	GUUGUCCGUGGUUGGGUUU	UUUGGUUCUACCAUGAACA	AACAAGUCACAGUCGGUGA
21063	21081	21033	71112	21135	21153	21171	21189	21207	21225	21243	21261	21279	21297	21315	21333	21351	21369	21387	21405	21423	21441	21459	21477	21495	21513	21531	21549	21567	21585	21603	21621	21639	21657	21675	21693	21711	21729	21747	21765	21783	21801
1171	11/2	2/1	11/4	1175	1176	1177	1178	1179	1180	1181	1182	1183	1184	1185	1186	1187	1188	1189	1190	1191	1192	1193	1194	1195	1196	1197	1198	1199	1200	1201	1202	1203	1204	1205	1206	1207	1208	1209	1210	1211	1212
GGUGGUUCUAUAGCU	HOGANICO ICACOLLINACA	**************************************	AAGCUUAUGGGCCAU	UCAUGGUGGACAGCU	\dashv	UCAUCGGAAGCAUUUU		cuuggcaagccgaagc	CAAAUUGAUGGCUAUACCA	AUGCAUGCUAACUACA		CCUAUCCAGUUGUCUL	-	\rightarrow		AUGUCUCUUAAGGAGA	CAAAUCAAUGAUAUGAUUU	UAUUCUCUUCUGGAAAAG	GGUAGGCUUAUCAUUA	-	GUUUCAAGUGAUAUUCUUG	_	\dashv			UGCACCACUUUUGAUG	-	_	AUGAGGGGGUUUACUAUC	-	UCAGACACUCUUUAUU		CCAUUUUAUUCUAAUG	_	Н			GCCACAGAGAAAUCAA		UUUGGUUCUACCAUGA	AACAAGUCACAGUCGGUGA
21063	21087	211033	71117	21135	21153	21171	21189	21207	21225	21243	21261	21279	21297	21315	21333	21351	21369	21387	21405	21423	21441	21459	21477	21495	21513	21531	21549	21567	21585	21603	21621	21639	21657	21675	21693	21711	21729	21747	21765	21783	21801

CUCHANO 1214 2.1833 ACUADAMOGNUO UNAMAGNUO UT 1215 2.1813 ACAANUCAAAGUUACAUGUUA UUUG 1216 2.1883 GACAGUUCAAAGUUACAUGUUA 1216 2.1881 CAAGAAAGGGUUGACAUGUUACAUGUUA UUUG 1216 2.1887 LOGAUUUAAAGGGUUGACAGAGAGUUA 1218 2.1881 CAAGAAGUUAAAGGGUUGACAGAGAGUUACAGAGAGAGUUAAAGUAGAGAGAG	AUGAUGAUGACAAUUCUA
21873 UGUGACAACCCUUUCUUUG 1216 21891 21891 GCUGUUUCUAAACCCAUGG 1217 21909 21909 GGUACACAGACACAUGCA 1217 21909 21909 GGUACACAGACACAUACUA 1218 21927 21927 21909 21909 21909 21981 UACAUAUCUGAUGUU 1221 21981 21983 UACAUAUCUGAUGUU 1221 22017 22017 CACUUACGAGAGUUUGUGU 1224 22035 22017 CACUUACGAGAGUUUGUGU 1224 22035 22017 CACUUACGAGAGUUUGUGU 1226 22017 22017 CACUUACGAGAGUUUUAAGC 1228 2217 22018 UUUAAAAUAAAAUAAAGACCUUU 1228 2217 22019 UUUCGCUUUUAAGCUUC 1228 2217 22017 UCUGGUUUUUAAGCUUU 1231 2217 2218 GCUCAAGACAUUUCACCUU 1232 2217 2218 ACCUCAUCAGAUCAUUCACCUU 1234 2221 2219 CCUCUUGGUAUUUCACCUC 1236 </td <td>1215</td>	1215
GCUGUUUCUAAACCCAUGG 1217 21909 GCUGUUUCUAAACCCAUGG 1217 21927 AUGAUAUUCGACAUACUA 1218 21945 UUUUAAUUCGACUUUU 1220 21981 UUUUAAUUCGACUUUU 1221 21981 UACAUAUUCGAGU 1222 21999 AAGUCAGGUAAUUUUAAAC 1222 22017 UCCUUGAUGUUUAAAC 1222 22017 CACUUACGAGAGUUUAUAAC 1222 22017 CACUUACGAGAGUUUAUAAC 1225 22017 CACUUACGAGAUUUGAGC 1226 22017 CACUUACGAGCCUUUGA 1221 22083 GCUAUCGAGCCUUUGAAC 1232 22179 CCUCUUGGAUUUAACCUU 1232 22173 CCUCUUGGAUUUUAACCUU 1236 22213 ACGUCAGCUUUAACUUU 1236 2221 CCUCUUGCCUCAGUUU 1236 2237 CCUCUUGCCUCAAGUCCU 1240 2236 AACACUCACACUUUCAAUCCAC 1242 2236 CUUCCUCAAAUCCUCU 1243 22413 CUUCCUCAAACCUUUCAAUCACCUUUCAAUCA	1216
GGUACACAGACAUACUA 1218 21927 AUGAUAUUCGAUAUGCAGU 1219 21945 AUGAUAUUCGAGU 1220 21981 UUUUAAUUCGAGU 1220 21981 UUUAAUUCGAGAA 1222 21981 UCCCUUGAUGCCUUUU 1222 21989 AAGUCAGGUAAUUUAAAC 1222 22017 CACUUACGAGAGUUUGUA 1224 22053 UUUAAAAAUAAAGAUGGU 1225 22071 GCCUAUCGAGCUUUGAACCUU 1228 22107 UUUCGUAUCAACCUUUGA 1230 2215 AAGUCAGCUUUUAACCUU 1232 22179 CCUCUUGGUUUUAACCUU 1232 22170 UCUGGUUUUAACCUUU 1235 22215 ACCUCAGCCUUUUCACCUU 1235 22215 ACCUCAGCUCAUUUCACCUU 1236 22281 ACCUCAGCUCAUUUAAAGC 1236 22281 ACCUCAGCUCAUUUAAAGC 1236 22281 ACAAUCACACAGAUCCUCA 1236 22281 ACAAUCACACAGAUCCUCA 1240 2341 UCUGCUCAAAGGAAUCCUCA	1217
AUGAUAUUCGAUAAUGCAU 1219 21945 UUUUAAUUCGAGU 1220 21981 UUUUAAUUCGAGU 1220 21981 UUUAAUUGCACUUUUCGAGU 1221 21981 UACAUAUUGAAGA 1222 21999 AAGUCAGGUAAUUUUAAAC 1223 22017 UUUAAAAAUAAAGAUGGU 1224 22053 UUUUAAAAAUAAGAUGGU 1225 22071 GCCUUACGAGCUUUAAGAUG 1228 22107 UUUAAAAAUAAGACUUUGA 1223 22107 UUUCUUUAAGAUUCACCUU 1232 2215 GCUCAAGACCUUUCACCUU 1232 2215 AACAAUUUUAAGACCUUUGA 1232 22179 CCUCUUGGUUUUAACCUU 1235 22215 ACAAAUUUUAAGCCUUUU 1235 22215 ACCUCAGCCUUUUAAGCCU 1236 22217 CCUCUUGGCUAUUUAAGCCUUU 1236 22287 ACCACUACACUUUAAGCCUUU 1241 22331 CCUCUUGCCUCAGAAUCCUC 1241 22341 UCUGCUCAAACUCUCAGAGACCUUUGAGAGACCUUUGAGAGAGA	1218
UUUDAUUGCACUUUCGAGU 1220 21963 UUUDAUUGCACUUUCGAGU 1221 21981 UACAUAUCUGAUGCCUUUU 1221 21981 UCGCUUGAUGUUUCAGAAA 1222 21999 AAGUCAGGUAAUUUAAGC 1224 22035 UUUDAAAAUUAAGAUUGGGU 1224 22057 GCUAUCCAACCUAUUGA 1228 22717 UUUCCUCAUUUAACAUUC 1232 22107 UCUGGUAUUUAACAUUC 1230 2215 ACCUCAGCACAUUC 1231 22215 ACCUCAGCACAUUC 1236 2216 ACCUCAGCACAUUUCACCUG 1236 2216 ACCUCAGCACAUUUCACCUG 1236 22215 ACCUCAGCACAUUUCACCUG 1236 22215 ACCUCAGCACAUUUCACCUG 1236 22216 ACCUCAGCACAUUCCUCAACUCUC 1236 22216 ACCUCAGCACAUUUCACCUCU 1236 22217 ACCUCAGCACAUUUCACCUCU 1240 2231 ACCUCAGCAUCACUCUCACAAUCCUC 1240 2231 ACCUCACACACACAAUCCUC 1241 222413 A	1219
UCGCUUGAUGAUGAGAA AGCUAGGUAGAUGUUUAAAC CACUAGGGUAAUUUAAGG GGCUAUCCUUUUAAGAGU UCUGGUUUUUAACCUUU CCUCUUGGUUCGUAUUUAAGGCA ACCUCAGGCUAUUUAAGCCAUU CCUCUUGGUUUUAACCUUU CCUCUUGGUUUUAACCUUU CCUCUUGGUUUUAACCUUU CCUCUUGGUUUUAACCUUU CCUCUUGGUUUUUAACCUUU CCUCUUGGUUUUUAACCUUU CCUCUUGGUUUUAACCUUU CCUCUUGGUUUUAACCUUU CCUCUUGGUUUUUAACCUUU CCAAAUUUUUAAACCUUUU CCAAAUUCACCUUUUAAACC CCAAAUUUUUAAACCUUUUAAACC CCAAAUUCACCAGAAAUCCCUUUU CCAAAUUCACCUUUUAAACC CCAAAUCACCUUUUUAAACC CCAAAUCACCUUUUAAACC CCAAAUCACAAAAAUCCAC CCAAAUCACAAAAAAAA	1220
AAGUCAGGUAAUUUUAAAC 1223 22017 CACUUACGAGAGUUUGUGU 1224 22035 UUUAAAAAUAAAGAUGGGU 1225 22053 UUUAAAAAUAAAGAUGGGU 1226 22051 UUUCUCUAUGUUUAAGGU 1226 22071 GGCUAUCAACCUAUGG 1228 22107 UCUGGUUUUAACACUUUGA 1239 22161 AAACCUAUUUUAAACAUUA 1233 22161 ACAAAUUUUAAACAUUC 1234 22215 ACAAAUUUUAAACAUUU 1236 22215 ACGUCAGCUAUUU 1236 22215 ACGUCAGCUAUUUAAAGC 1236 22216 ACGUCAGCUAUUUAAAGC 1236 22215 ACGUCAGCUAUUUAAAGC 1236 22251 CCAACUACAAAAUGGUA 1238 22287 ACGAUCACAGAUGCUAUU 1236 22305 GAUUGUUCCAAAAUCCUCAAAAUCCUCAAAUCCACAAACUUUAACCUUUAACCAAACUUUAACCAAAGCAAACUUUAACCAAAGCAAACAAA	1222
CACUUACGAGAGUUUGUGU 1224 22035 UUUAAAAAUAAAGAUGGGU 1225 22053 UUUUAAAAAUAAAGAUGGGU 1225 22053 UUUUCUCUAUCUUUAAGAUG 1226 22071 GGCUAUCAACCUUUGA 1228 22107 UCUGGUUUUAACACUUUGA 1238 2215 AAACCUAUUUAACACUUUGA 1233 22161 ACAAAUUUUAACACCUUU 1233 22161 ACAAAUUUUAACACCUUU 1233 22161 ACAACUUUGAGCCAUUU 1234 22215 ACAACUUUGAGCCAUUU 1236 22217 CCUCAAGACCUUUUCACCUG 1236 22287 ACGACACACAUUUAAAGC 1236 22287 ACGACACACAAAUCCAC 1236 2233 UUUGUCAAAAUCCAC 1240 2233 ACAAUCACAGAAUCCACAAAUCCA 1241 22341 UCUGUUAAGACUUUACCA 1242 22359 AUUGACCACAAACUCUCAGAGAU 1246 22413 GUUGUUCCCUCAGGAGAU 1246 22485 GUUGUCCACAAACUCUCU 1248 22467 GCUCACAAAACU	1223
UUUDAAAAUDAAGAUGGGU 1225 22053 UUUDAAAAUDAAGGU 1225 22053 UUUCCUAUGUUDAUAGAUG 1226 22071 GGCUAUCGACCUAUAGAUG 1228 22107 GUAGUUCGUAUCACCUU 1228 22107 UCUGGUUUUAACACUUUGA 1239 2215 AAACCUAUUUAACACUUUG 123 2215 ACAAAUUUUAAGGCCAUUC 1232 2215 ACAAAUUUUAAGGCCAUUC 1232 22179 CCUCUUGGUAUUUAAGCC 1234 22215 ACAACUACAGCUUUCACCUG 1235 22215 ACGACACAGCUUUCACCUG 1234 22287 ACGACACAGCUUUUAAGCC 1240 2233 UUUGGCUAUUAUGCUC 1241 22359 ACAAUCACAGGAAUCCAC 1241 22359 ACAAUCACAGGAAUUUACC 1244 22359 AUUGCCUCAAACUCUU 1244 22395 GUUGUGACCUCUAAUU 1245 22413 GUUGUCCCUCAGGAGAU 1246 22485 GUUGUCACAAACUCUCU 1248 22485 GUUGUAAAAUUCCUU	1224
UUUCUCUAUGUUNAUAAGG 1226 22071 UUUCUCUAUGUUNAUAAGA 1227 22089 GGCUAUCAACCUAUAGAUG 1228 22107 GUAGUUCUUAACACUUUGA 1228 22107 UCUGGUUUUAACACUUUGA 1230 22143 CCUCUUGGUUUUAACAUUC 1232 22179 CCUCUUGGUAUUAAGCCUAUU 1232 22179 CCUCUUGGUAUUUAAGCCUAUU 1233 22215 ACAAAUUUUAAGCCUAUU 1234 22215 ACGUCAGCUAUUUCACCUG 1234 22215 ACGUCAGCUAUUUAAAGC 1236 22287 ACGUCAGCUAUUUAAAGC 1236 22287 ACGUCAGCUAUUUAAAGC 1236 22287 ACGACUACAAAUCCAC 1240 2233 CUUGGUUCUCAAAUGCU 1241 22350 ACAAUCACAAAUCCAC 1242 22350 ACAAUCACAAGGAAUUUACC 1244 22350 GUUGUUCCCUCAAAAUCCUU 1244 22395 GUUGUUCCCUCAGGAGAUU 1241 22413 GUUGUUCCCUCAGGAGAUU 1241 22413 GCUCACUCA	1225
GGCUAUCAACCUAUAGAUG 1227 22089 GGCUAUCAACCUAUAGAUG 1228 22107 GUAGUUCUAACACUUUGA 1228 22107 UCUGGUUUUAACACUUUGA 1230 22143 AAACCUAUUUUAACACUUC 1231 22107 ACAAAUUUUAAGACCAUUC 1232 22179 ACAAAUUUUAAGACCAUUC 1233 22197 ACAAAUUUUAAGACCUAUU 1234 22215 ACAACUAGACCUAUU 1234 22215 ACGUCAGCUAUUUAAAGC 1236 22287 ACGUCAGCUAUUUAAAGC 1236 22287 ACGACUACAAUCCAC 1236 22287 ACGACACACAAAUCCAC 1240 22323 AUUGUUCAAAUCCAC 1240 22353 AUUGCUCAAAUCCAC 1241 22355 AUUGCUCAAAUCCAC 1242 22359 AUUGAACCUCUAAUUACC 1244 22355 GUUGUUCCCUCAGGAGAUU 1247 22449 UUUGAGAGUUCUUU 1241 2241 AUUACAAACUUCUCUCU 1248 22467 GCUACUAAAUUCUCUC 124	1226
GUAGUUCGUGAUCUNGCUU 1228 22107 UCUGGUUUUAACACUUUGA 1229 22125 AAACCUAUUUUAACACUUUG 1230 22143 CCUCUUGGUUUUAACAUUC 1231 22161 ACAAAUUUUAAGACCAUUC 1232 22179 CCUCAGCCUUUUCACCUG 1233 22215 ACAAUUUUAGAGCCAUUC 1234 22251 CCUCAAGACCUCAUUUCACCUG 1236 22251 ACGUCAGCUCAUUUAAAGC 1236 22251 ACGACACUACAAAUCCAC 1236 22287 ACGACACAAAUCCAC 1240 22323 AUUGUUCUCAAAUCCAC 1241 22341 ACAAUCACAGAUGCUUUGAGA 1242 22359 ACAAUCACACAAAUCCAC 1241 22350 AUUGCUGAACUUUAAGC 1244 22350 AUUGCUGAACUUUUAAGG 1244 22350 GUUGUUCCCUCAGGAGAGAUUUCCCUUAAGACUUUUUAAUG 1246 22413 GUUGUUCAAACUUUUUAAUG 1248 22467 GCUACUAAACUUCUGGAGAGAGA 1248 22467 GCUACUAAAACUUCUGGAGAGAGAA 1250 2250	1227
UCUGGULUUAACACUUUGA 1229 22125 AAACCUAUUUUAACACUUUGC 1230 22143 CCUCUUGGUAUUAACAUUA 1231 22161 ACAAAUUUUAGAGCCAUUC 1232 22179 CUUACAGCCUUUUCACCUG 1233 22197 GCUCAAGACAUUUGGGGCA 1234 22215 ACGUCAGCCUAUU 1235 22217 GCUCAAGACCUCAUU 1236 22251 ACGUCAGCUAUUUAAAGC 1236 22251 ACGACUACAGUCCAC 1236 22287 ACGACUACAAUGCUCA 1237 22287 ACGACUACAAAUGCUCA 1236 22377 CCAACUACAAAUCCAC 1241 22355 ACAAUCACAAAUCCAC 1241 22355 AUUGCUCAAAUGCU 1241 22355 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUCAAACUUUUAAUG 1248 22467 GCUACUAAACUUCUG 1248 22485 GUUCUAAAAUUCCUUCUG 1249 2280 GUUCUAAAAUUUCUG 1248<	1228
AAACCUAUUUUAAGUUGC 1230 22143 CCUCUUGGUAUUAACAUUA 1231 22161 ACAAAUUUUAGAGCCAUUC 1232 22179 CUUACAGCCUUUCACCUG 1233 22197 GCUCAAGACAUUUGGGGCA 1234 22215 ACGUCAGCUCAUUU 1236 22251 ACGUCAGCUCAUUUAAAGC 1236 22251 UUUGUUGACAAUUUAAAGC 1236 22287 ACAAUCAGAUGCUGUUG 1237 22287 ACAAUCAGAUGCUGUUG 1239 22353 CUUGCUGAAAUCCAC 1240 22323 CUUGCUGAAAUCCAC 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGCUCAAAUCCAC 1244 22359 GUUGUUCCCUCAGGAGAUG 1244 22359 GUUGUUCCCUCAGGAGAUG 1244 22395 GUUGUUCAAAUUCCUU 1246 22413 GUUGUUGAGAGUUUUUAAUG 1248 22467 GCUACUAAACUUUUCUG 1249 22485 GUUGUAAAAUUCCUUCUG 1250 22501 GUUGUGAGAGAGAGAGAGAA	1229
CCUCUUGGUAUUAACAUUA 1231 22161 ACAAAUUUUAGAGCCAUUC 1232 22179 CUUACAGCCUUUUCACCUG 1233 22197 GCUCAAGACUUUGGGGCA 1234 22215 GCUCAAGACUUUGGGGCA 1234 22215 ACGUCAGCUAUUUAAAGC 1236 22231 UUUGUUGGCUAUUUAAAGC 1236 22287 ACGACUACAAAUGCUA 1238 22287 ACGACUACAAAUGCUA 1231 22377 CCAACUACAAAUGCUCA 1241 22341 UCUGUUCACAAAUGCU 1241 22341 UCUGUUCACAAAUGCU 1241 22359 AUUGACAAAGGAAUUUACC 1243 22351 GUUGUUCCCUCAGGAGAUG 1244 22355 GUUGUUCCCUCAGGAGAUG 1244 22355 AUUGACAAACUUCUCUAAUG 1248 22431 AUUACAAACUUCUCUUCUG 1248 22487 GUUGUUCAAAUUCCCUUCUG 1248 22487 GUUGUUCAAAUUCCUUCUG 1248 22487 GUUGUUCAAAUUCCUUCUG 1248 22487 GUUGUUCAAAUUCCCUUCUG 1248 22487 GUUGUUCAAAUUCCUUCUG 1248 22487 GUUGUUCAAAUUCCUUCUG 1250 22503 140 GUUGCUGAUUACUGUGC 1252 22539 CUUCUACAACUUAAGUGCU 1251 22521 160 GUUGCUGAUCAACUUUUU 1253 22557 1	1230
ACAAAUUUUAGAGCCAUUC 1232 22179 ACAAAUUUUAGAGCCAUUC 1233 22179 CUUACAGCCUUUUCACCUG 1233 22197 GCUCAAGACAUUUGGGGCA 1234 22215 ACGUCAGCUGUUUGGGCCA 1234 22233 UUUGUUGGCUAUUU 1235 22261 CCAACUACAUUUAAAGC 1236 22251 CCAACUACAAUGCUCA 1237 22269 AAGUAUGAUGAAAUGCUC 1240 22323 CUUGCUCAAAUGCUC 1241 22341 UCUGUUAAGACUCAAUGCU 1241 22359 AUUGACAAAGGAAUGCU 1244 22359 AUUGACCUCAAAUGCU 1244 22359 AUUGACCUCAAAUGCU 1245 22413 GUUGUUCCCUCAGGAGAUG 1246 22431 AUUGACAACUUCUCUCU 1246 22431 AUUUCAAACUUCUCU 1248 22467 GCUACUAAAUUCCCUUCUG 1248 22485 GCUACUAAACUUCUG 1250 22503 AAAAAAUUCCCUUCUG 1251 22521 GUUGCUGAGAGAGAGA 1251	1231
CUUACAGCCUUUUCACCUG 1233 22197 6CUCAAGACAUUUGGGGCA 1234 22215 ACGUCAAGACAUUUGGGGCA 1234 22215 ACGUCAGCUAUUU 1235 22233 ACGUCAGCUAUUUUAAAGC 1236 22251 CCAACUACACUUUAAAGC 1237 22269 AAGUAUGUUCAAAUGCUA 1231 22377 CCAACUCAAAUGCUC 1241 22323 CUUGCUGAACUUUGAGA 1242 22359 AUUGACAAACUUUAAUGC 1244 22355 AUUGACAAACUUUAAUG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCCUCAGGAGAUG 1248 22413 GUUGUUCAAACUUUUAAUG 1248 22467 GCUACUAAAACUUCUGUG 1249 22485 GUUGUUAAAGUUCCUUCUG 1249 22485 GUUGUUAAAUUCCCUUCUG 1250 22503 AAAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUCAACUUUUU 1251 22521 CUCUACAACUUAAGUGCU 1251 22521 CUUUUCAACCUUUAAGCCU 1251 22521 CUUUUCAACCUUUAAGCCU 1251 22521 CUUUUCAACCUUUAAGCCU 1252 22539 CUUUUCAACCUUUAAGCCU 1254 22557 7	1232
GCUCAAGACAUUUGGGGCA 1234 22215 ACGUCAGCUGAUU 1235 22233 ACGUCAGCUGCAGCCUAUU 1235 22233 UUUGUUGGCUAUUUAAAGC 1236 22251 CCAACUACAUUUAAAGC 1236 22287 AAGUAUGAUGAAAUGCUA 1237 22269 AAGUAUGAUGAAAUGCUA 1241 22323 CUUGCUGAACUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUUACC 1244 22359 AUUGACAAAGGAAUUACC 1244 22355 GUUGUUCCCUCAAGAUG 1244 22355 GUUGUUCCCUCAGGAGAUG 1247 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUGAGAUUCCCUU 1247 22485 GCUACUAAAUUCCCUUCUG 1248 22485 GCUACUAAAAUUCCCUUCUG 1249 22485 GCUACUAAAAUUCCCUUCUG 1250 22501 GCUACUAAAAUUCCUUCUG 1251 22521 GUUGCUGAGAGAGAGAA 1250 22539 GUCUAUAGAGAUGUGC <td>1233</td>	1233
ACGUCAGCUGCAGCCUAUU 1235 22233 UUUGUUGGCUAUUUAAAGC 1236 22251 UUUGUUGGCUAUUUAAAGC 1236 22251 CCAACUACAUUUAUGCUCA 1237 22269 AAGUAUGAUGAAAUGCUA 1238 22287 ACAAUCACAGAUGCUGUUG 1240 22323 CUUGCUCAAAUCCAC 1241 22341 UCUGUUAAGACUCAAGA 1242 22359 AUUGACAAGGAAUGCU 1244 22359 AUUGACAAAGGAAUG 1246 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUGAGAUUCCCUU 1247 22485 GCUACUAAAUUCCCUUCUG 1248 22467 GCUACUAAAAUUCCCUUCUG 1251 22501 GCUACUAAAAUUCCUUCUG 1251 22521 GCUACUAAAAUUCCUAAAUUCUG 1251 22521 GUUGCUGAACAUGUGC 1252 22539 CUCUACAACUCAACAUCU	1234
UUUGUUGGCUAUUUAAAGC 1236 22251 CCAACUACAUUUAAAGC 123 22269 AAGUAUGAUGAAAUGGUA 123 22287 AAGUAUGAUGAAAUGGUA 1238 22387 ACAAUCACAGAUGCUGUUG 1240 22323 GAUUGUUCUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUGC 1244 22359 AUUGACAAAGGAAUG 1244 22355 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUGAACUUCCUUAU 1247 22449 AUUACAAACUUGUGCCUU 1246 22431 AUUACAAACUUCUGUCCUU 1248 22467 GCUACUAAAUUCCCUUCUG 1248 22485 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCUUCUG 1251 22501 GCUACUAAAAUUCCCUUCUG 1251 22521 GCUACUAAAAUUCCUUCUG 1251 22521 GUUGCUAAAAUUCUG 1251 22521 GUUGCUAAACUUCUGUG 1252 22539 CUCUACAACUCAACAUUUU	1235
CCAACUACAUUNAUGCUCA 1237 22269 AAGUAUGAUGAAAAUGGUA 1238 22287 AAGUAUGAUGAAAUGGUA 1238 22305 ACAAUCACAGAUGCUGUUG 1240 22323 GAUUGUUCUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUUUACC 1244 22357 CAGACCUCUAAUUUCAGGG 1244 22355 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUCCCUCAGGAGAUG 1246 22413 GUUGUUGAGAUUCCCUU 1247 22449 UUUGAGAGCUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1248 22485 GCUACUAAAUUCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1251 22501 GCUACUAAAAUUCCCUUCUG 1251 22521 GUUGCUGAUUAAUUUU 1251 22521 GUUGCUGAACUUUUU 1253 22551 CUCUACAACUUAAGUGCU 1253 22551	1236
AAGUAUGAUGAAAUGGUA 1238 22287 ACAAUCACAGAUGCUGUUG 1239 22305 GAUUGUUCUCAAAAUCCAC 1240 22323 GAUUGUUCUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUUUACC 1243 22377 CAGACCUCUAAUUUCAGGG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22443 AUUGUGAGAUUCCUUAUA 1246 22449 AUUACAAACUUCUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1250 22503 AAAAAAAUUCCAUCUAAUUCUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22521 GUUGCUGAACUCAACUUUU 1251 22539 CUCUACAACUUAAGUGCU 1253 22557 CUCUACAACCUUUAAGUGCU 1254 22557	1237
ACAAUCACAGAUGCUGUUG 1239 22305 GAUUGUUCUCAAAAUCCAC 1240 22323 CUUGCUGAACUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUUUACC 1243 22377 CAGACCUCUAAUUUCAGGG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCCUCAGGAGAUG 1246 22431 AUUACAAACUUCUCUCUU 1247 22449 UUUGAGGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUUUG 1249 22485 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1250 22503 AAAAAAUUCCAUCUAAUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22521 GUUGCUAACUCAACUUUU 1253 22539 CUCUACAACUUAAGUGCU 1253 22537	1238
GAUUGUUCUCAAAAUCCAC 1240 22323 CUUGCUGAACUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUUUACC 1243 22377 CAGACCUCUAAUUUCAGGG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCUCAGGAGAUG 1246 22431 AUUACAAACUUCUGUCUU 1247 22449 UUUGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1250 22503 AAAAAAUUCCAUCUGUGC 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22521 GUUGCAACUCAACAUUUU 1253 22539 CUCUACAACUUAAGUGCU 1253 22537	1239
CUUGCUGAACUCAAAUGCU 1241 22341 UCUGUUAAGAGCUUUGAGA 1242 22359 AUUGACAAAGGAAUUUACC 1243 22377 CAGACCUCUAAUUUCAGGG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCUCAGGAGAUG 1245 22413 GUUGUGAGUUCCUUAUA 1246 22431 AUUACAAACUUCUGUCUU 1247 22449 UUUGAGGAGGAGAA 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22539 CUCUACAACUUAAGUGCU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1240
UCUGUNAAGAGCUUNGAGA 1242 22359 AUUGACAAAGGAAUUNACC 1243 22377 CAGACCUCUAAUUUCAGGG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCCUCAGGAGAUG 1245 22431 AUUGAGAGUUCCCUNANA 1246 22431 AUUGAGAGUUCCUNANUG 1247 22449 UUUGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GUCUAUGGAGAGAA 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22521 GUUGCUGAUUACUCUGUGC 1252 22539 CUCUACAACUUAAGUGCU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1241
AUUGACAAAGGAAUUUACC 1243 22377 CAGACCUCUAAUUUCAGGG 1244 22385 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCCUCAGGAGAUG 1245 22431 GUUGUGAGUUCCUAAUA 1246 22431 AUUACAAACUUCUGUCUU 1247 22449 UUUGGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22521 GUUGCUGAUUACUCUGUGC 1252 22539 CUCUACAACCUUUAAGUGCU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1242
CAGACCUCUAAUUUCAGGG 1244 22395 GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUUCCCUCAGGAGAUG 1245 22431 GUUGUGAGAUUCCCUAAUA 1246 22431 AUUACAAACUUGUGUCUU 1247 22449 UUUGGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GUCUAUGCAGGAGAA 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1243
GUUGUUCCCUCAGGAGAUG 1245 22413 GUUGUGAGAUUCCCUAAUA 1246 22431 AUUACAAACUUGUGUCCUU 1247 22449 AUUGGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GCUACUAAAUUCCCUUCUG 1250 22503 AAAAAAUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22539 CUCUACAACUCAACAUUUU 1253 22537 UUUUCAACCUUUAAGUGCU 1254 22557	1244
GUUGUGAGAUUCCCUAAUA 1246 22431 AUUACAAACUUGUGUCCUU 1247 22449 UUUGGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GUCUAUGCAUGGAGAA 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1245
AUUACAAACUUGUGUCCUU 1247 22449 UUUGGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GUCUAUGCAUGGAGAA 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1251 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1246
UNUGGAGAGGUUUUUAAUG 1248 22467 GCUACUAAAUUCCCUUCUG 1249 22485 GUCUAUGCAUGGAGAA 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1252 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1247
GCUACUAAAUUCCCUUCUG 1249 22485 GUCUAUGCAUGGAGAGAA 1250 22503 AAAAAAUUUCUAAUUGUG 1251 22521 GUUGCUGAUUACUCUGUGG 1252 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22557	1248
GUCUAUGCAUGGAGAGAA 1250 22503 4AAAAAAUUUCUAAUUGUG 1251 22521 CUUGCUGAUUACUCUGUGC 1252 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22575	1249
AAAAAAUUUCUAAUUUGUG 1251 22521 GUUGCUGAUUACUCUGUGC 1252 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22575	1250
GUUGCUGAUUACUCUGUGC 1252 22539 CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22575	1251
CUCUACAACUCAACAUUUU 1253 22557 UUUUCAACCUUUAAGUGCU 1254 22575	1252
UNUUCAACCUUUAAGUGCU 1254 22575	1253
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Н	-	CUGCAUAGACAUUGGAGAA	CCUUGACUACAAAAGAAUC	UNUGUCUNACANCANCOCC	CAGUUUGUCCUGGCGCUAU	UAUAAUCAGCAAUAACACC	CAUCUGGCAAUUUAUAAUU	GGACACCCAUGAAAUC	UCCUAGUAUUCCAAGCAAG	UUGAAGUAGCAUCAAUGUU	UAUAAUUAUAAUUACCAGU	GUCUAAGAUACCUAUAUUU	AGGGCCUAAGCUUGCCAUG		CAGGGGAGAAAGGCACAUU	GGGUGCAAGGUUUGCCAUC	AACAAUUAAGAGCAGGUGG	AAUCAUUUAAUGGCCAAUA	UAGUGGUGUAAAAACCAUA	GUUGGUAGCCAAUGCCAGU	GUACUACAACUCUGUAAGG	UNAAAAGUUCAAAAGAAAG				-	-	-	-	GGCCAAAUUGUUGAAAUGG	UGAAAUCAGAAACAUCACG	GAUCUCGAACGGAAUCAGU	AUAUUUCAGAUGUUUUAGG	CGCAAGGUGAAAUGUCUAA	CACUUACACCCCCAAAAGC	UUGUUCCAGGUGUAAUUAC	CAACUUCAGAUGAAGCAUU	CAUCUUGAUAUAGAACAGC	AAACAUCAGUGCAGUUAAC	CUGCAUGAAUUGCUGUAGA	AAGCUGGUGUGAGUGAUC
22593	22611	22629	22647	22665	22683	22701	22719	22737	22755	22773	22791	22809	22827	22845	22863	22881	22899	22917	22935	22953	22971	22989	23007	23025	23043	23061	23079	23097	23115	23133	23151	23169	23187	23205	23223	23241	23259	23277	23295	23313	23331
1255	1256	1257	1258	1259	1260	1261	1262	1263	1264	1265	1266	1267	1268	1269	1270	1271	1272	1273	1274	1275	1276	1277	1278	1279	1280	1281	1282	1283	1284	1285	1286	1287	1288	1289	1290	1291	1292	1293	1294	1295	1296
\dashv	\dashv	UNCUCCAAUGUCUAUGCAG	GAUUCUUUUGUAGUCAAGG	GGAGAUGAUGUAAGACAAA	AUAGCGCCAGGACAAACUG	GGUGUUAUUGCUGAUUAUA	AAUUAUAAAUUGCCAGAUG		CUUGCUUGGAAUACUAGGA	AACAUUGAUGCUACUUCAA	ACUGGUAAUUAUAAUUAUA		CAUGGCAAGCUUAGGCCCU	-	AAUGUGCCUUUCUCCCCUG	_	CCACCUGCUCUNAAUUGUU	UAUUGGCCAUUAAAUGAUU	UAUGGUUUUUACACCACUA	ACUGGCAUUGGCUACCAAC	_	CUUUCUUUGAACUUUUAA		_	_1	-	_	ACUGGUGUGUNAACUCCUU	\dashv	-	-	-1				GUAAUUACACCUGGAACAA		_	GUDAACUGCACUGAUGUUU	UCUACAGCAAUUCAUGCAG	GAUCAACUCACACCAGCUU
22575	22593	22611	22629	22647	22665	22683	22701	22719	22737	22755	22773	22791	22809	22827	22845	22863	22881	22899	22917	22935	22953	22971	22989	23007	23025	23043	23061	23079	23097	23115	23133	23151	23169	23187	23205	23223	23241	23259	23277	23295	23313
1255	1256	1257	1258	1259	1260	1261	1262	1263	1264	1265	1266	1267	1268	1269	1270	1271	1272	1273	1274	1275	1276	1277	1278	1279	1280	1281	1282	1283	1284	1285	1286	1287	1288	1289	1290	1291	1292	1293	1294	1295	1296
\rightarrow	\rightarrow	UUCUCCAAUGUCUAUG		_	35 AUAGCGCCAGGACAACUG	-	_	GAUUUCAUGGGUUGU	CUUGCUUGGAAUACUA	AACAUUGAUGCUACUI	73 ACUGGUAAUUAUAAUUAUA	_	CAUGGCAAGCUUAGG	UUUGAGAGAGACAUAI		GAUGGCAAACCUUGC/	_		-			CUUUCUUUUGAACUUI	\rightarrow	17 UGUGGACCAAAAUUAUCCA	_	3 CAGUGUGUCAAUUUAAUU	_	-	-		CGUGAUGUUUCUGAUI	ACUGAUUCCGUUCGA(CCUAAAACAUCUGAAA		GCUUUUGGGGGGUGUA				7 GUUAACUGCACUGAUGUUU	UCUACAGCAAUUCAUG	3 GAUCAACUCACACCAGCUU
22575	22593	22611	22629	22647	22665	22683	22701	22719	22737	22755	22773	22791	22809	22827	22845	22863	22881	22899	22917	22935	22953	22971	2298	23007	23025	23043	23061	23079	23097	23115	23133	23151	23169	23187	2320	23223	23241	23259	23277	23295	23313

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UGGCGCAUAUUUCUACUG GGAAACAAUGUUUCCAGA GGAAACAAUGUUUCCAGA AUAGGAGCUGACCUCUUA GACAUUCCUAUUGAGUGCG GACAUUCCUAUUGAGUGCG GACAUUCCUAUUGAGUGCG GACAUUCCUAUUGAGUGCG GACAUUCCUAUUGAGUAA GAUACCCAUUCCUAUACC GCAUUCCUAUUCCAAAAU GCAUUCCUAUUCCAAAAU GGCAUUCCUAUACCCAAAAAU AAAACCUCCGUAGUAACC GCAAAACCUCCGUAGAAAA AAAACCUCCGUAGAAAAA AAAACCUCCGUAGAAAAA AAAACCUCCGUAGAAAAA AAAACCUCCGUAGAAAAA AAAACCUCCGUAGAAAAA AAAACCUCCGUAGAAAAA AAAACCUCCGUAGAAAAA AAAACCUCCAAAAAAAAAA	UGGCGCAUAUAUUCUACUG GGAAACAAUGUAUUCCAGA	ACUCAAGCAGGCUGUCUUA	AUAGGAGCUGAGCAUGUCG	GACACUUCUUAUGAGUGCG	GACAUUCCUAUUGGAGCUG	GGCAUUGUGCUAGUUACC	CAUACAGUUUCUUUAUUAC	CGUAGUACUAGCCAAAAAU	UCUAUUGUGGCUUAUACUA	AUGUCUUUAGGUGCUGAUA	AGUUCAAUUGCUUACUCUA	AAUAACACCAUUGCUAUAC	CCUACUAACUUUCAAUUA	AGCAUUACUACAGAAGUAA	AUGCCUGUUUCUAUGGCUA	AAAACCUCCGUAGAUUGUA	AAUAUGUACAUCUGCGGAG	GAUUCUACUGAAUGUGCUA	AAUUJGCUUCUCCAAUAUG	GGUAGCUUUUGCACACAC	CUAAAUCGUGCACUCUCAG	GGUAUUGCUGCUGAACAGG	GAUCGCAACACACGUGAAG	GUGUUCGCUCAAGUCAAAC	CAAAUGUACAAAACCCCAA	ACUUUGAAAUAUUUGGUG	GGUUUUAAUUUUUCACAAA	AUAUUACCUGACCCUCUAA	AAGCCAACUAAGAGGUCUU	UNDANUGAGGACUUGCUCU	UUUAAUAAGGUGACACUCG	SCUGAUGCUGGCUUCAUGA AACCAALIALICCCAAIICCC	AAGCAAUGGIGAIIIAAIIGGIA	AGAGALICI CALIII IGLIGGG	CAGAGUICAAUGGACIIUA	ACAGUGUUGCCACCUCUGC	CUCACUGAUGAUAUGAUUG	GCUGCCUACACUGCUGCUC	CUAGUUAGUGGUACUGCCA
UGGCGCAUAUAUUCUACUG GGAAACAAUGUAUUCUACUG GGAAACAAUGUAUUCCAGA ACUCAAGCAGCCACGUCUCUA AUAGGAGCUGCAGCAGAAAU ACACCUNCUUAUGAGCGAG GGCAUUUGUACUACCAAAAU ACACCUUCUUAGGAGCUA ACACCUUCUAUGCCAAAAAU ACACCUCCUAUUCCAAUAC CCUAGUACACUUCCUAAAAA ACACCACUUCCAAAAAU ACACCUCCAAAAAU ACACCUCCAAAAAA ACACCACUUCCCAAAAAA ACACCACUUCCCAAAAAA ACACCACUUACCAAAAA ACACCACUUACCAAAAA ACACCACUUACCAAAAA ACACCACCUCCAAAAAA ACACCACCUCCAAAAAA ACACCACCUCCAAAAAA ACACCACCUCCAAAAAA ACACCACCUCCAAAAAA ACACCACCUCCAAAAAAAA	23331	23367	23385	23403	23421	23439	23457	23475	23493	23511	23529	23547	23565	23583	23601	23619	23637	23655	23673	23691	23709	23727	23745	23763	23781	23799	23817	23835	23853	238/1	23889	23907	2367	23061	23979	23997	24015	24033	24051
UGGGGCAUAUAUUCC GGAAACAACAGGCGGG AUAGGAGCUGGCGGG GACAUUCGUAGCOAGG GGCAUUUGGGCUAGG GGCAUUUGGGCUAGG GGCAUUUGGCCAACAGGCCAACACCG AUAGGCCCGUAGGCCAACACCG GGUAGCCCCGUAGGCCAACACCG GGUAGCCCCGCAACACCG GGUAGCCCCCAACACCCAACACCG GGUAGCCCCCAACACCCAACACCCAACACCCAACACCCCAACACCCC	1297	1299	1300	1301	1302	1303	1304	1305	1306	1307	1308	1309	1310	1311	1312	1313	1314	1315	1316	1317	1318	1319	1320	1321	1322	1323	1324	1325	1326	1327	1328	1329	1221	1332	1333	1334	1335	1336	1337
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24105	24141	24159	24177	24195	24213	24231	24249	24267	24285	24303	24321	24339	24357	24375	24393	24411	24429	24447	24465	24483	24501	24519	24537	24555	24573	24591	24609	24627	24645	24663	24681	24699	24717	24735	24753	24771	24789	24807	24825	24843
1339	1341	1342	1343	1344	1345	1346	1347	1348	1349	1350	1351	1352	1353	1354	1355	1356	1357	1358	1359	1360	1361	1362	1363	1364	1365	1366	1367	1368	1369	1370	1371	1372	1373	1374	1375	1376	1377	1378	1379	1380
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24087	24123	24141	24159	24177	24195	24213	24231	24249	24267	24285	24303	24321	24339	24357	24375	24393	24411	24429	24447	24465	24483	24501	24519	24537	24555	24573	24591	24609	24627	24645	24663	24681	24699	24717	24735	24753	24771	24789	24807	24825
1339	1341	1342	1343	1344	1345	1346	1347	1348	1349	1350	1351	1352	1353	1354	1355	1356	1357	1358	1359	1360	1361	1362	1363	1364	1365	1366	1367	1368	1369	1370	1371	1372	1373	1374	1375	1376	1377	1378	1379	1380
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27129	27165	27183	27201	27219	27237	27255	27273	27291	27309	27327	27345	27363	27381	27399	27417	27435	27453	27471	27489	27507	27525	27543	27561	27579	27597	27615	27633	27651	27669	27687	27705	27723	27741	27759	27777	27795	27813	27831	27849	27867
1508	1509	1510	1511	1512	1513	1514	1515	1516	1517	1518	1519	1520	1521	1522	1523	1524	1525	1526	1527	1528	1529	1530	1531	1532	1533	1534	1535	1536	1537	1538	1539	1540	1541	1542	1543	1544	1545	1546	1547	1548
UNAUGAGGACUUUCAGGAU	ACGUUAUAUAAGUUCAAU	UAGUGAGACAAUUAUUAA	AGCCUCUAACUAAGAAGAA	AUUAUUCGGAGUUAGAUGA	AUGAAGAACCUAUGGAGUU	UAGAUUAUCCAUAAAACGA	AACAUGAAAAUUAUUCUCU	UNCCUGACAUUGAUUGUAU	UUUACAUCUUGCGAGCUAU	UAUCACUAUCAGGAGUGUG	GUUAGAGGUACGACUGUAC	CUACUAAAAGAACCUUGCC	CCAUCAGGAACAUACGAGG	GGCAAUUCACCAUUUCACC	CCUCUUGCUGACAAUAAAU	_		GCUUGUGCUGACGGUACUC	CGACAUACCUAUCAGCUGC	CGUGCAAGAUCAGUUUCAC	CCAAAACUUUUCAUCAGAC	CAAGAGGAUCAACAAG	GAGCUCUACUCGCCACUUU	UNUCUCAUUGUUGCUGCUC	CUAGUAUUUUAAUACUUU	UGCUUCACCAUUAAGAGAA	AAGACAGAAUGAAUGAGCU	UCACUUDAAUUGACUUCUA	AUUUGUGCUUUUUAGCCUU	UNCUGCUAUUCCUUGUUUU	UAAUAAUGCUUAUUAUAUU	UUUGGUUUUCACUCGAAAU	UCCAGGAUCUAGAAGAACC	CUUGUACCAAAGUCUAAAC	CGAACAUGAAACUUCUCAU	UNGUNUGACUUGUAUUUC	CUCUAUGCAGUUGCAUAUG	GCACUGUAGUACAGCGCUG	GUGCAUCUAAUAAACCUCA	AUGUGCUUGAAGAUCCUUG
27111	27147	27165	27183	27201	27219	27237	27255	27273	27291	27309	27327	27345	27363	2/381	27399	27417	27435	27453	27471	27489	27507	27525	27543	27561	27579	27597	27615	27633	27651	27669	27687	27705	27723	27741	27759	27777	27795	27813	27831	27849
1507	1509	1510	1511	1512	1513	1514	1515	1516	1517	1518	1519	1520	1521	1522	1523	1524	1525	1526	1527	1528	1529	1530	1531	1532	1533	1534	1535	1536	1537	1538	1539	1540	1541	1542	1543	1544	1545	1546	1547	1548
27111 UUAUGAGGACUUUCAGGAU	_	+-	-		AUGAAGAACCUAUGG	UAGAUUAUCCAUAAA	AACAUGAAAAUUAUU			UAUCACUAUCAGGAG		CUACUAAAAGAACCU	27363 CCAUCAGGAACAUACGAGG	GGCAAUUCACCAUUU		UNUGCACUAACUUGC	AGCACACACUUUGCL	_	CGACAUACCUAUCAG	_	_			\dashv	CUAGUAUUUUUAAUA		AAGACAGAAUGAAUG	UCACUUUAAUUGACU	_	_			UCCAGGAUCUAGAAG	CUUGUACCAAAGUCU	27759 CGAACAUGAAACUUCUCAU			27813 GCACUGUAGUACAGCGCUG		27849 AUGUGCUUGAAGAUCCUUG

		1	Г	Γ	Г			Γ									<u> </u>															_	_							Γ
3242	3244	3245	3246	3247	3248	3249	3250	3251	3252	3253	3254	3255	3256	3257	3258	3259	3260	3261	3262	3263	3264	3265	3266	3267	3268	3269	3270	3271	3272	3273	3274	3275	3276	3277	3278	3279	3280	3281	3282	3283
GCCUCUGCUUCCCUCUGCG	ACUACGUGAUGAGGAGCGA	AUUUCUUGAAUUACCGCGA	ACUGCUGCCAGGAGUUGAA	AGCAGGAGAAUUUCCCCUA	ACCUCCGCUAGCCAUUCGA	CGCGAGGGCAGUUCACCA	UCUGUCUAGCAGCAAUAGC	GCUCUCAAGCUGGUUCAAU	GCCUUUACCAGAAACUUUG	nneeccnnennennee	AGAUUUCUUAGUGACAGUU	UUUAGAUGCCUCAGCAGCA	ACGUUUUUGGCGAGGCUUU	GUACUGUUUUGUGGCAGUA	AAAUGCUUGAGUGACGUUG	UUCUGGACCACGUCUCCCA	GAAAUUUCCUUGGGUUUGU	GAUUAGGUCUUGGUCCCCG	GUAAUCAGUUCCUUGUCUG	AAUUUGCGGCCAAUGUUUG	ACUUGGAGCAAAUUGUGCA	UCCAAAGAAUGCAGAGGCA	CAUGCCAAUGCGUGACAUU	UCCCGAAGGUGUGACUUCC	AUGAUAAGUCAGCCAUGUU	AUCCAAUUUAAUGGCUCCA	GAAUUGUGGAUCUUUGUCA	CAGUAUGACGUUGUCUUUG	GUCAAUGUGCUUGUUCAGC	UGGGAAUGUUUUGUAUGCG	CUUUUUAGGCUCUGUUGGU	AGUCUUUUUCUUUUGUCC	CAAAGGCUGAGCUUCAUCA	cnncnnnnencncneceec	AAGAGUCACAGUGGGCUGC	CAUGUCAGCCGCAGGAAGA	UUGUCUGGAGAAAUCAUCC	ACUCAUGGAAUUUUGAAGU	UGAAUCAGCAGAAGCUCCA	╙
28641	28677	28695	28713	28731	28749	28767	28785	28803	28821	28839	28857	28875	28893	28911	28929	28947	28965	28983	29001	29019	29037	29055	29073	29091	29109	29127	29145	29163	29181	29199	29217	29235	29253	29271	29289	29307	29325	29343	29361	29379
1591	1593	1594	1595	1596	1597	1598	1599	1600	1601	1602	1603	1604	1605	1606	1607	1608	1609	1610	1611	1612	1613	1614	1615	1616	1617	1618	1619	1620	1621	1622	1623	1624	1625	1626	1627	1628	1629	1630	1631	1632
CGCAGAGGGAAGCAGAGGC	UCGCUCCUCAUCACGUAGU	UCGCGGUAAUUCAAGAAAU	UUCAACUCCUGGCAGCAGU	UAGGGGAAAUUCUCCUGCU	ucgaaugecuagegeageu	UGGUGAAACUGCCCUCGCG	GCUAUUGCUGCUAGACAGA	AUUGAACCAGCUUGAGAGC	CAAAGUUUCUGGUAAAGGC	CCAACAACAAGGCCAA	AACUGUCACUAAGAAAUCU	UGCUGCUGAGGCAUCUAAA	AAAGCCUCGCCAAAAACGU	UACUGCCACAAAACAGUAC	CAACGUCACUCAAGCAUUU	UGGGAGACGUGGUCCAGAA	ACAAACCCAAGGAAAUUUC	CGGGGACCAAGACCUAAUC	CAGACAAGGAACUGAUUAC	CAAACAUUGGCCGCAAAUU	NGCACAAUUUGCUCCAAGU	NGCCNCNGCANNCNNGGA	AAUGUCACGCAUUGGCAUG	GGAAGUCACACCUUCGGGA	AACAUGGCUGACUUAUCAU	UGGAGCCAUUAAAUUGGAU	UGACAAAGAUCCACAAUUC	CAAAGACAACGUCAUACUG	GCUGAACAAGCACAUUGAC	CGCAUACAAAACAUUCCCA	ACCAACAGAGCCUAAAAAG	GGACAAAAGAAAAGACU	UGAUGAAGCUCAGCCUUUG	GCCGCAGAGACAAAGAAG	GCAGCCCACUGUGACUCUU	UCUUCCUGCGGCUGACAUG	GGAUGAUUUCUCCAGACAA	ACUUCAAAAUUCCAUGAGU	UGGAGCUUCUGCUGAUUCA	AACUCAGGCAUAAACACUC
28623	28659	28677	28695	28713	28731	28749	28767	28785	28803	28821	28839	28857	28875	28893	28911	28929	28947	28965	28983	29001	29019	29037	29055	29073	29091	29109	29127	29145	29163	29181	29199	29217	29235	29253	29271	29289	29307	29325	29343	29361
1591	1593	1594	1595	1596	1597	1598	1599	1600	1601	1602	1603	1604	1605	1606	1607	1608	1609	1610	1611	1612	1613	1614	1615	1616	1617	1618	1619	1620	1621	1622	1623	1624	1625	1626	1627	1628	1629	1630	1631	1632
23 CGCAGAGGGAAGCAGAGGC G41 CGGCAGGCCUCUCU	+	377 UCGCGGUAAUUCAAGAAAU	-	\dashv	UCGAAUGGCUAGCGC		GCUAUUGCUGCUAGA	85 AUUGAACCAGCUUGAGAGC	CAAAGUUUCUGGUAA	-		UGCUGCUGAGGCAUC	\dashv	\dashv	CAACGUCACUCAAGC/	\dashv	ACAAACCCAAGGAAAI	CGGGGACCAAGACCU	-		-	-	AAUGUCACGCAUUGG	GGAAGUCACACCUUC	-				-		\dashv	GGACAAAAAGAAAAA(-		-	-	-	UGGAGCUUCUGCUGA	61 AACUCAGGCAUAAACACUC
28623	28659	28677	28695	28713	28731	28749	28767	28785	28803	28821	28839	28857	28875	28893	28911	28929	28947	28965	28983	29001	29019	29037	29055	29073	29091	29109	29127	29145	29163	29181	29199	29217	29235	29253	29271	29289	29307	29325	29343	29361

antisense strand. The upper and lower sequences in the Table can further comprise a chemical modification having Formulae Istructure B, BNN, NN, BNSN, or NSN, where B stands for any terminal cap moiety, N stands for any nucleotide (e.g., thymidine) Formula I). The upper sequence is also referred to as the sense strand, whereas the lower sequence is also referred to as the example about 1, 2, 3, or 4 nucleotides in length, preferably 2 nucleotides in length, wherein the overhanging sequence of the and s stands for phosphorothioate or other internucleotide linkage as described herein (e.g. internucleotide linkage having The 3'-ends of the Upper sequence and the Lower sequence of the siNA construct can include an overhang sequence, for lower sequence is optionally complementary to a portion of the target sequence. The overhang can comprise the general VII or any combination thereof (see for example chemical modifications as shown in Table V herein).

Table III: SARS synthetic siNA and Target Sequences

Target Pos	Target	SeqID	RPI#	Aliases	Sequence	SeqID
UGAAL	UGAAUGAAGAGGUUGCCAUCAUU	3303		SARS:1657U21 siRNA sense	AAUGAAGAGGUUGCCAUCATT	3311
UGU	UGUUGCAUCUCCACAGGAGUGUA	3304		SARS:1166U21 siRNA sense	UUGCAUCUCCACAGGAGUGTT	3312
CUC	CUCAAAGCAAGGGACUUUACCGU	3305		SARS:2383U21 siRNA sense	CAAAGCAAGGGACUUUACCTT	3313
SUG	CUGUGUAAAUGGCCUCAUGCUCU	3306		SARS:2600U21 siRNA sense	GUGUAAAUGGCCUCAUGCUTT	3314
nnn	UNUGUECUUGCUECUGUCUACAG	3307		SARS:26574U21 siRNA sense	UGUGCUUGCUGCUGUCUACTT	3315
ACU	ACUUGUCAUUGGUGCUGUGAUCA	3308		SARS:26792U21 siRNA sense	UUGUCAUUGGUGCUGUGAUTT	3316
3	UUGAACCAGCUUGAGAGCAAAGU	3309		SARS:28788U21 siRNA sense	GAACCAGCUUGAGAGCAAATT	3317
ದ್ದ	ecnnennnacconcueecnconen	3310		SARS:26531U21 siRNA sense	UNGUUUCCUCUGGCUCUUTT	3318
<u>-</u>		2202		SARS:1675L21 siRNA (1657C)	TTHINGINGIOOVVOOCHVOIL	2240
3		3		SARS-11841 21 siRNA (1166C)		500
<u></u>	UGUUGCAUCUCCACAGGAGUGUA	3304		antisense	CACUCCUGUGGAGAUGCAATT	3320
}		1000		SARS:2401L21 siRNA (2383C)		
3	CUCAAAGCAAGGACUUUACCGU	3302		antisense	GGUAAAGUCCCUUGCUUUGII	3321
Š	CHGHGHAAAHGGCCHCAHGCHCH	3306		SARS.ZOIGLZI SIRINA (ZOUUC) antisense	AGCALIGAGGCCALIIIIACACTT	3333
		8		SARS: 265921 21 siRNA (26574C)		7700
Ħ	UUUGUGCUUGCUGUCUACAG	3307		antisense	GUAGACAGCAGCACATT	3323
				SARS:26810L21 siRNA (26792C)		
PC	<u>ACUUGUCAUUGGUGCUGUGAUCA</u>	3308		antisense	AUCACAGCACCAAUGACAATT	3324
Ξ		0000		SARS:28806L21 siRNA (28788C)		100
키	UUGAALLAGLUUGAGAGLAAAGU	3309		antisense	UNDECUCARECUEEUUCII	3372
ز		2240		SARS:26549L21 siRNA (26531C)	***************************************	0000
3 5	***************************************	2 2 2				0250
3 9		3303		SARS: 165/UZ1 SIRNA Stab04 sense	-1.	332/
3	UGUUGCAUCUCCACAGGAGUGUA	3304		SARS:1166U21 siRNA stab04 sense	B uuGcAucuccAcAGGAGuGII B	3328
리	CUCAAAGCAAGGGACUUUACCGU	3305		SARS:2383U21 siRNA stab04 sense	B cAAAGcAAGGGAcuuuAccTT B	3329
징	CUGUGUAAAUGGCCUCAUGCUCU	3306		SARS:2600U21 siRNA stab04 sense	B GuGuAAAuGGccucAuGcuTT B	3330
밁	UUUGUGCUUGCUGUCUACAG	3307		SARS:26574U21 siRNA stab04 sense	B uGuGcuuGcuGcuGucuAcTT B	3331
Ą	ACUUGUCAUUGGUGCUGUGAUCA	3308		SARS:26792U21 siRNA stab04 sense	B uuGucAuuGGuGcuGuGAuTT B	3332
3	UUGAACCAGCUUGAGAGCAAAGU	3309		SARS:28788U21 siRNA stab04 sense	B GAAccAGcuuGAGAGcAAATT B	3333
ပ္ပ	ecnnennnacacaecacanen	3310		SARS:26531U21 siRNA stab04 sense	B uuGuuuccucuGGcucuuTT B	3334
n	UGAAUGAAGAGGUUGCCAUCAUU	3303		SARS:1675L21 siRNA (1657C) stab05 antisense	uGAuGGcAAccucuucAuuTsT	3335
				SARS:1184L21 siRNA (1166C) stab05	4	
ฐ	UGUUGCAUCUCCACAGGAGUGUA	3304		antisense	cAcuccuGuGGAGAuGcAATsT	3336

CUCAAAGCAAGGGACUUUACCGU	3305	SARS:2401L21 siRNA (2383C) stab05 antisense	GGuAAAGucccuuGcuuuGTsT	3337
CUGUGUAAAUGGCCUCAUGCUCU 3306		SARS:2618L21 siRNA (2600C) stab05 antisense	AGCAUGAGGCCAUUUACACTST	3338
UUUGUGCUUGCUGCUGUCUACAG 3307		SARS:26592L21 siRNA (26574C) stab05 antisense	GuAGACAGCAGCACATST	3339
ACUUGUCAUUGGUGCUGUGAUCA 3308		SARS:26810L21 siRNA (26792C) stab05 antisense	AucAcAGcAccAAuGAcAATsT	3340
UUGAACCAGCUUGAGAGCAAAGU 3309	1	SARS:28806L21 siRNA (28788C) stab05 antisense	unuGcucucAAGcuGGuucTsT	3341
GCUUGUUUUCCUCUGGCUCUUGU 3310	1	SARS:26549L21 siRNA (26531C) stab05 antisense	AAGAGccAGAGGAAAAcAATsT	3342
UGAAUGAAGAGGUUGCCAUCAUU 3303		SARS:1657U21 siRNA stab07 sense	B AAuGAAGAGGuuGccAucATT B	3343
UGUUGCAUCUCCACAGGAGUGUA 3304		SARS:1166U21 siRNA stab07 sense	B uuGcAucuccAcAGGAGuGTT B	3344
CUCAAAGCAAGGGACUUUACCGU 3305		SARS:2383U21 siRNA stab07 sense	B CAAAGCAAGGGACUUUACCTT B	3345
		SARS:2600U21 siRNA stab07 sense	B GuGuAAAuGGccucAuGcuTT B	3346
UUUGUGCUUGCUGCUGUCUACAG 3307		SARS:26574U21 siRNA stab07 sense	B uGuGcuuGcuGcuGucuAcTT B	3347
-	Ì	SARS:26792U21 siRNA stab07 sense	B uuGucAuuGGuGcuGuGAuTT B	3348
UUGAACCAGCUUGAGAGCAAAGU 3309	ı	SARS:28788U21 siRNA stab07 sense	B GAAccAGcuuGAGAGcAAATT B	3349
GCUUGUUUCCUCUGGCUCUUGU 3310	İ	SARS:26531U21 siRNA stab07 sense	B uuGuuuccucuGGcucuuTT B	3350
HGAAHGAAGAGGHHGCCAHCAHH		SARS:1675L21 siRNA (1657C) stab11	+ +	1 3
+-		SARS:11841 21 ciPNA (1166C) etab11	uchuc ochaccucuuchuu s I	3331
UGUUGCAUCUCCACAGGAGUGUA 3304		antisense	cAcuccu Gu GGA GAu GcAATsT	3352
		SARS:2401L21 siRNA (2383C) stab11		
CUCARAGCAAGGGACUUUACCGU 3305		antisense	GGuAAAGucccuuGcuuuGTsT	3353
CUGUGUAAAUGGCCUCAUGCUCU 3306		SARS:2618L21 siRNA (2600C) stab11 antisense	AGcAuGAGGccAuuuAcAcTsT	3354
		SARS:26592L21 siRNA (26574C)	+ +	
+		SARS:26810L21 siRNA (26792C)	GUAGACAGCAGCACAISI	3333
ACUUGUCAUUGGUGCUGUGAUCA 3308		stab11 antisense	AucAcAGcAccAAuGAcAATsT	3356
UUGAACCAGCUUGAGAGCAAAGU 3309		SARS:28806L21 siRNA (28788C) stab11 antisense	uuuGcucucAAGcuGGuucTsT	3357
ecunennuccucueecucunen 3310	İ	SARS:26549L21 siRNA (26531C)	44G4Gcc4G4GG4AAAcaateT	3350
╀		O A D C. 40 E 21 LO A C.	181888888888888888888888888888888888888	0000
+		SARS:1657U21 siRNA stab08 sense	AAuGAAGAGGuuGccAucATsT	3359
+		SARS:1166U21 siRNA stab08 sense	uuGcAucuccAcAGGAGuGTsT	3360
ACUUNACCGU		SARS:2383U21 siRNA stab08 sense	CAAAGCAAGGAcuuuAccTsT	3361
CUGUGUAAAUGGCCUCAUGCUCU 3306	_	SARS:2600U21 siRNA stab08 sense	GuGuAAAuGGccucAuGcuTsT	3362

SARS:26792U21 siRNA stab08 sense	3308 SARS:26792U21 siRNA stab0
SARS:28788U21 siRNA stab08 sense	3309 SARS:28788U21 siRNA stab0
SARS:26531U21 siRNA stab08 sense	3310 SARS:26531U21 siRNA stab08
SARS:1675L21 siRNA (1657C) stab08	SARS:1675L21 siRNA (1657C)
antisense	3303 antisense
SARS:1184L21 siRNA (1166C) stab08	SARS:1184L21 siRNA (1166C) s
antisense	3304 antisense
SARS:2401L21 siRNA (2383C) stab08	SARS:2401L21 siRNA (2383C)
antisense	3305 antisense
SARS:2618L21 siRNA (2600C) stab08	SARS:2618L21 siRNA (2600C)
antisense	3306 antisense
SARS:26592L21 siRNA (26574C)	SARS:26592L21 siRNA (2657
stab08 antisense	3307 stab08 antisense
SARS:26810L21 siRNA (26792C)	SARS:26810L21 siRNA (267
stab08 antisense	3308 stab08 antisense
SARS:28806L21 siRNA (28788C)	SARS:28806L21 siRNA (287
stab08 antisense	3309 stab08 antisense
SARS:26549L21 siRNA (26531C)	SARS:26549L21 siRNA (2653
stab08 antisense	GCUUGUUUCCUCUGGCUCUUGU 3310 stab08 antisense
 	3309 3309 3306 3306 3307 3308 3309 3309

Uppercase = ribonucleotide

u,c = 2'-deoxy-2'-fluoro U, C \underline{A} = 2'-O-methyl Adenosine \underline{G} = 2'-O-methyl Guanosine T = thymidine

B = inverted deoxy abasic

s = phosphorothioate linkage

A = deoxy Adenosine G = deoxy Guanosine

Table IV

Non-limiting examples of Stabilization Chemistries for chemically modified siNA constructs

Chemistry	pyrimidine	Purine	сар	p=S	Strand
"Stab 00"	Ribo	Ribo	TT at 3'-ends		S/AS
"Stab 1"	Ribo	Ribo	-	5 at 5'-end 1 at 3'-end	S/AS
"Stab 2"	Ribo	Ribo	-	All linkages	Usually AS
"Stab 3"	2'-fluoro	Ribo	-	4 at 5'-end 4 at 3'-end	Usually S
"Stab 4"	2'-fluoro	Ribo	5' and 3'-ends	-	Usually S
"Stab 5"	2'-fluoro	Ribo	-	1 at 3'-end	Usually AS
"Stab 6"	2'-O-Methyl	Ribo	5' and 3'-ends	-	Usually S
"Stab 7"	2'-fluoro	2'-deoxy	5' and 3'-ends	-	Usually S
"Stab 8"	2'-fluoro	2'-O- Methyl	-	1 at 3'-end	Usually AS
"Stab 9"	Ribo	Ribo	5' and 3'-ends	-	Usually S
"Stab 10"	Ribo	Ribo	_	1 at 3'-end	Usually AS
"Stab 11"	2'-fluoro	2'-deoxy	-	1 at 3'-end	Usually AS
"Stab 12"	2'-fluoro	LNA	5' and 3'-ends		Usually S
"Stab 13"	2'-fluoro	LNA		1 at 3'-end	Usually AS
"Stab 14"	2'-fluoro	2'-deoxy		2 at 5'-end 1 at 3'-end	Usually AS
"Stab 15"	2'-deoxy	2'-deoxy		2 at 5'-end 1 at 3'-end	Usually AS
"Stab 16	Ribo	2'-O- Methyl	5' and 3'-ends		Usually S
"Stab 17"	2'-O-Methyl	2'-O- Methyl	5' and 3'-ends		Usually S
"Stab 18"	2'-fluoro	2'-O- Methyl	5' and 3'-ends	1 at 3'-end	Usually S
"Stab 19"	2'-fluoro	2'-O- Methyl	3'-end		Usually AS
"Stab 20"	2'-fluoro	2'-deoxy	3'-end		Usually AS
"Stab 21"	2'-fluoro	Ribo	3'-end		Usually AS
"Stab 22"	Ribo	Ribo	3'-end -		Usually AS

CAP = any terminal cap, see for example Figure 10.

All Stab 1-22 chemistries can comprise 3'-terminal thymidine (TT) residues

All Stab 1-22 chemistries typically comprise about 21 nucleotides, but can vary as described herein.

S = sense strand AS = antisense strand

Table V

A. 2.5 µmol Synthesis Cycle ABI 394 Instrument

Reagent	Equivalents	Amount	Wait Time* DNA	Wait Time* 2'-O-methyl	Wait Time*RNA
Phosphoramidites	6.5	163 µL	45 sec	2.5 min	7.5 min
S-Ethyl Tetrazole	23.8	238 µL	45 sec	2.5 min	7.5 min
Acetic Anhydride	100	233 μL	5 sec	5 sec	5 sec
N-Methyl Imidazole	186	233 µL	5 sec	5 sec	5 sec
TCA	176	2.3 mL	21 sec	21 sec	21 sec
Iodine	11.2	1.7 mL	45 sec	45 sec	45 sec
Beaucage	12.9	645 μL	100 sec	300 sec	300 sec
Acetonitrile	NA	6.67 mL	NA	NA	NA

B. 0.2 µmol Synthesis Cycle ABI 394 Instrument

Reagent	Equivalents	Amount	Wait Time* DNA	Wait Time* 2'-O-methyl	Wait Time*RNA
Phosphoramidites	15	31 µL	45 sec	233 sec	465 sec
S-Ethyl Tetrazole	38.7	31 µL	45 sec	233 min	465 sec
Acetic Anhydride	655	124 µL	5 sec	5 sec	5 sec
N-Methyl Imidazole	1245	124 µL	5 sec	5 sec	5 sec
TCA	700	732 µL	10 sec	10 sec	10 sec
lodine	20.6	244 µL	15 sec	15 sec	15 sec
Beaucage	7.7	232 µL	100 sec	300 sec	300 sec
Acetonitrile	NA	2.64 mL	NA	NA	NA

C. $0.2\,\mu mol\,Synthesis\,Cycle\,96$ well Instrument

Reagent	Equivalents:DNA/ 2'-O-methyl/Ribo	Amount: DNA/2'-O- methyl/Ribo	Wait Time* DNA	Wait Time* 2'-O- methyl	Wait Time* Ribo
Phosphoramidites	22/33/66	40/60/120 μL	60 sec	180 sec	360sec
S-Ethyl Tetrazole	70/105/210	40/60/120 μL	60 sec	180 min	360 sec
Acetic Anhydride	265/265/265	50/50/50 µL	10 sec	10 sec	10 sec
N-Methyl Imidazole	502/502/502	50/50/50 μL	10 sec	10 sec	10 sec
TCA	238/475/475	250/500/500 µL	15 sec	15 sec	15 sec
lodine	6.8/6.8/6.8	80/80/80 µL	30 sec	30 sec	30 sec
Beaucage	34/51/51	80/120/120	100 sec	200 sec	200 sec
Acetonitrile	NA	1150/1150/1150 µL	NA	NA	NA

- Wait time does not include contact time during delivery.
 - Tandem synthesis utilizes double coupling of linker molecule

CLAIMS

What we claim is:

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1. A chemically synthesized double stranded short interfering nucleic acid (siNA) molecule that directs cleavage of a severe acute respiratory syndrome (SARS) virus RNA via RNA interference, wherein:

- a. each strand of said siNA molecule is about 19 to about 23 nucleotides in length;
- b. one strand of said siNA molecule comprises nucleotide sequence having sufficient complementarity to said SARS virus RNA for the siNA molecule to direct cleavage of the SARS virus RNA via RNA interference; and
- c. said siNA molecule does not require the presence of nucleotides having a 2'-hydroxy group for mediating RNA interference.
- 2. The siNA molecule of claim 1, wherein said siNA molecule comprises no ribonucleotides.
- 15 3. The siNA molecule of claim 1, wherein said siNA molecule comprises ribonucleotides.
 - 4. The siNA molecule of claim 1, wherein one strand of said double-stranded siNA molecule comprises a nucleotide sequence that is complementary to a nucleotide sequence of a SARS virus gene or a portion thereof, and wherein a second strand of said double-stranded siNA molecule comprises a nucleotide sequence substantially similar to the nucleotide sequence or a portion thereof of said SARS virus RNA.
 - 5. The siNA molecule of claim 4, wherein each strand of the siNA molecule comprises about 19 to about 23 nucleotides, and wherein each strand comprises at least about 19 nucleotides that are complementary to the nucleotides of the other strand.
- The siNA molecule of claim 1, wherein said siNA molecule comprises an antisense region comprising a nucleotide sequence that is complementary to a nucleotide sequence of a SARS virus gene or a portion thereof, and wherein said siNA further comprises a sense region, wherein said sense region comprises a nucleotide sequence substantially similar to the nucleotide sequence of said SARS virus gene or a portion thereof.

7. The siNA molecule of claim 6, wherein said antisense region and said sense region comprises about 19 to about 23 nucleotides, and wherein said antisense region comprises at least about 19 nucleotides that are complementary to nucleotides of the sense region.

- The siNA molecule of claim 1, wherein said siNA molecule comprises a sense region and an antisense region, and wherein said antisense region comprises a nucleotide sequence that is complementary to a nucleotide sequence of RNA encoded by a SARS virus gene, or a portion thereof, and said sense region comprises a nucleotide sequence that is complementary to said antisense region.
- 10 9. The siNA molecule of claim 6, wherein said siNA molecule is assembled from two separate oligonucleotide fragments wherein one fragment comprises the sense region and a second fragment comprises the antisense region of said siNA molecule.
 - 10. The siNA molecule of claim claim 6, wherein said sense region is connected to the antisense region via a linker molecule.
- 15 11. The siNA molecule of claim 10, wherein said linker molecule is a polynucleotide linker.
 - 12. The siNA molecule of claim 10, wherein said linker molecule is a non-nucleotide linker.
- 13. The siNA molecule of claim 6, wherein pyrimidine nucleotides in the sense region are 2'-O-methyl pyrimidine nucleotides.
 - 14. The siNA molecule of claim 6, wherein purine nucleotides in the sense region are 2'-deoxy purine nucleotides.
 - 15. The siNA molecule of claim 6, wherein pyrimidine nucleotides present in the sense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides.
- 25 16. The siNA molecule of claim 9, wherein the fragment comprising said sense region includes a terminal cap moiety at the 5'-end, the 3'-end, or both of the 5' and 3' ends of the fragment comprising said sense region.
 - 17. The siNA molecule of claim 16, wherein said terminal cap moiety is an inverted deoxy abasic moiety.
- 30 18. The siNA molecule of claim 6, wherein pyrimidine nucleotides of said antisense region are 2'-deoxy-2'-fluoro pyrimidine nucleotides

19. The siNA molecule of claim 6, wherein purine nucleotides of said antisense region are 2'-O-methyl purine nucleotides.

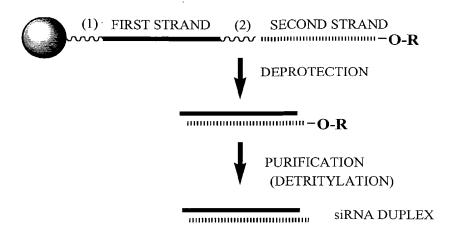
- 20. The siNA molecule of claim 6, wherein purine nucleotides present in said antisense region comprise 2'-deoxy- purine nucleotides.
- 5 21. The siNA molecule of claim 18, wherein said antisense region comprises a phosphorothioate internucleotide linkage at the 3' end of said antisense region.
 - 22. The siNA molecule of claim 6, wherein said antisense region comprises a glyceryl modification at the 3' end of said antisense region.
- 23. The siNA molecule of claim 9, wherein each of the two fragments of said siNA molecule comprise 21 nucleotides.

15

- 24. The siNA molecule of claim 23, wherein about 19 nucleotides of each fragment of the siNA molecule are base-paired to the complementary nucleotides of the other fragment of the siNA molecule and wherein at least two 3' terminal nucleotides of each fragment of the siNA molecule are not base-paired to the nucleotides of the other fragment of the siNA molecule.
- 25. The siNA molecule of claim 24, wherein each of the two 3' terminal nucleotides of each fragment of the siNA molecule are 2'-deoxy-pyrimidines.
- 26. The siNA molecule of claim 25, wherein said 2'-deoxy-pyrimidine is 2'-deoxy-thymidine.
- 20 27. The siNA molecule of claim 23, wherein all 21 nucleotides of each fragment of the siNA molecule are base-paired to the complementary nucleotides of the other fragment of the siNA molecule.
- 28. The siNA molecule of claim 23, wherein about 19 nucleotides of the antisense region are base-paired to the nucleotide sequence of the RNA encoded by a SARS virus gene or a portion thereof.
 - 29. The siNA molecule of claim 23, wherein 21 nucleotides of the antisense region are base-paired to the nucleotide sequence of the RNA encoded by a SARS virus gene or a portion thereof.
- 30. The siNA molecule of claim 9, wherein the 5'-end of the fragment comprising said antisense region optionally includes a phosphate group.

31. A pharmaceutical composition comprising the siNA molecule of claim 1 in an acceptable carrier or diluent.

Figure 1



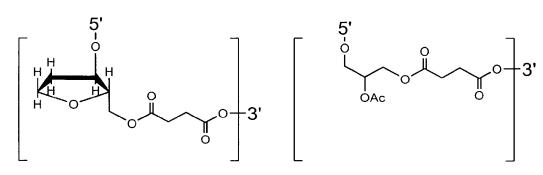
= SOLID SUPPORT

R = TERMINAL PROTECTING GROUP FOR EXAMPLE: DIMETHOXYTRITYL (DMT)

(1) = CLEAVABLE LINKER
(FOR EXAMPLE: NUCLEOTIDE SUCCINATE OR
INVERTED DEOXYABASIC SUCCINATE)

= CLEAVABLE LINKER

(FOR EXAMPLE: NUCLEOTIDE SUCCINATE OR INVERTED DEOXYABASIC SUCCINATE)



INVERTED DEOXYABASIC SUCCINATE LINKAGE

GLYCERYL SUCCINATE LINKAGE

Figure 2

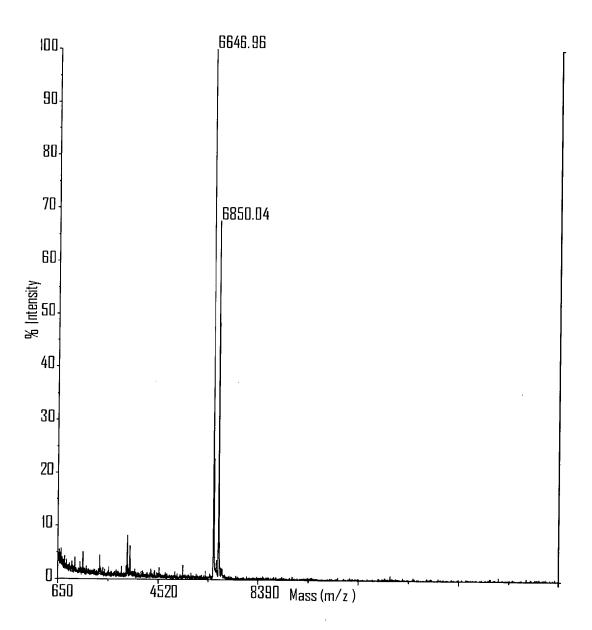


Figure 3

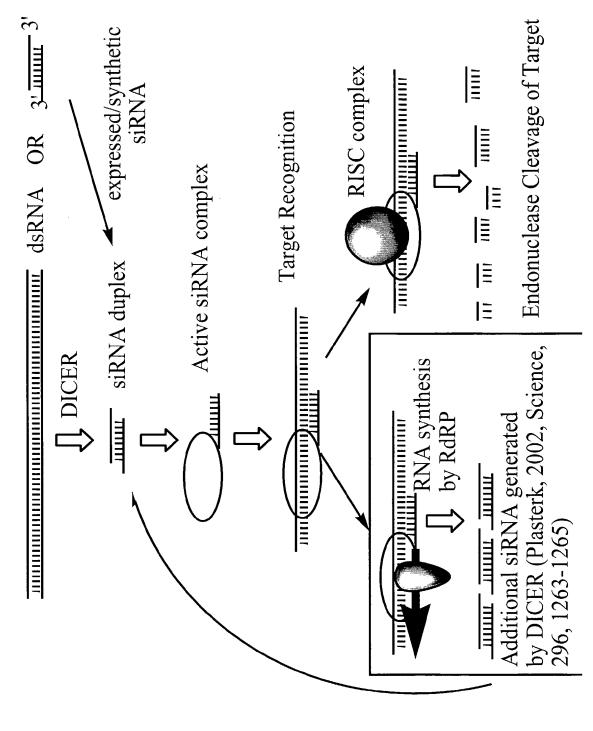


Figure 4

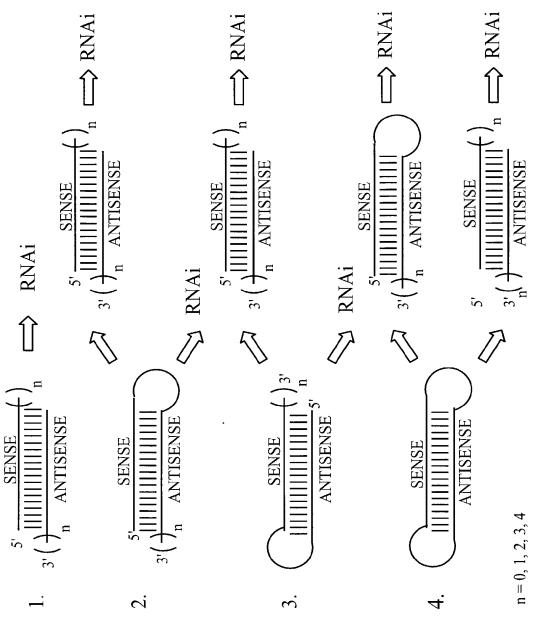
```
SENSE STRAND (SEQ ID NO 3375)
                 ALL POSITIONS RIBONUCLEOTIDE EXCEPT PÓSITIONS (N N)
       5'-
                 -3'
       3'-
           ..L-(N<sub>c</sub>N) NNNNNNNNNNNNNNNNN
                                                              -5'
                          ANTISENSE STRAND (SEQ ID NO 3376)
                  ALL POSITIONS RIBONUCLEOTIDE EXCEPT POSITIONS (N N)
                        SENSE STRAND (SEQ ID NO 3377)
        ALL PYRIMIDINES = 2'-FLUORO AND ALL PURINES = 2'-OM EXCEPT POSITIONS (N N)
                 -3'
B
       3'-
            L-(N<sub>s</sub>N) NNNNNNNNNNNNNNNNNNN
                                                             -5'
                       ANTISENSE STRAND (SEQ ID NO 3378)
       ALL PYRIMIDINES = 2'-FLUORO AND ALL PURINES = 2'-O-ME EXCEPT POSITIONS (N N)
                          SENSE STRAND (SEQ ID NO 3379)
              ALL PYRIMIDINES = 2'-O-ME OR 2'-FLUORO EXCEPT POSITIONS (N N)
       5'-
                -31
       3'-
             -5
                          ANTISENSE STRAND (SEQ ID NO 3380)
                    ALL PYRIMIDINES = 2'-FLUORO EXCEPT POSITIONS (N N)
                        SENSE STRAND (SEQ ID NO 3381)
      ALL PYRIMIDINES = 2'-FLUORO EXCEPT POSITIONS (N N) AND ALL PURINES = 2'-DEOXY
                -3'
D
           L-(N<sub>s</sub>N) NNNNNNNNNNNNNNNNNNN
      3'-
                                                             -5'
                       ANTISENSE STRAND (SEQ ID NO 3378)
       ALL PYRIMIDINES = 2'-FLUORO AND ALL PURINES = 2'-O-ME EXCEPT POSITIONS (N N)
                          SENSE STRAND (SEQ ID NO 3382)
                  ALL PYRIMIDINES = 2'-FLUORO EXCEPT POSITIONS (N N)
                -3'
\mathbf{E}
          L-(NN) NNNNNNNNNNNNNNNNNNNN
                                                             -5'
                       ANTISENSE STRAND (SEQ ID NO 3378)
       ALL PYRIMIDINES = 2'-FLUORO AND ALL PURINES = 2'-O-ME ÉXCEPT POSITIONS (N N)
                        SENSE STRAND (SEQ ID NO 3381)
      ALL PYRIMIDINES = 2'-FLUORO EXCEPT POSITIONS (N N) AND ALL PURINES = 2'-DEOXY
               -3'
H
            3'-
                                                             -5'
                      ANTISENSE STRAND (SEO ID NO 3383)
      ALL PYRIMIDINES = 2'-FLUORO EXCEPT POSITIONS (N N) AND ALL PURINES = 2'-DEOXY
```

POSITIONS (NN) CAN COMPRISE ANY NUCLEOTIDE, SUCH AS DEOXYNUCLEOTIDES (eg. THYMIDINE) OR UNIVERSAL BASES B = ABASIC, INVERTED ABASIC, INVERTED NUCLEOTIDE OR OTHER TERMINAL CAP THAT IS OPTIONALLY PRESENTL = GLYCERYL or B THAT IS OPTIONALLY PRESENTS = PHOSPHOROTHIOATE OR PHOSPHORODITHIOATE that is optionally absent

Figure 5

```
SENSE STRAND (SEQ ID NO 3384)
                                                                                   -3'
                      iB-AUGUGCUUGAAGAUCCUUGTT-iB
          3'-
                 L-T<sub>S</sub>TUACACGAACUUCUAGGAAC
                                                                                   -5'
                                ANTISENSE STRAND (SEQ ID NO 3385)
                                  SENSE STRAND (SEQ ID NO 3386)
          5'-
                         <u>augugcuugaagauccuugTT</u>
                                                                                    -3'
B
                                                                                    -5'
          3'-
                  L-T<sub>S</sub>Tu<u>a</u>c<u>a</u>c<u>a</u>c<u>g</u>a<u>a</u>cuucu<u>a</u>gg<u>a</u>ac
                                 ANTISENSE STRAND (SEQ ID NO 3387)
                                  SENSE STRAND (SEQ ID NO 3388)
          5'-
                       iB-AuGuGcuuGAAGAuccuuGTT-iB
                                                                                    -31
          3'-
                  L-T<sub>S</sub>T u A c A c G A A c u u c u A G G A A c
                                                                                    -5'
                                 ANTISENSE STRAND (SEQ ID NO 3389)
                                 SENSE STRAND (SEQ ID NO 3390)
         5'-
                      iB-AuGuGcuuGAAGAuccuuGTT-iB
                                                                                   -31
D
         3'-
                   L-T<sub>S</sub>T u <u>a c a c g a a c u u c u a g g a a c</u>
                                                                                   -5'
                                ANTISENSE STRAND (SEQ ID NO 3387)
                                 SENSE STRAND (SEQ ID NO 3391)
         5'-
                        iB-A u G u G c u u G A A G A u c c u u G T T-iB
                                                                                   -3'
\mathbf{E}
         3'-
                     L-T<sub>S</sub>Tu<u>a</u>c<u>a</u>c<u>a</u>a<u>c</u>g<u>a</u>acuucu<u>a</u>gg<u>a</u>ac
                                                                                   -5'
                                ANTISENSE STRAND (SEQ ID NO 3387)
                                 SENSE STRAND (SEQ ID NO 3390)
          5'-
                         iB-AuGuGcuuGAAGAuccuuGTT-iB
                                                                                   -3'
F
          3'-
                    L-T<sub>S</sub>T u A c A c G A A c u u c u A G G A A c
                                                                                   -5'
                                ANTISENSE STRAND (SEQ ID NO 3392)
         lower case = 2'-O-Methyl or 2'-deoxy-2'-fluoro
                                                      ITALIC UPPER CASE = DEOXYIB = INVERTED DEOXYABASICL = GLYCERYL MOIETY or iB OPTIONALLY PRESENTS = PHOSPHOROTHIOATE (
         italic lower case = 2'-deoxy-2'-fluoro
         underline = 2'-O-methyl
                                                         PHOSPHORODITHIOATE OPTIONALLY PRESEN
```

Figure 6



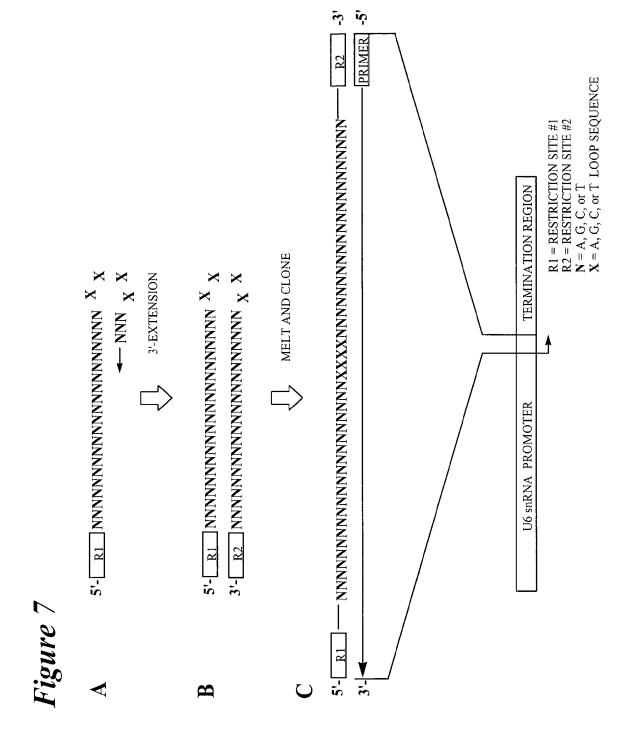


Figure 8

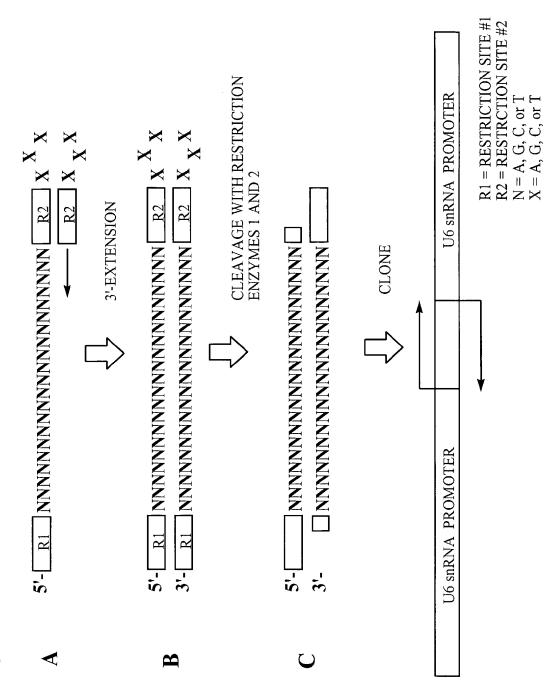
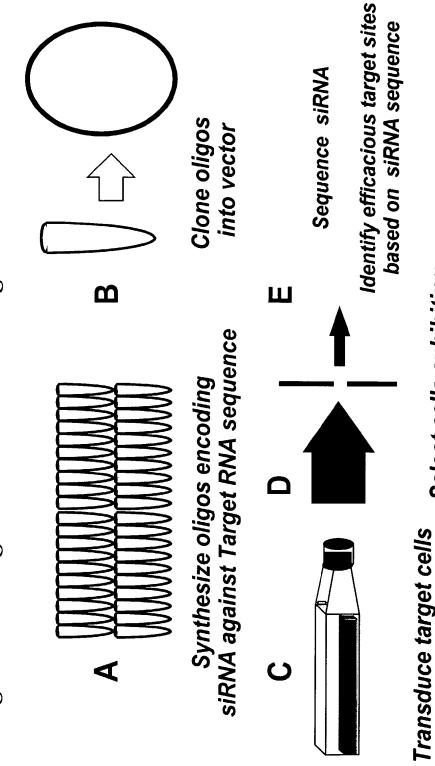


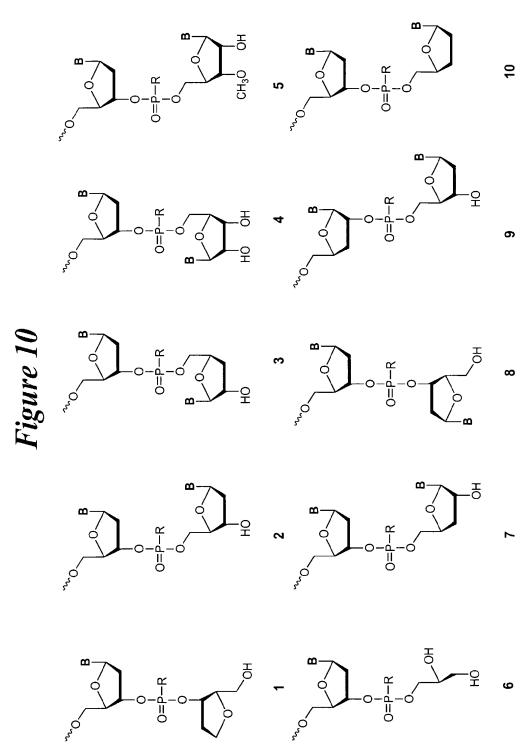
Figure 9: Target site Selection using siRNA



Select cells exhibiting

desired phenotype

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R = O, S, N, alkyl, substituted alkyl, O-alkyl, S-alkyl, alkaryl, or aralkyl B = Independently any nucleotide base, either naturally occurring or chemically modified, or optionally H (abasic).

Figure 11: Modification Strategy

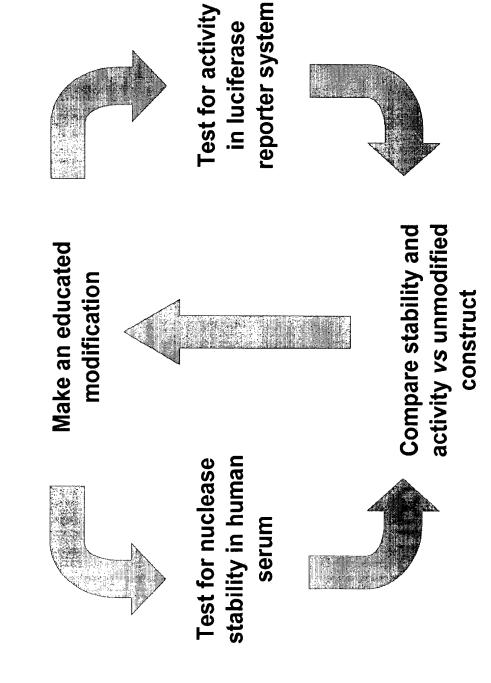
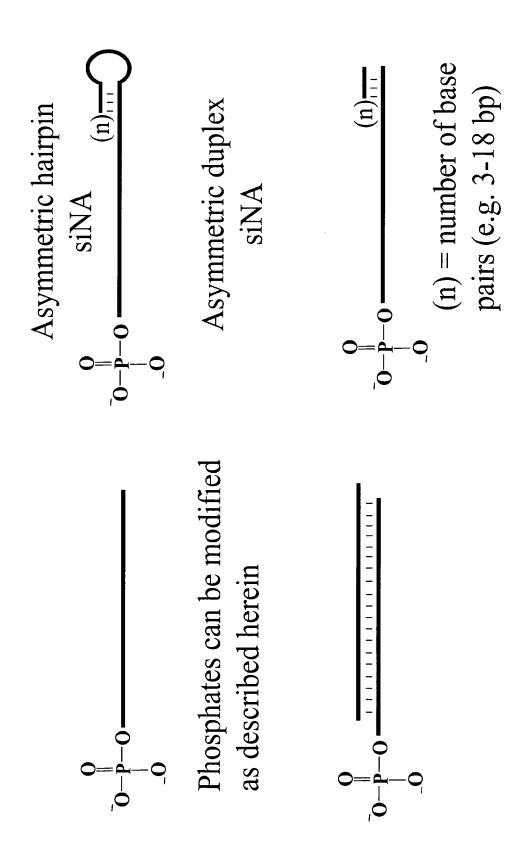


Figure 12: Phosphorylated siNA constructs



modifications herein

Figure 13: 5'-phosphate modifications

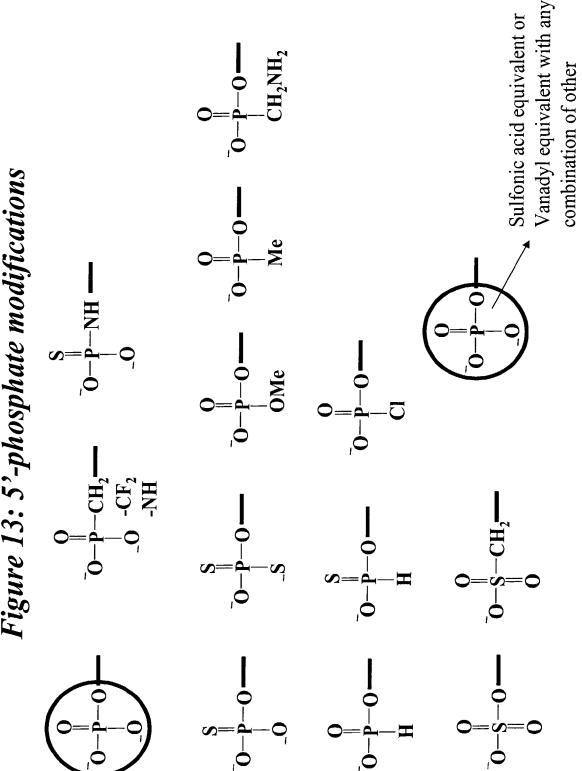


Figure 14A: Duplex forming oligonucleotide constructs that utilize palindrome

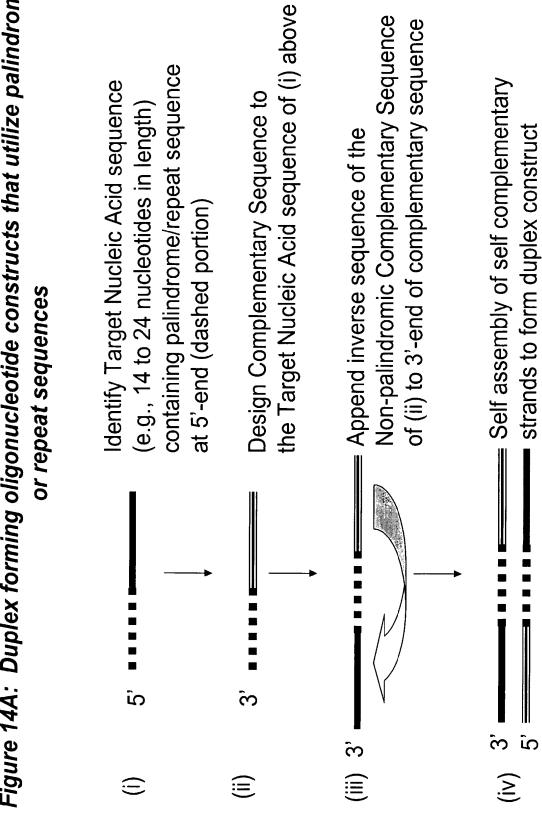


Figure 14B: Example of a duplex forming oligonucleotide sequence that utilizes a palindrome or repeat sequence

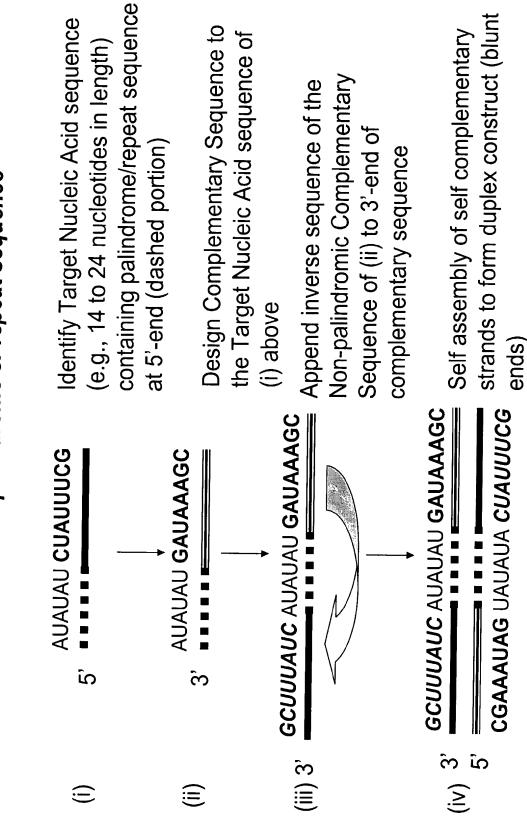


Figure 14C: Example of a duplex forming oligonucleotide sequence that utilizes a palindrome or repeat sequence, self assembly

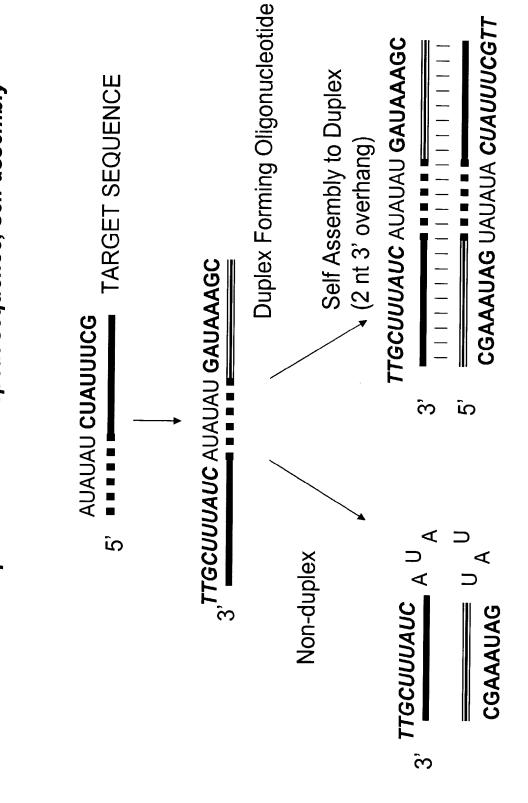


Figure 14D: Example of a duplex forming oligonucleotide sequence that utilizes a palindrome or repeat sequence, self assembly and inhibition

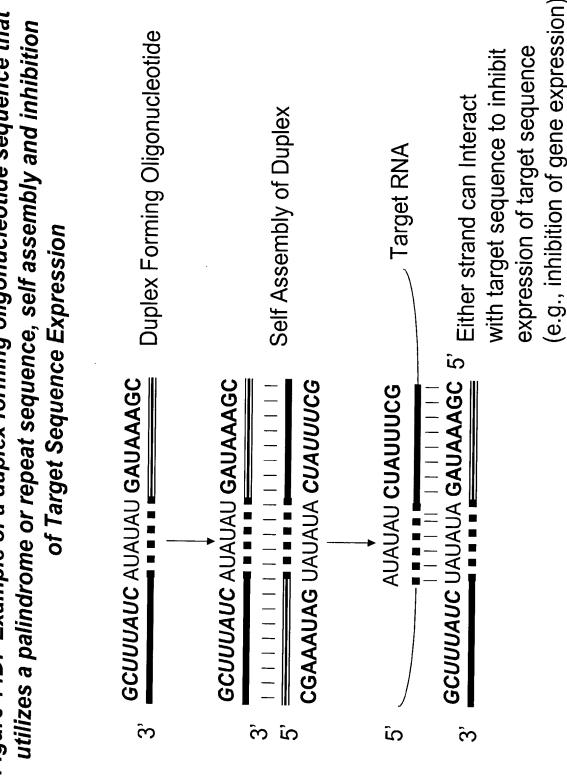


Figure 15: Duplex forming oligonucleotide constructs that utilize artificial palindrome or repeat sequences

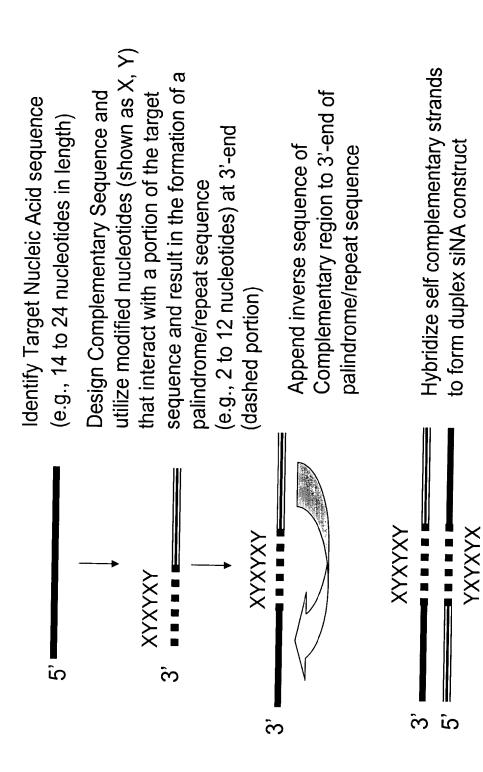


Figure 16: Examples of double stranded multifunctional siNA constructs with

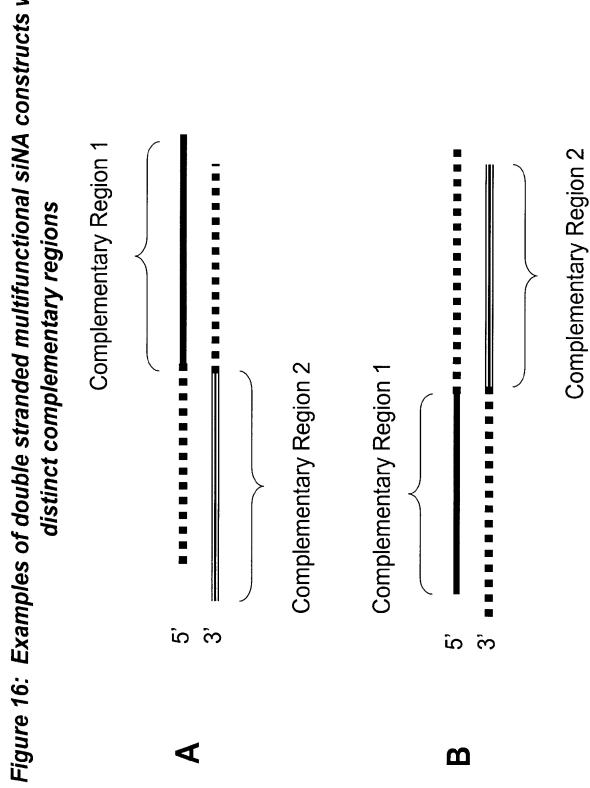
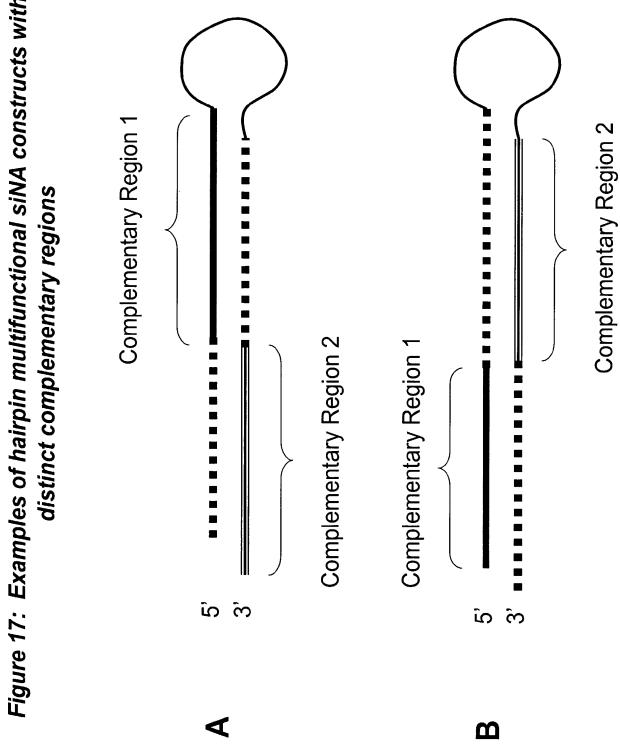
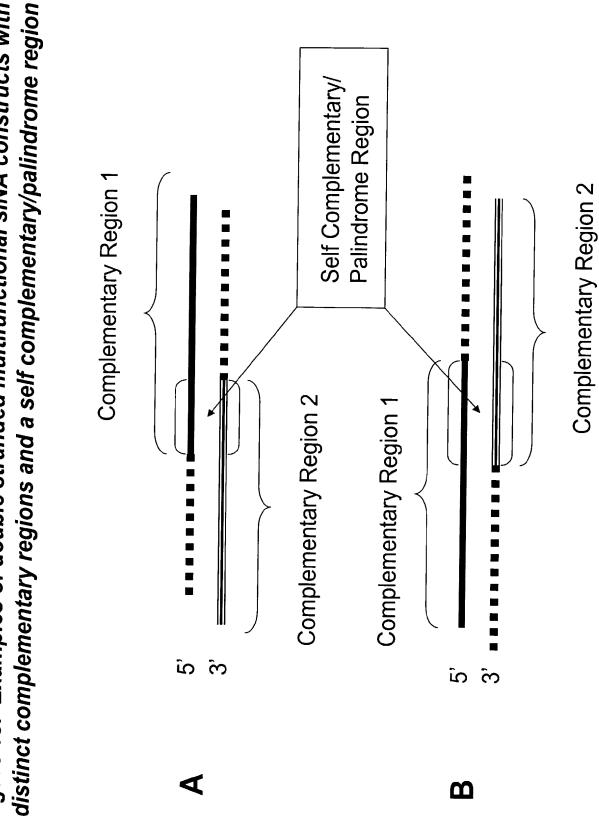


Figure 17: Examples of hairpin multifunctional siNA constructs with



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Figure 18: Examples of double stranded multifunctional siNA constructs with



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Figure 19: Examples of hairpin multifunctional siNA constructs with

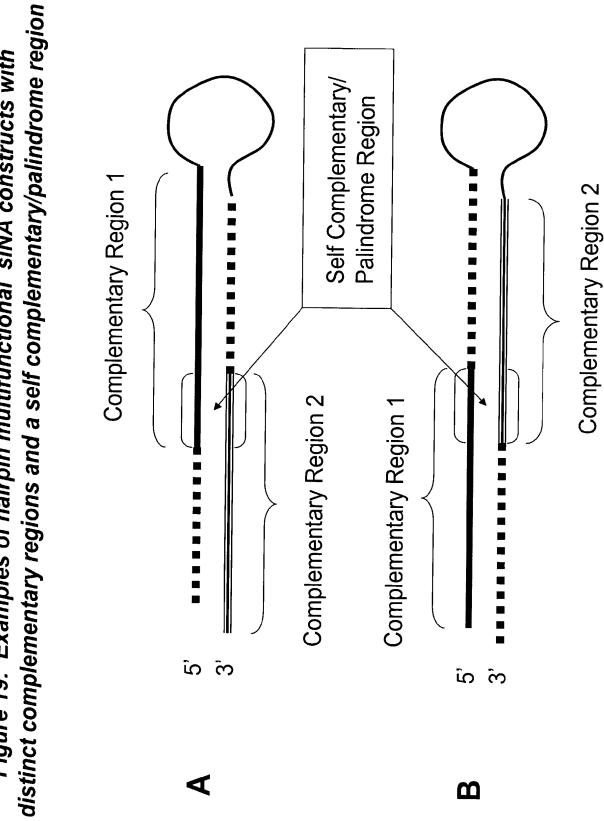
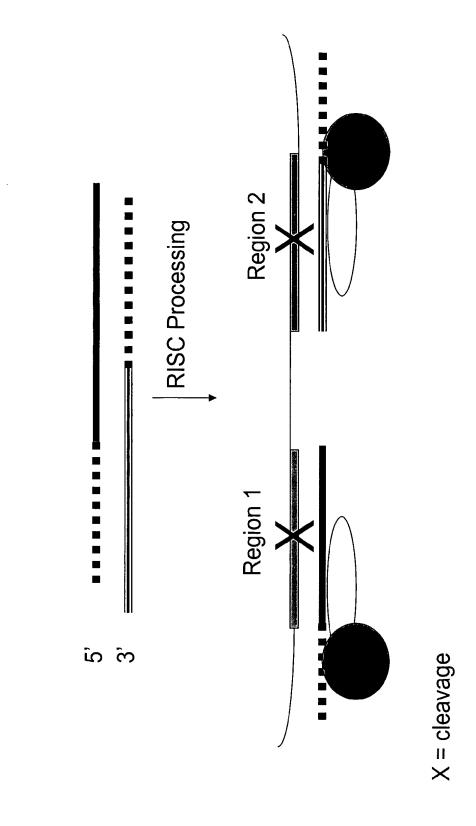


Figure 20: Example of multifunctional siNA targeting two separate Target 1 RNA Target 2 RNA **RISC Processing** Target nucleic acid sequences OR က်က် X = cleavage

Figure 21: Example of multifunctional siNA targeting two regions within the same target nucleic acid sequence



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